University of Reading

Antimicrobial & antiparasitic use and resistance in British sheep and cattle: a systematic review

Article

Accepted Version

Creative Commons: Attribution-Noncommercial-No Derivative Works 4.0

Hennessey, M., Whatford, L., Payne-Gifford, S., Johnson, K. F. ORCID: https://orcid.org/0000-0002-5088-1163, Van Winden, S., Barling, D. and Häsler, B. (2020) Antimicrobial & antiparasitic use and resistance in British sheep and cattle: a systematic review. Preventive Veterinary Medicine, 185. 105174. ISSN 0167-5877 doi: 10.1016/j.prevetmed.2020.105174 Available at https://centaur.reading.ac.uk/95051/

It is advisable to refer to the publisher's version if you intend to cite from the work. See Guidance on citing.
Published version at: http://dx.doi.org/10.1016/j.prevetmed.2020.105174
To link to this article DOI: http://dx.doi.org/10.1016/j.prevetmed.2020.105174

Publisher: Elsevier

All outputs in CentAUR are protected by Intellectual Property Rights law, including copyright law. Copyright and IPR is retained by the creators or other copyright holders. Terms and conditions for use of this material are defined in the End User Agreement.

University of Reading

www.reading.ac.uk/centaur

CentAUR

Central Archive at the University of Reading

Reading's research outputs online

Antimicrobial & antiparasitic use and resistance in British sheep and cattle production systems: a systematic review

Mathew P. Hennessey*[1], Louise Whatford[1], Sophie Payne-Gifford[2], Kate Johnson[2], Steven Van Winden[1], David Barling[2], Barbara Häsler[1]

*Corresponding author; mphennessey@rvc.ac.uk

[1] Veterinary Epidemiology, Economics and Public Health Group, Department of Pathobiology and Population Sciences, Royal Veterinary College, London, UK

[2] Centre for Agriculture Food and Environmental Management, School of Life Science, University of Hertfordshire

Abstract

A variety of antimicrobials and antiparasitics are used to treat British cattle and sheep to ensure animal welfare, a safe food supply, and maintain farm incomes. However, with increasing global concern about antimicrobial resistance in human and animal populations, there is increased scrutiny of the use of antimicrobials in food-producing animals.

This systematic review sought to identify and describe peer and non-peer reviewed sources, published over the last ten years, detailing the usage of, and resistance to, antimicrobials and antiparasitics in sheep and cattle farming systems in Britain as well as identify knowledge gaps. Applying the PRISMA review protocol and guidelines for including grey literature; Scopus, Web of Science, Medline, and government repositories were searched for relevant articles and reports. Seven hundred and seventy titles and abstracts and 126 full-text records were assessed, of which 40 scholarly articles and five government reports were included for data extraction.

Antibiotic usage in sheep and cattle in Britain appear to be below the UK average for all livestock and tetracyclines and beta-lactam antibiotics were found to be the most commonly used. However, the poor level of coverage afforded to these species compared to other livestock reduced the certainty of these findings. Although resistance to some

antibiotics (using *Escherichia coli* as a marker) appears to have decreased in sheep and cattle in England and Wales over the last few years, levels of resistance remain high to the commonly used antibiotics. The small number and fragmented nature of studies identified by this review describing anthelmintic usage, and the lack of available national sales data, prevented the identification of trends in either sheep or cattle.

We recommend that additional efforts are taken to collect farm or veterinary level data on antimicrobial usage and resistance, especially in sheep, which appear from this review to be a neglected species in this field. Additionally, metrics produced by this data should be generated in a way to allow for maximum comparability across species, sectors, and countries.

Introduction

The use of antimicrobial and antiparasitic agents allow the control of pathogens in order to increase animal health, welfare, and productivity in livestock settings which are challenged by disease (Page and Gautier 2012). However, the increased use of these agents over the last 70 years has led to the development of resistance to treatment with subsequent negative health and economic effects (Heymann 2006). Antimicrobial resistance is recognised as a global health threat, and is predicted to develop into a leading cause of human fatality by 2050, with an annual cost to the global economy of 100 trillion US dollars (O'Neill 2016). Anthelmintic resistance while primarily species specific, is a major cause of poor productivity and economic loss in livestock production systems globally (Shalaby 2013).

While the interactions between human, animal, and environmental microbiomes are complex and not fully understood, evidence exists linking the use of antibiotics in one microbiome to the prevalence of resistant organisms in another; occupational exposure to livestock has been reported as a risk for human health, particularly among veterinarians, farmers, livestock cullers, and slaughterhouse workers, who are exposed to organisms such as livestock associated methicillin resistant *Staphylococcus aureus* (MRSA) and *Coxiella burnetii* (Klous *et al.* 2016; Rossi *et al.* 2017; Tang *et al.* 2017). While reducing the use of antimicrobials in one population is known to be correlated with a reduction in resistance in the same population, evidence linking reductions of use in livestock with reductions of

resistant organisms in humans is currently scarce (Dorado-García *et al.* 2016; Tang *et al.* 2017; Træholt Franck *et al.* 2017; Veldman *et al.* 2017; Bennani *et al.* 2020). Thus, while measures to reduce antimicrobial usage in farming provide safeguarding mechanisms to protect their therapeutic use in livestock, delineating the benefit such measures have to protect the therapeutic use of antimicrobials in humans remains challenging.

Although there are calls to govern the use of antimicrobials at an international level (Woolhouse *et al.* 2015; Padiyara *et al.* 2018), with guidance documents and action plans from global bodies such as the World Health Organisation (WHO), Food and Agriculture Organisation (FAO), and the World Organisation for Animal Health (OIE), (FAO 2016; OIE 2016; WHO 2019), there is no legally binding international treaty (no Montreal or Kyoto protocol) on how they should be used or documented (Heymann and Ross 2019). At a national level, there are various best practice guidelines available to antimicrobial and antiparasitic users in livestock in Britain, such as the UK government's One Health report on antibiotic use and resistance (VMD 2019a) and five-year action plan for antimicrobial resistance (DHSC 2019), the British Veterinary Association's policy statement on the responsible use of antimicrobials in food producing animals (BVA 2019), and the industry led initiatives Sustainable Control of Parasites in Sheep (SCOPS 2019) and Control of Worms Sustainably (COWS 2019a). To date, the use of antimicrobials in livestock in Britain is governed by EU (indirectly) and national legislation, which include the 2006 ban on antibiotics being used as growth promoters and a 2018 proposal to restrict the routine use of prophylactic and metaphylactic antibiotics (due to come into effect in 2022) (European Parliament 2019). Although possible to repeal EU legislation post-Brexit, it is likely the UK will adopt this legislation after its exit as the UK has been one of the forerunners of effective voluntary strategies to reduce antimicrobial use driven by strong private-public partnerships and private industry involvement and leadership.

In Britain, the Veterinary Medicines Directorate (VMD; an agency of the Department of Environment Farming and Rural Affairs) regulates medicine registration and use. The National Office of Animal Health (NOAH) and the Responsible Use of Medicines in Agriculture Alliance (RUMA), two industry initiatives, set the background of what antimicrobials are available and how they are used in livestock. And yet, apart from pigs and poultry, the level of use of antimicrobials in British livestock production is relatively

unknown at farm level. Often, due to multi-species registration of medicines, amounts of antimicrobials are stated at livestock level and not species or farm level. Although farmers are legally required to record the amount of antimicrobials they have used (DEFRA 2019), this data is used for individual farm management and farm assurance schemes, and not stored in a central database and therefore not readily available for antimicrobial usage surveillance.

Usage of antibiotics is calculated through national sales data by the VMD, and while this inferred usage has good coverage for some livestock species (for example usage in salmon farming is 100% complete), there is only 30% coverage for dairy cattle, 5.5% coverage for beef cattle, and no known sales data coverage for sheep (VMD 2018). Additionally, as antimicrobials are often registered to multiple livestock species, sales cannot be reliably related to a certain species, unless the drug of use is solely registered to said species (for example products solely licensed to fish). The VMD collects antibiotic sales data and usage data. Antibiotic sales data are submitted by pharmaceutical companies to the VMD on their previous year's sales of antimicrobials authorised for use in animals in accordance with veterinary medicine regulations 2013. Antibiotic usage data are collected and submitted voluntarily by different livestock stakeholders to the VMD. This was the result of a collaboration between RUMA and the VMD and first published in 2014 with only usage data from the poultry sector until more data became available in the subsequent years. Additionally, although the UK participates in mandatory EU-wide antibiotic resistance monitoring, in 2018 samples were only taken from poultry (VMD 2018), and so understanding the links between antimicrobial usage and resistance at the animal and farm level is challenging.

Cattle and sheep are the two most commonly produced red meat species in Britain and understanding the level of usage and resistance of/to anti-infective agents is an important aspect of the national agenda for controlling antimicrobial resistance and ensuring the sustainability of domestic meat production, especially given the changing horizon ahead by leaving the governance of the EU behind. Consequently, the aim of this study was to conduct a systematic review on the use and resistance of antimicrobials and antiparasitics in cattle and sheep production systems in Britain to provide an overview of the current situation and identify gaps in knowledge.

Methods

Search strategy

A systematic literature review was conducted in line with PRISMA guidelines (Moher *et al.* 2015). First, an *a priori* protocol was produced which set out the primary and secondary objectives and the review question; namely to (1) identify and describe the existing literature detailing the level of usage and resistance to antimicrobials and antiparasitics in British[1] sheep and cattle production systems, and (2) identify any research gaps within this topic. Inclusion criteria were defined based on the *population*, *intervention*, *comparison*, *outcomes* of an article, and *study* design framework (PICOS, adapted from Chatterjee *et al*., (2018)) and included; English language, peer-reviewed texts and reports, which had a focus on sheep and/or cattle raised for meat production in Britain (England, Wales, and Scotland) published in the last ten years; further details are given in Supp. 1 (section 6). The search was conducted on the 11th and 12th June 2019 in Scopus, Web of Science and Medline databases. These three databases were selected to provide a high level of article recall across biomedical articles (Bramer *et al.* 2017).

Search terms were derived using the Boolean operator OR for the following four themes, (1) anti-infective agent, (2) livestock population[2], (3) location, and (4) focus, before being combined using the Boolean operators 'AND' and 'AND NOT' (Table 1). The term 'UK or United Kingdom' was included at this stage to screen for any articles which may contain information on England, Scotland, or Wales.

Table 1. Search terms used to build the systematic review

Anti-infective agent	**Livestock population**	**Location**	**Focus**	**Exclude**
(antimicrobial* OR "anti microbial*" OR antibiotic* OR "anti biotic*" OR	AND (livestock OR cattle OR beef OR cow OR cows OR calf OR calv* OR	AND (GB OR "Great Britain" OR England OR English OR wales OR welsh	AND (use OR using OR usage OR resis* OR	AND NOT "New south wales"

[1] British (English, Scottish, and Welsh) production systems were the focus of this review (rather than the whole of the United Kingdom)

[2] As around half of British beef is supplied from the dairy sector (through calves and cull cows) (AHDB 2017) the use of antibiotics in dairy cows was considered a relevant indicator of antibiotic use in red meat production.

antifungal* OR "anti fungal*" OR antiprotozoal* OR "anti protozoal*" OR bactericid* OR bacteriostat* OR anti-infective* OR "anti infective*" OR antiviral* OR "anti viral*" OR vermifuge* OR antiparasitic* OR "anti parasitic*" OR anthelmintic* OR antihelmintic* or wormer)	heifer* OR bull OR bulls OR bovine OR sheep OR lamb* OR ewe OR ewes OR ram OR rams OR ovine OR dairy)	OR Scotland OR Scottish OR UK OR "united kingdom")	treatment* OR incidence OR prevalence OR risk OR "risk factor" OR driver)

144

145 To complement the search in scientific databases and achieve a complete systematic
146 review, grey literature was searched using the methodology described by Mahood *et al.*
147 (2014) to screen for data sets and reports. Rather than using open search engines (e.g.
148 Google.com) which may result in unreliable sources, we targeted government data sets
149 (Piasecki *et al.* 2018). The UK's government's data repositories[3] were searched using the
150 same search terms and parameters as described in Table 1. The only difference is that the
151 government search function is not as sophisticated; only using the Boolean operator 'AND'.

152

153 ## Relevance screening and full text appraisal

154 After duplicate removal, two reviewers (MH and LW) independently reviewed the same 10%
155 of the articles (n=69), selected by random using a random number generator in Excel, by
156 title and abstract using the PICOS inclusion criteria. Once both reviewers had screened the
157 sample articles, the conclusion on whether to include or exclude were compared in order to
158 measure the inter-rater reliability using observed proportional agreement and Cohen's
159 kappa, calculated manually using the method described by Cohen (1960) (Supp. 1; part 8).
160 Observed proportional agreement between the two observers was 91.3%, with a
161 corresponding Cohen's kappa of 0.812 indicating strong level inter-rater reliability IRR. The
162 reviewers discussed the six articles on which they disagreed in order to reach a consensus

[3] https://www.gov.uk/search/research-and-statistics

and to clarify the screening criteria. Given the high level of IRR, it was deemed acceptable to allow a single reviewer (MH) to screen the remaining articles and apply inclusion and exclusion criteria. Full text appraisal of the remaining articles was completed by two independent reviewers (MH and LW). Grey literature records were screened for relevance using the same PICOS inclusion criteria. During the review process citation lists were examined to check recall accuracy and to identify possible additional articles for inclusion in the review.

Data extraction

Data was extracted from both the included scientific articles and reports into Microsoft Excel (version 16.33); capturing data on the target population, area of interest, geographic location, study design, and outcome indicators (such as the number of farms using antimicrobials, percentage of bacterial isolates resistant to antibiotics, or proportion of farms with anthelmintic resistance) (a summary of which is presented in Supp. 2). Where reports contained disaggregated data (such as antibiotic resistance profiles by species, region, and year), this data was extracted and collated to allow visualisation of trends. Where sources contained data relating to the United Kingdom, rather than Britain (the focus of this review), data was disaggregated into constituent countries.

Results

Summary of articles

A total of 773 articles were screened for this review: 687 primary articles identified through searching Scopus, Web of Science, and Medline, 83 documents and reports identified through a grey literature search, and 3 additional articles identified by examining the citation lists of these primary articles. All articles were written in English; no exclusion of articles was done based on language.

Figure 1. Flow chart documenting literature retrieval and criteria used to select articles and reports for inclusion in the systematic review of anti-infective agents in sheep and cattle populations in Britain.

Descriptive statistics of selected articles and reports

Of the final 40 articles half focused solely on cattle, 19 focused solely on sheep, and one article contained data on both species. Most articles (29/40) contained data on resistance to anti-infective agents while fewer articles (15/40) contained data on the usage of anti-infective agents (Table 2). Four articles contained data relating to more than one area of interest.

Table 2. Topic areas covered in articles

Area of interest	Number of articles	% of articles
Antibiotic usage	10	25
Antibiotic resistance	16	40
Anthelmintic usage	6	15
Anthelmintic resistance	12	30
Anti-ectoparasitic resistance	2	5

NB. Total number of articles and reports exceeds 40 as some records contained data on more than one area of interest

The grey literature reports included two relevant data series; annual data for Veterinary Antimicrobial Resistance and Sales Surveillance (VARSS) published by the Veterinary Medicines Directorate (VMD) in 2013, 2015 and 2018, and reports on antibiotic usage from the task force for Responsible Use of Medicines in Agriculture (RUMA) published in 2018 and 2019.

A total of 36 articles (90%) covered population data from England, 25 (62.5%) from Wales, and 20 (50%) from Scotland (total number of articles exceeds 40 as many articles contained data on more than one country).

Antibiotic use

Antibiotic usage was detailed in the results of nine (23%) of the articles (five focused on cattle and four focused on sheep) (Table 3). Seven of the nine articles (78%) targeted farmers for data collection using a questionnaire-based approach and in the remaining two veterinary sales data were used.

The five reports used antibiotic sales data collected from veterinary practices and pharmaceutical companies as part of nationwide antibiotic use surveillance. For cattle, data on antibiotic usage were reported by RUMA and the VMD over a four- and five-year period, respectively. The RUMA reports use benchmark values for antibiotic usage in dairy cattle provided by two groups of dairy farms from Kite Consulting and Solway Vets (n=674) and

from Kingshay consultants (n=409). The 2019 RUMA report contained information on 3,458 beef farms (representing 5.5% of British production) and 2,978 dairy farms (30% of the national herd) collected from veterinary practice sales data by FarmVet Systems[4]. For sheep, the reports contained information on antibiotic usage from a single study by Davies *et al*. (2017) already included in this review.

Table 3. Studies on antibiotic usage

Study	Population	Location	Random sampling	Sample size (farms)	Method of data collection
Rutherford et al., 2015	Sheep	EN, WA	No	19	Questionnaire
Davies et al., 2017	Sheep	EN, SC, WA	No	207	Practice sales data
O'Kane et al., 2017	Sheep	EN	Yes	1294	Questionnaire
Lima et al., 2019	Sheep	EN, SC, WA	No	648	Questionnaire
Brunton et al., 2012	Cattle	EN, WA	Yes	557	Questionnaire
Horseman et al., 2013	Cattle	EN, SC, WA	Yes	84	Questionnaire
Hyde et al., 2017	Cattle	EN	No	332	Practice sales data
Fujiwara et al., 2018	Cattle	EN, WA, SC	No	148	Questionnaire
Higham et al., 2018	Cattle	EN, WA, SC	No	372	Questionnaire

EN = England, SC = Scotland, WA = Wales

The majority of the studies produced a proportional outcome metric related to a particular farming practice (for example; the % of farmers using antibiotics to treat lameness). Two studies used practice sales data and details of farm flock and herd compositions to generate estimates of antibiotic use in milligrams per population corrected unit (mg/PCU), defined daily doses vet (DDDvet), and defined course doses vet (DCDvet).

Antibiotic usage in sheep

The three studies looking at antibiotic usage in sheep from farm level data described usage regarding the treatment of footrot (one of the lead causes of lameness in sheep) and new born lambs; the proportion of farmers using antibiotic injections to treat footrot was found to be 24.4% (O'Kane *et al.* 2017), and the proportion of farmers administering prophylactic antibiotics to new born lambs was 26.8% in a general population of sheep farms (Lima et al., 2019) and 73.7% in a population of sheep farms which reported to have joint ill present (Rutherford et al., 2015).

[4] FarmVet Systems, provided by software company VetIMPRESS; www.vetimpress.com

In the study by Davies *et al*. (2017) which looked at antibiotic use in 207 sheep farms, antibiotic usage was found to have a mean mg/PCU of 11.38 (s.d. 15.35, range 0-116.9), 1.47 DDDvet (s.d. 2.1), and 0.39 DCDvet per ewe per flock. The most common classes of antibiotics used were; tetracyclines (57.4%), penicillins (23.7%), and aminoglycosides (10.7%). Antibiotics were predominately administered parenterally (84.4% of the time).

Antibiotic usage in cattle

The five studies looking at antibiotic usage in cattle described the treatment of mastitis and lameness in dairy cattle. Mastitis was found to be the most common reason for the use of antibiotics (Higham *et al*., 2018), with 93% of farmers using antibiotic intra-mammary tubes to treat mastitis during the lactation (Brunton *et al*., 2012), and 96% of farmers using antibiotic dry cow intra-mammary tubes (Fujiwara *et al*., 2018). Regarding lameness treatment (sole ulcer, sole bruising, and white line disease) 55% of farmers reported using injectable antibiotics as an option to treat clinical cases (Horseman et al., 2013).

In the study by Hyde et al. (2017) on 332 dairy farms, antibiotic usage was found to have a mean mg/PCU of 22.11 (range 0.36-97.79), 4.22 DDDvet (range 0.05-20.29), and 1.93 DCDvet (range 0.01-6.74). The most common type of antibiotics used were beta-lactams and aminoglycosides which comprised 42.8% and 20.9% respectively. Parenteral treatment was the most common route of administration (78.1% of the time).

The VMD and RUMA reports contained antibiotic consumption data from 2014-2018 for dairy and beef production systems and are shown in tables 4 and 5.

Table 4. Antibiotic usage in cattle by class (VMD 2019b)

Antibiotic	**Beef mg/kg (%)**	**% change 2017-2018**	**Dairy mg/kg (%)**	**% change 2017-2018**
Penicillin and 1st generation cephalosporins	5.0 (24)	+28	5.5 (32)	+8
Tetracyclines	7.3 (35)	-16	3.2 (19)	+14
Aminoglycosides	3.8 (18)	+31	3.5 (20)	+13
Macrolides	1.7 (8)	+13	1.9 (11)	-2
Trimethoprim/sulphonamides	1.3 (6)	+30	1.9 (11)	+20

Table 5. Antibiotic usage in beef and dairy cattle (RUMA 2019; VMD 2019b)

	Baseline (2016)[5]	2017-2018	2018-2019	% change compared to baseline
Total usage (mg/kg)				
FarmVet Systems				
Beef	-	19	21	
Dairy	26.2	16	17	-29.2
Kite consultants & Solway Vets				
Dairy	26.2	23.7	21.9	-16.4
Kingshay consultants				
Dairy	26.2	20.5	17.3	-34.0
Intramammary tubes (DCDVet)				
UK-VARSS				
Dry cow	0.732	0.547	0.644	-12
Lactating cow	0.808	0.694	0.776	-4
Kite consultants & Solway Vets				
Dry cow	0.732	0.5	0.46	-37
Lactating cow	0.808	0.66	0.55	-32
Kingshay consultants				
Dry cow	0.732	0.522	0.519	-29
Lactating cow	0.808	0.801	0.601	-26

Antibiotic resistance

Of the 40 articles, 16 contained information about antibiotic resistance; 12 (75%) about resistance in cattle, three (19%) in sheep and one of the studies contained information about both cattle and sheep (6%) (Table 6).

Nine of the studies (56%) conducted bacterial identification and resistance testing from samples collected from farms (e.g. from bulk milk tanks or clinical cases) while the remaining seven studies (44%) analysed pre-existing laboratory data. From the 16 studies, eight (50%) focused on *Enterobacteriaceae* species with *Escherichia coli* (*E. coli*) being the most common organism profiled, followed by *Staphylococcus aureus (S. aureus)* in 4/16 (25%). Two studies (13%) used a form of random sampling in their study design.

Table 6. Studies on antibiotic resistance

Study	Population	Location	Random sampling	Sample size	Source of samples	Organism
Wu et al., 2014	Sheep	EN, SC, WA	No	41	Pre-existing laboratory samples	*Campylobacter jejuni*

[5] Baseline data taken from a single source; FarmVet Systems

Rutherford et al., 2015	Sheep	EN, WA	No	25	On farm sampling (sheep with joint ill)	*Streptococcus dysgalactiae*
Angell et al., 2015	Sheep	EN, WA	No	20	On farm sampling (sheep with CODD lesions)	*Treponema spp.*
Cheney et al., 2015	Sheep	EN, WA	No	101	Pre-existing laboratory samples	*Escherichia coli*
García-Álvarez et al., 2011	Cattle	EN	No	940	Pre-existing laboratory samples	*Staphylococcus aureus*
Warner et al., 2011	Cattle	EN, WA	No	65	On farm sampling *(not stated from where)*	*Escherichia coli*
Wu et al., 2012	Cattle	EN, WA	No	34	Pre-existing laboratory samples	*Escherichia coli*
Paterson et al., 2012	Cattle	EN, SC, WA	No	1500	On farm sampling (bulk milk tank)	*Staphylococcus aureus*
Randall et al., 2014	Cattle	EN, WA	Yes	103	On farm sampling (waste milk samples)	*Entero-bacteriaceae*
Ayling et al., 2014	Cattle	EN, SC, WA	No	45	Pre-existing laboratory samples	*Mycoplasma bovis*
Paterson et al., 2014	Cattle	EN, SC, WA	Yes	1090	On farm sampling (bulk milk samples)	*Staphylococcus aureus*
Cheney et al., 2015	Cattle	EN, WA	No	534	Pre-existing laboratory samples	*Escherichia coli*
Thomas et al., 2015	Cattle	EN, SC, WA	No	-	On farm sampling (mastitis cases)	*Escherichia coli* *Staphylococcus aureus* *Strep. uberis*
MacFadyen et al., 2018	Cattle	EN, WA	No	1100	On farm sampling (bulk milk samples)	*Macrococcus caseolyticus*
Mellor et al., 2019	Cattle	EN, SC, WA	No	1115	Pre-existing laboratory samples	*Salmonella typhimurium*
Mueller-Doblies et al., 2018	Cattle	EN, SC, WA	No	45,336	Pre-existing laboratory samples	*Salmonella typhimurium*
Velasova et al., 2019	Cattle	EN, SC, WA	No	40	On farm sampling (faecal samples)	*Entero-bacteriaceae*

NB. EN = England, SC = Scotland, WA = Wales

The 2018 VARSS reports contained information on antibiotic resistance in both sheep and cattle (as well as other animals) collated from samples sent to the Animal and Plant Health Agency (APHA) laboratories for diagnostic purposes (VMD 2019b). Antibiotic resistance was reported for the major livestock bacterial pathogens (such as species causing mastitis and respiratory disease) as well as marker bacterial species significant to human health (such as *E. coli* and *Salmonella spp.*) collected from livestock faecal samples (Table 7).

Table 7. Samples submitted to APHA included in the 2018 VARSS report (VMD 2019b)

Population	**Location**	**Sample size**	**Organism**	**Data range available**
Sheep	UK	22	*Streptococcus dysgalactiae*	2011-2018

Sheep	UK	81	*Mannheimia haemolytica*	2011-2018
Sheep	UK	50	*Biberstein trehalosi*	2011-2018
Sheep	EN, WA	72-161	*Escherichia coli*	2013-2018
	SC	67		
Sheep	EN, WA	276	*Salmonella spp.*	2013-2018
	SC	68		
Cattle	UK	110	*Escherichia coli*	2011-2018
Cattle	UK	32	*Streptococcus dysgalactiae*	2011-2018
Cattle	UK	84	*Streptococcus uberis*	2011-2018
Cattle	UK	36	*Staphylococcus aureus*	2011-2018
Cattle	UK	76	*Pasteurella multocida*	2011-2018
Cattle	UK	44	*Mannheimia haemolytica*	2011-2018
Cattle	EN+WA	208	*Escherichia coli*	2013-2018
	SC	157-313		
Cattle	EN+WA	489	*Salmonella spp.*	2013-2018
	SC	140		

NB. EN = England, SC = Scotland, WA = Wales, UK = United Kingdom

Antibiotic resistance in sheep

The four studies investigating antibiotic resistance in sheep reported on four different organisms; *E.coli, Campylobacter jejuni* (*C. jejuni*)*, Streptococcus dysgalactiae* (*S. dysgalactiae*)*,* and *Treponema* species*.* In their study of antibiotic resistance of *E. coli* from diseased farm livestock, Cheney et al. (2015), found that 57.4% of non-verotoxigenic *E. coli* were resistant to at least one antimicrobial and the highest level of resistance for tetracycline (56.4% of isolates), sulphonamides (48.5%), ampicillin (37.6%), and streptomycin (31.7%). A study of abortion associated with *C. jejuni* by Wu et al. (2014) found that of the 42 isolates, 17.1% were resistant to nalidixic acid, 9.8% resistant to clindamycin, 4.9% resistant to tetracyclines, and 2.4% resistant to azithromycin (the authors did not state what percentage of isolates were resistant to at least one antimicrobial). In a study of *S. dysgalactiae* isolated from sheep with joint ill, Rutherford et al. (2015) reported that all 25 isolates were resistant to tetracycline. Angell et al. (2015) tested the in-vitro susceptibility of contagious ovine digital dermatitis associated *Treponema* species and found that all 20 isolates were susceptible to ten different antibiotics.

The VARSS 2018 report showed high a level of resistance to tetracyclines in *S. dysgalactiae* and *Mannheimia haemolytica* (Table 8; VARSS 2018).

Table 8. Antibiotic resistance in major sheep pathogens taken from VARSS 2018 report

		Resistant isolates (%)						
	Number of isolates	Ampicillin	Amoxicillin/ clavulanate	Enrofloxacin	Trimethoprim	Tetracycline	Neomycin	Tylosin
Common mastitis pathogens:								
Streptococcus dysgalactiae	22	0	0			77.3		0
Common respiratory pathogens:								
Mannheimia haemolytica	81	2.5	0	0	0	46.9		
Bibersteinia trehalosi	50	0	0	0	0	2.0		

NB. In sheep, *Mannheimia haemolytica* can also cause mastitis

303

304 High levels of antibiotic resistance were reported in isolates of *E. coli* from sheep in England,
305 Wales, and Scotland, with the highest levels detected to tetracycline, ampicillin, and
306 spectinomycin in all countries, streptomycin in England and Wales, and
307 amoxicillin/clavulanate in Scotland (Figure 2; VARSS 2013-2018). Levels of resistance were
308 found to be decreasing in *E. coli* in sheep in England and Wales, while levels of resistance in
309 sheep in Scotland showed an increase over the last two years.

310 **Figure 2.** Percentage of *E. coli* isolates from sheep resistant to different antibiotics in (A) England and Wales,
311 and (B) Scotland

312

313 In 2018, the highest level of resistance in *Salmonella spp.* from sheep in England and Wales
314 was to streptomycin (7.6% of isolates), and in Scotland was to sulphonamide compounds
315 (11.8% of isolates) (Figure 3; VARSS 2013-2018).

316 **Figure 3.** Percentage of *Salmonella* isolates from sheep resistant to different antibiotics in (A) England and
317 Wales, and (B) Scotland

318

319 **Antibiotic resistance in cattle**

320 Four studies reported on the resistance profiles to *S. aureus;* two examining isolates from
321 mastitis cases and two examining isolates from bulk milk samples. Thomas et al., (2015)
322 found that of the 38 *S. aureus* isolates from mastitis cases, 31.6% were resistant to penicillin
323 G, and García-Álvarez et al., (2011) found that of the 940 *S. aureus* isolates from mastitis
324 cases, 2.6% were resistant to methicillin, though none were positive for the *mecA* gene

(used to confirm methicillin-resistant *S. aureus* [MRSA]). Paterson et al. (2012) identified 300 MRSA isolates from 1500 bulk milk samples and found that seven of the isolates (originating from five geographically remote locations) were *mecA* positive and belonged to the clonal complex CC398. Another study from the same author documented the presence of *mecC* MRSA in ten out of 375 (2.7%) English farms and one sample of *mecA* MRSA (Paterson et al., 2014).

Three articles described three miscellaneous bacteria; *Mycoplasma bovis, Streptococcus uberis (S. uberis),* and *Macrococcus caseolyticus*. Ayling et al., (2014) reported that *Mycoplasma bovis* had shown increasing levels of resistance over a five-year period (between 2004 and 2009), demonstrated by rising MIC50 levels, though as minimum inhibitory concentrations to define resistance have not been set for this bacterium the prevalence of resistance could not be stated. Thomas et al., (2015) reported that in 39 isolates of *S. uberis,* 12.8% and 7.7% were resistant to tetracycline and erythromycin respectively. In their study of *Macrococcus caseolyticus,* MacFayden et al., (2018) found that all the 33 isolates grown from bulk milk tanks were positive for *mecB* and *mecD*.

Studies which investigated *Enterobacteriaceae* species included those which looked for extended spectrum beta lactamase (ESBL) markers in various bacteria and those which reported on resistance in specific bacterial species (Table 9).

Table 9. Antibiotic resistance in *Enterobacteriaceae* species

Study	**Source of samples**	**Resistance**
Randall et al., 2014	Waste milk samples (n=103)	6.8% samples positive for ESBL
Velasova et al., 2019	Faecal samples (n=40)	25% samples positive for ESBL
Warner et al., 2011	On farm sampling (n=65)	ESBL *E. coli* found on 43.1% of farms
Cheney et al., 2015	Pre-existing lab samples (n=534)	84.1% non-VTEC *E. coli* resistant to at least one antibiotic 56.5% VTEC *E. coli* resistant to at least one antibiotic
Wu et al., 2012	Pre-existing lab samples (n=34)	61.7% of *E. coli* with at least one antibiotic resistant gene
Mueller-Doblies et al., 2018	Pre-existing lab samples (n=244)	69.2% of *Salmonella* isolates resistant to one of more antibiotics
Mellor et al., 2019	Pre-existing lab samples (n=1115)	85.4% of *Salmonella* isolates resistant to one of more antibiotics 74.7% of *Salmonella* isolates resistant to three or more antibiotics

ESBL= Extended spectrum beta lactamase; *E. coli* = *Escherichia coli;* VTEC= Verotoxigenic *E. coli*

Cheney et al. (2015), found high levels of resistance in *E. coli* to sulphonamides (73.6% of isolates), tetracycline (70.7% of isolates), ampicillin (69.5% of isolates), and streptomycin (48.5% of isolates). The VARSS 2018 report recorded a high level of resistance to tetracyclines in the following bacterial species: *S. dysgalactiae, Pasteurella multocida, S. uberis,* and *Mannheimia haemolytica* and a high level of resistance to neomycin in *S. uberis* (Table 10; VMD 2018).

Table 10. Antibiotic resistance in major cattle pathogens taken from VARSS 2018 report

		Resistant isolates (%)						
	Number of isolates	Ampicillin	Amoxicillin/ clavulanate	Enrofloxacin	Trimethoprim	Tetracycline	Neomycin	Tylosin
Common mastitis pathogens:								
Escherichia coli	110	21.8	5.5	2.7	6.4	13.6	2.7	
Streptococcus dsygalactiae	32	0	0			87.5	3.1	0
Streptococcus uberis	84	0	0			34.5	45.2	11.9
Staphylococcus aureus	36	27.8	0			2.8	0	2.8
Common respiratory pathogens:								
Pasteurella multocida	76	2.6	0	0	0	51.3		
Mannheimia haemolytica	44	2.3	0	0	0	50		

Overall the level of antibiotic resistance in *E. coli* reported was higher in cattle from England and Wales compared to Scotland. In all countries the highest levels of resistance were recorded to ampicillin and tetracycline. Resistance levels were found to be decreasing in *E. coli* from cattle in England and Wales. While resistance levels were also found to be decreasing in *E. coli* from cattle in Scotland from 2013 to 2017, resistance increased in 2018 (Figure 4; VMD 2013-2018).

Figure 4. Percentage of *E. coli* isolates from cattle resistant to different antibiotics in (A) England and Wales, and (B) Scotland

In 2018 the highest level of resistance in *Salmonella spp.* from cattle in England and Wales was to streptomycin and sulphonamide compounds (both 13.9% of isolates), and in Scotland was to sulphonamide compounds (15.7% of isolates) (Figure 5; VMD 2013-2018).

Figure 5. Percentage of *Salmonella* isolates from cattle resistant to different antibiotics in (A) England and Wales, and (B) Scotland

Anthelmintic use

Of the 40 articles, six (15%) looked at anthelmintic usage; five in sheep and one in cattle (Table 11). All of the studies used farm level data to measure usage and was either captured by farmers self-reporting through questionnaires (n=5), or by ascertaining baseline usage levels before conducting trials into anthelmintic resistance (n=1). No reports were found reporting anthelmintic usage.

Anthelmintics are separated into five major groups; broad spectrum anthelmintics active against major species of helminths and some ectoparasites (groups 1-3); group 1-BZ (benzimidazoles), group 2-LV (imidazothiazoles, including levamisole), group 3-ML (macrocyclic-lactones), and newer generation anthelmintics (groups 4 & 5); group 4-AAD (amino-acetonitrile derivatives), and group 5-SI (spiro-indoles, such as derquantel, available as combination products) (Kaminsky *et al.* 2008; Little *et al.* 2011).

Table 11. Studies on anthelmintic usage

Study	Population	Location	Random sampling	Sample size (farms)	Method of data collection
Burgess et al., 2012	Sheep	EN, SC, WA	No	118	Questionnaire
Morgan et al., 2012	Sheep	EN, SC, WA	Yes	600	Questionnaire
Crilly et al., 2015	Sheep	SC	No	38	Questionnaire
Learmount et al., 2016	Sheep	EN, WA	No	14	Routine usage recorded as part of trial
Lima et al., 2019	Sheep	EN, SC, WA	No	615	Questionnaire
Bellet et al., 2018	Cattle	EN	No	43	Questionnaire

NB. EN = England, SC = Scotland, WA = Wales

Anthelmintic use in sheep

Of the six studies, two described the routine use of anthelmintics. In a study of 118 sheep farms, Burgess et al., (2012) reported that 99% of farmers gave treatment against nematodes and in a study of 600 farms, Morgan et al., (2012) reported that 93%, 67%, and 58% of farmers routinely treated against nematodes, liver fluke, and tapeworms respectively. Two studies reported on specific farming practices; in their study of 615 sheep

388 farms, Lima *et al.* (2019) reported that farmers administered a group four or five
389 anthelmintic (monepantel and derquantel) to 32% and 28% of ewes and rams at quarantine.
390 Crilly et al., (2015) reported that 27 out of 38 farmers (71%) used moxidectin (a macrocyclic
391 lactone) for the periparturient treatment of ewes. Macrocyclic lactones (group three
392 anthelmintics) were reported by three studies to be the most commonly used anthelmintic
393 against nematodes; 56% of 118 farms (Burgess et al., 2012), 47% of 600 farms (Morgan et
394 al., 2012), and 84% (SCOPS farms[6]) and 70% (non SCOPS farms) in a study of 14 farms
395 (Learmount et al., 2016). Benzimidazoles (group one anthelmintics) were reported to be
396 used against nematodes in 31% of 118 farms (Burgess et al. 2012), 26% of 600 farms
397 (Morgan et al., 2012), and 7% (SCOPS farms) and 21% (non SCOPS farms) in a study of 14
398 farms (Learmount et al., 2016). Levamisole (group two anthelmintics) had the lowest
399 reported use, ranging from 28-31% of 118 farms (Burgess et al., 2012), 16% of 600 farms
400 (Morgan et al., 2012), to 9% of 14 farms (Learmount et al., 2016).

401 The mean number of times ewes were treated annually for nematodes (any class of
402 anthelmintic) was reported to be 2.0 (Burgess et al., 2012), 2.35 (s.d. 1.48, range 0-12)
403 (Morgan et al., 2012), and 2.4 (Learmount et al., 2016). The mean number of times lambs
404 were treated for nematodes was reported to be 3.3 (Burgess et al., 2012), 3.55 (s.d. 2.76,
405 range 0-16) (Morgan et al., 2012), and 4.1 (Learmount et al., 2016). Learmount et al., (2016)
406 also reported that those farms following the SCOPS guidelines used significantly fewer
407 treatments in both ewes (ewes on SCOPS farms being treated between zero and three times
408 per year compared to non-SCOPS farms treating between zero and five times per year) and
409 lambs (lambs on SCOPS farms being treated between zero and five times per year compared
410 to non-SCOPS farms treating between zero and eight times per year), though it should be
411 noted that this study only contained seven SCOPS and seven non SCOPS farms.

412

413 **Anthelmintic usage in cattle**

414 Only one study, (Bellet et al., 2018) consisting of 43 farms reported on the use of
415 anthelmintics in cattle and found that farmers routinely used anthelmintics on 85% and 44%

[6] SCOPS – Sustainable Control of Parasites in Sheep (SCOPS 2019)

of their young stock and adult cows respectively. As with the sheep studies, the most common anthelmintic class used in young stock was macrocyclic lactones (89% of farms), which is consistent with the industry led cattle parasite guideline Control of Worms Sustainably (COWS) which recommend macrocyclic lactones as a first line treatment against the parasites *Ostertagia ostertagi* and *Cooperia oncophora* (COWS 2019b).

Anthelmintic resistance

Twelve of the 40 studies (30%) reported on anthelmintic resistance; ten in sheep and two in cattle (Table 12). No grey literature sources were found reporting anthelmintic resistance.

Faecal egg count reduction tests (FECRT) were used to test for resistance in the majority (n=9) of the studies; other tests for resistance were the larval development test (LDT) (n=4), egg hatch test (n=1), and farmer self-reported resistance (n=1).

Table 12. Studies on anthelmintic resistance

Study	Population	Location	Random sampling	Sample size (farms)	Method of resistance testing
Taylor et al., 2009	Sheep	EN, WA	No	40	FECRT & LDT
Mitchell et al., 2010	Sheep	WA	No	122	LDT
Burgess et al., 2012	Sheep	EN, SC, WA	No	118	Self-reported resistance
Jones et al., 2012	Sheep	EN, WA	No	11	FECRT
Daniel et al., 2012	Sheep	EN, SC, WA	No	25	FECRT
Stubbings and SCOPS, 2012	Sheep	EN, SC, WA	No	16	FECRT
Thomas, 2015	Sheep	WA	No	58	FECRT, LDT, EHT
Learmount et al., 2016a	Sheep	EN, WA	No	14	LDT
Glover et al., 2017	Sheep	EN	No	27	FECRT
Kamaludeen et al., 2019	Sheep	EN, WA	Partly	74	FECRT
McArthur et al., 2011	Cattle	SC	No	4	FECRT
Geurden et al., 2015	Cattle	EN, SC, WA	No	10	FECRT

EN = England, SC = Scotland, WA = Wales
FECRT = Faecal egg count reduction test, LDT = Larval development test, EHT = Egg hatch test

Anthelmintic resistance in sheep

Eight of the studies reported on the resistance of nematodes to anthelmintics, either generally, or specifically for *Teladorsagia* and *Trichostrongylus* (Table 13). In their study of 122 sheep farms in Wales, Mitchell et al., (2010) reported nematodes resistance in 100 farms (82.0%) consisting of resistance to benzimidazole only, benzimidazole and levamisole, and to levamisole only, in 56 (46%), 38 (31%), and six (5%), of farms respectively. In another study of 58 sheep farms in Wales, Thomas (2015) reported nematode resistance in 47 farms (81%), consisting of resistance to benzimidazoles, levamisole, and macrocyclic lactones in 44 (75.9%), 32 (55.2%), and 33 (56.9%) of farms respectively. Ten farms had single resistance, 16 farms had double resistance, 13 had triple resistance; and 7 had triple resistance plus moxidectin (ibid). In a study of 25 sheep farms in England, Glover et al., (2017) reported resistance for benzimidazoles, levamisole, and macrocyclic lactones in 24 (96%), 15 (60%), and 18 (67%) of farms. Three farms had single resistance (to benzimidazoles), 11 farms had double resistance, and ten had triple resistance (ibid).

Table 13. Nematode resistance

Study	No of farms	Nematode	Overall	1-BZ	2-LV	3-ML
Taylor et al., 2009	40	*Teladorsagia*		97.5%	40%	
		Trichostrongylus		44%	50%	
Mitchell et al., 2010	122	Unspecified	82%	77%	37%	
Burgess et al., 2012	118	*Trichostrongylus*	18%	17.8%	3.4%	
Jones et al., 2012	11	*Trichostrongylus*				55%
Stubbings and SCOPS, 2012	16	*Trichostrongylus*				62.5%
Thomas, 2015	58	Unspecified	81%	75.9%	55.2%	56.9%
Glover et al., 2017	25	Unspecified	96%	96%	60%	67%
Learmount et al., 2016a	14	*Teladorsagia*		100%		
		Trichostrongylus		100%		

1-BZ = group 1 (Benzimidazole), 2-LV = group 2 (Levamisole), 3-ML = group 3 (macrocyclic lactone)

Two studies reported on the resistance of *Fasciola hepatica* (liver fluke) in sheep to triclabendazole. In a study of 26 farms in England and Wales, Kamaludeen et al., (2019) reported that 21 of the farms (80.8%) showed a reduction in triclabendazole efficacy with nine farms showing a complete lack of efficacy and no change in post treatment faecal egg count. Daniel et al., (2012) reported that of 15 farms in the study, seven (six in Wales and

one in Scotland) were found to have triclabendazole resistance, though there was no indication of resistance in the ten farms sampled from England.

Anthelmintic resistance in cattle

Two studies reported on the resistance to macrocyclic lactones (ivermectin and moxidectin) to *Cooperia oncophora* and *Ostertagia ostertagi* though both studies contained a small number of farms. McArthur et al., (2011) reported that three out of four farms had FECRT results consistent with *Cooperia* resistance to ivermectin. Geurden et al., (2015) reported that out of ten farms, one and five farms had confirmed and inconclusive resistance to moxidectin respectively, and three and four farms had confirmed and inconclusive resistance to ivermectin respectively; resistant species were *Cooperia* and *Ostertagia.*

Anti-ectoparasitic usage & resistance

Two articles contained data concerning ectoparasites, one on the usage and one on the resistance of anti-ectoparasitics. Crilly et al., (2015), reported that 61% of farms (39% using injectable macrocyclic lactones and 21 using organophosphate dips) in Scotland use whole flock treatment for *Psoroptes ovis* (sheep scab), and Doherty et al., (2018), reported on the novel resistance of *Psoroptes ovis* to macrocyclic lactones in a study of four farms in England and Wales.

Discussion

General

Although the importance of anti-infectives and the risk of resistance development are widely discussed (DANMAP 2016, Dorado-Garcia *et al.* 2016, Veldman et al., 2017), we identified a low number of publications (40 papers and two report series) reporting use or resistance in sheep and cattle in Britain. There were marked differences between the number of papers focussing on cattle compared to sheep, with 60% of the papers focusing on usage and 76% on resistance in cattle only. Similarly, both report series only contained primary antimicrobial usage data in cattle and not in sheep. Cattle, especially dairy, may be

the greater focus of attention due to the more intensive way they are farmed, with increased contact time between professionals (both farmers and veterinarians) compared to sheep. Other ways that cattle gain more attention than sheep is that beef markets are offered more protections under the EU's Common Market Organisation than sheep markets and additionally, beef is consumed, exported and imported more than sheep meat (AHDB 2019a, 2019b). This gap in interest and knowledge of what appears to be a neglected species warrants more attention and research.

Antibiotic usage

From the data extracted in this review, antibiotic use in sheep and cattle in Britain appear similar to each other, similar to the level observed in poultry, and below the UK average for all livestock (which is elevated by the relatively high usage levels reported in pigs). The marked difference to pig production is likely due to the less intensive nature of production compared to the pig sector, where prophylactic and metaphylactic use of antibiotics to avoid infectious diseases occurs in many farrow-to-finish and fattening farms (Lekagul *et al.* 2019). While poultry production in the Britain is often highly intensive, the ability to achieve high levels of biosecurity (such as occurs in closed housing systems) support production systems that are not heavily reliant on antibiotics (DEFRA 2020). However, a major caveat of these findings is the poor level of coverage afforded to sheep and cattle (especially beef production systems) in Britain; small sampling sizes with frequent use of convenience sampling over random sampling are likely to lead to unrepresentative results. In comparison, the pig sector utilises an electronic medicine book (eMB-pigs) to allow farmers to regularly upload antibiotic usage and represents 87% of UK pig producers (DHSC 2019).

Mastitis being the most common use for antibiotics in dairy cattle in Britain is consistent with other high dairy producing countries such as the USA and New Zealand (Denis *et al.* 2009; Landers *et al.* 2012). Antibiotic usage in dairy cattle due to mastitis has followed a downward trend over the last three years showing reductions in both total usage and in dry and lactating cow treatments. As with other livestock production systems in the UK, tetracyclines and beta-lactam antibiotics (penicillins and first generation cephalosporins) were commonly used antibiotics in sheep and cattle (VMD 2019b), and reflects the WHO's position on restricting the use certain antibiotics (such as third and fourth generation cephalosporins and fluoroquinolones) in non-human species (WHO 2019). Globally,

517 tetracyclines remain the most commonly used antibiotics in livestock production, and are
518 one of the antibiotic groups used for growth promotion in countries which have yet to ban
519 this practice (Granados-Chinchilla and Rodríguez 2017).

520 Many of the scholarly articles described antibiotic usage using in a proportional metric
521 focused at the farm level. While these types of metrics are potentially useful for comparing
522 temporal and spatial trends and providing relatively easy ways of measuring use before and
523 after an intervention, they remain specific to a species, disease, or practice, and are not
524 readily comparable outside of their own sector. However, in this review there were limited
525 instances of proportional metrics being used to make serial or temporal comparisons, thus
526 limiting their usefulness. Furthermore, as the proportional metrics are set at the farm level,
527 they may inflate the magnitude of usage compared to metrics set at the level of individual
528 animals. The production of quantifiable metrics, such as mg/PCU or mg/kg, provide a
529 standardised approach allowing comparisons of usage between species, sectors (livestock
530 and human), and countries, and are advocated as harmonised indicators by both the
531 European Centre for Disease Prevention and Control and the UK One Health report on
532 antibiotic use (VMD 2019a). However, metrics such as mg/kg do not account for the
533 variation in dosage of different antibiotics; for example, newer generation drugs may have a
534 lower mg/kg dose than older ones; thus limiting the use of new generation drugs in favour
535 of older ones may lead to a higher overall mg/kg despite effective antibiotic stewardship
536 (Mills *et al.* 2018). To compensate for this, metrics such as the defined daily dose can be
537 utilised, where the total mg of medicine used is divided by the daily dose, but add an
538 additional level of complexity to data generation. Quantifiable metrics can either be
539 generated from a 'top down' (or consumption level) approach, using national sales data and
540 estimations of total livestock populations (as in the VMD or RUMA reports) and so remain
541 aggregated at the species level; or from a 'bottom up' approach, using veterinary practice
542 sales and farm holding data (as used by Davies *et al.* (2017) and Hyde *et al.* (2017)), and so
543 be more complex and time consuming to generate than consumption level data.
544 Consumption level data can also face problems when antibiotics are licenced for use in
545 more than one species and assumptions need to be made on how usage is divided across
546 species. Given the requirement of farm assurance schemes for farmers to keep records of
547 antibiotic usage, and the high level of digitalisation of veterinary practice sales data,

548 generating additional ‘bottom up’ quantifiable metrics with a wider coverage than is
549 currently available should be possible, albeit not necessarily feasible; Jones-Diette *et al.*
550 (2016) state that veterinary research using electronic records is hindered by the multitude
551 of practice management systems used in the UK.

552 ## Antibiotic resistance

553 Although resistance to some antimicrobials (using *E.coli* as a marker) appears to have
554 decreased in sheep and cattle in England and Wales over the last few years, levels of
555 resistance remain high, particularly for tetracyclines, penicillins, aminoglycosides and
556 sulphonamides in both species and there is some evidence of increasing levels of resistance
557 in Scotland. Additionally, many of the sheep and cattle pathogens responsible for
558 economically important issues such as mastitis and respiratory diseases have high levels of
559 resistance to tetracyclines, one of the most commonly used antibiotics. However, as these
560 findings are derived from bacterial samples submitted to veterinary laboratories selection
561 bias should be considered. Given that submitting samples for bacterial culture and
562 sensitivity is not routine practice for all cases of mastitis or respiratory disease the data will
563 likely reflect the more troublesome clinical cases which have not responded to first line
564 treatment, and so resistance levels in the general population may be lower than reported
565 here. With the exception of ampicillin and neomycin in cattle, resistance of pathogens to
566 other major groups of antibiotics remains low for both species, providing, at least for now,
567 effective alternative treatment options.

568 From a One Health perspective, monitoring the levels of antibiotic resistance in zoonotic
569 pathogens in animals forms an important part of national action plans to tackle
570 antimicrobial resistance. The high level of antibiotic resistance observed in *E. coli* in both
571 sheep and cattle is concerning given that ruminants are an important reservoir for zoonotic
572 verotoxigenic *E. coli* (Fairbrother and Nadeau 2006). While it may appear encouraging to
573 observe that resistance in *E. coli* to enrofloxacin remains below 10% for both sheep and
574 cattle across Britain, fluoroquinolones remain an important group of antibiotics in the
575 treatment of *E. coli* in humans, with increasing reports of resistance in people with uro-
576 genital infections (Zaytoun *et al.* 2011; Talan *et al.* 2016). As with *E.* coli, livestock play an
577 important role in the zoonotic transmission of Salmonella, a major cause of human food
578 poisoning. The lower rate of antibiotic resistance seen in *Salmonella* in sheep and cattle

579 compared to *E.* coli is reflected in findings from other ruminant populations (Scott *et al.*
580 2012). These lower rates of antibiotic resistance may be explained by the less ubiquitous
581 nature of *Salmonella* in ruminant intestinal tracts than *E. coli* (Fegan *et al.* 2004; RODRIGUEZ
582 *et al.* 2006) leading to a lower antibiotic resistance selection pressure for *Salmonella*.

583 ## Anthelmintics

584 Sheep gained more attention than cattle in the area of anthelmintic usage and resistance
585 which may be due to some of the inherent differences between these two species. Sheep
586 experience an increase in faecal parasite output around lambing related to a relaxation of
587 immunity at this time, thought to be more profound in the presence of twins (or triplets), a
588 common occurrence in this species (Fthenakis *et al.* 2015). There is a perception that cattle
589 suffer less with worm burdens than sheep (with the industry led COWS advising that adult
590 cows do not need monitoring for worms unless a problem occurs (COWS 2019a)) and our
591 finding that more data exists for sheep than cattle is reflected in global trends on
592 anthelmintic research (Sutherland and Leathwick 2011).

593 ### Anthelmintic usage

594 The small number and fragmented nature of studies identified by this review describing
595 anthelmintic usage, and the lack of available national sales data, prevented the
596 identification of trends in either sheep or cattle. Collecting data on anthelmintic usage may
597 be confounded by the fact that they are prescribed at a farm rather than animal level, but it
598 should still be possible to see serial and temporal trends. Given the negative economic
599 burden of parasites on livestock production (gastrointestinal parasites are estimated to cost
600 the British sheep industry £84 million annually (Nieuwhof and Bishop 2005)) and two major
601 industry led initiatives to control anthelmintic usage (SCOPS and COWS), this lack of data is
602 surprising, and warrants addressing. For example, it would be prudent to investigate
603 whether the difference identified by Learmount *et al.* (2016) in their small number of SCOPS
604 and non-SCOPS farms, exists on a wider scale, and thus be able to validate the benefit for
605 farmers to follow such guidelines.

606 ### Anthelmintic resistance

607 The high levels of resistance of nematodes in British sheep and cattle to group 1-3
608 anthelmintics is reflected by global trends in livestock (Mphahlele *et al.* 2019). This finding is

concerning, especially given the small number of group 4 and 5 anthelmintics currently available. However, as with anthelmintic usage, the small number of studies focusing on anthelmintic resistance identified by this review warrants attention. The SCOPS guidelines recommend that sheep farmers perform faecal egg counts every two to four weeks during the grazing seasons, and so it could be assumed that data exists at the farm or veterinary practice level detailing anthelmintic resistance on a wider scale than is currently reported.

Conclusion

From the findings of this review we recommend that additional data is needed to understand the current usage of antimicrobials in sheep, and the current usage of, and resistance to anthelmintics in sheep and cattle in Britain. Given the national importance of sheep farming, the lack of research afforded to this species identified by this review is concerning. As identified by two articles in this review, veterinary practice sales data provide a potential valuable resource for measuring antimicrobial usage if effective methods of collecting and collating data can be accomplished on a national scale. When collating and reporting data on antimicrobial usage, researchers and governing bodies should take efforts to produce metrics which are comparable across species, sectors, and time; some of the findings identified by this review were limited in their usefulness due to a lack of comparability. Currently, data on antibiotic resistance in sheep and cattle in Britain is subject to selection bias, being based on specimens from clinical cases, an issue which could be addressed though the development of an active surveillance system, though such a system would require access to adequate resources on a national scale. Additionally, efforts could be made to access data on anthelmintic resistance which exists as part of individual farm health plans so that an assessment can be made about the effectiveness of current strategies to control the development of resistance.

Declaration of interests:

Funding

This study was conducted as part of a project kindly funded by the Cadogan Charity with matched-funding provided by the Royal Veterinary College and the University of Hertfordshire.

Acknowledgments

We would like to acknowledge Houda Bennani for her input into the governance of antimicrobials.

References

AHDB (2017) *Beef Production from the Dairy Herd*, available: https://beefandlamb.ahdb.org.uk/wp-content/uploads/2017/08/Beef-production-from-the-dairy-herd.pdf.

AHDB (2019a) The UK Cattle Yearbook 2019 | AHDB [online], available: https://ahdb.org.uk/knowledge-library/the-uk-cattle-yearbook-2019 [accessed 10 Feb 2020].

AHDB (2019b) The UK Sheep Yearbook 2019 | AHDB [online], available: https://ahdb.org.uk/knowledge-library/the-uk-sheep-yearbook-2019 [accessed 10 Feb 2020].

Angell, J.W., Clegg, S.R., Sullivan, L.E., Duncan, J.S., Grove-White, D.H., Carter, S.D., Evans, N.J. (2015) 'In vitro susceptibility of contagious ovine digital dermatitis associated Treponema spp. isolates to antimicrobial agents in the UK.', *Veterinary dermatology*, 26(6), 484–485, available: http://ovidsp.ovid.com/ovidweb.cgi?T=JS&PAGE=reference&D=med11&NEWS=N&AN=26482550.

Ayling, R.D., Rosales, R.S., Barden, G., Gosney, F.L. (2014) 'Changes in antimicrobial susceptibility of Mycoplasma bovis isolates from Great Britain.', *The Veterinary record*, 175(19), 486, available: http://ovidsp.ovid.com/ovidweb.cgi?T=JS&PAGE=reference&D=med10&NEWS=N&AN=25185107.

Bellet, C., Green, M.J., Bradley, A.J., Kaler, J. (2018) 'A longitudinal study of gastrointestinal parasites in English dairy farms. Practices and factors associated with first lactation heifer exposure to &ITOstertagia ostertagi&IT on pasture', *JOURNAL OF DAIRY SCIENCE*, 101(1), 537–546.

Bennani, H., Mateus, A., Mays, N., Eastmure, E., Stärk, D.C.K., Häsler, B. (2020) 'Overview of Evidence of Antimicrobial Use and Antimicrobial Resistance in the Food Chain', *Antibiotics*.

Bramer, W.M., Rethlefsen, M.L., Kleijnen, J., Franco, O.H. (2017) 'Optimal database

672 combinations for literature searches in systematic reviews: a prospective exploratory
673 study', *Systematic reviews*, 6(1), 245, available:
674 https://pubmed.ncbi.nlm.nih.gov/29208034.

675 Brunton, L.A., Duncan, D., Coldham, N.G., Snow, L.C., Jones, J.R. (2012) 'A survey of
676 antimicrobial usage on dairy farms and waste milk feeding practices in England and
677 Wales.', *The Veterinary record*, [Comment in: Vet Rec. 2012 Oct 27;171(17):429; PMID:
678 23104790 [https://www.ncbi.nlm.nih.gov/pubmed/23104790]], 171(12), 296,
679 available:
680 http://ovidsp.ovid.com/ovidweb.cgi?T=JS&PAGE=reference&D=med8&NEWS=N&AN=2
681 2903925.

682 Burgess, C.G.S., Bartley, Y., Redman, E., Skuce, P.J., Nath, M., Whitelaw, F., Tait, A., Gilleard,
683 J.S., Jackson, F. (2012) 'A survey of the trichostrongylid nematode species present on
684 UK sheep farms and associated anthelmintic control practices.', *Veterinary*
685 *parasitology*, 189(2–4), 299–307, available:
686 http://ovidsp.ovid.com/ovidweb.cgi?T=JS&PAGE=reference&D=med8&NEWS=N&AN=2
687 2560313.

688 BVA (2019) Responsible Use of Antimicrobials [online], available:
689 https://www.bva.co.uk/take-action/our-policies/responsible-use-of-antimicrobials/
690 [accessed 14 May 2020].

691 Chatterjee, A., Modarai, M., Naylor, N.R., Boyd, S.E., Atun, R., Barlow, J., Holmes, A.H.,
692 Johnson, A., Robotham, J. V (2018) 'Quantifying drivers of antibiotic resistance in
693 humans: a systematic review', *The Lancet Infectious Diseases*, 18(12), e368–e378,
694 available: http://www.sciencedirect.com/science/article/pii/S1473309918302962.

695 Cheney, T.E.A., Smith, R.P., Hutchinson, J.P., Brunton, L.A., Pritchard, G., Teale, C.J. (2015)
696 'Cross-sectional survey of antibiotic resistance in Escherichia coli isolated from diseased
697 farm livestock in England and Wales.', *Epidemiology and infection*, 143(12), 2653–2659,
698 available:
699 http://ovidsp.ovid.com/ovidweb.cgi?T=JS&PAGE=reference&D=med11&NEWS=N&AN=
700 25613078.

701 Cohen, J. (1960) 'A Coefficient of Agreement for Nominal Scales', *Educational and*

Psychological Measurement, 20(1), 37–46, available: https://doi.org/10.1177/001316446002000104.

COWS (2019a) Promoting Sustainable Control of Cattle Parasites - COWS - Promoting Sustainable Control of Cattle Parasites [online], available: https://www.cattleparasites.org.uk/ [accessed 14 May 2020].

COWS (2019b) Gut and Lung Worms - COWS - Promoting Sustainable Control of Cattle Parasites [online], available: https://www.cattleparasites.org.uk/gut-and-lung-worms/ [accessed 10 Feb 2020].

Crilly, J.P., Jennings, A., Sargison, N. (2015) 'Patterns of faecal nematode egg shedding after treatment of sheep with a long-acting formulation of moxidectin.', *Veterinary parasitology*, 212(3–4), 275–280, available: http://ovidsp.ovid.com/ovidweb.cgi?T=JS&PAGE=reference&D=med11&NEWS=N&AN=26276580.

Daniel, R., van Dijk, J., Jenkins, T., Akca, A., Mearns, R., Williams, D.J.L. (2012) 'Composite faecal egg count reduction test to detect resistance to triclabendazole in Fasciola hepatica.', *The Veterinary record*, 171(6), 153–155, available: http://ovidsp.ovid.com/ovidweb.cgi?T=JS&PAGE=reference&D=med8&NEWS=N&AN=22791519.

Davies, P., Remnant, J.G., Green, M.J., Gascoigne, E., Gibbon, N., Hyde, R., Porteous, J.R., Schubert, K., Lovatt, F., Corbishley, A. (2017) 'Quantitative analysis of antibiotic usage in British sheep flocks.', *The Veterinary record*, 181(19), 511, available: http://ovidsp.ovid.com/ovidweb.cgi?T=JS&PAGE=reference&D=med13&NEWS=N&AN=29051311.

DEFRA (2019) *The Guide to Cross Compliance in England 2019*, available: https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment_data/file/764890/Cross_Compliance_2019_rules_v1.0.pdf [accessed 17 Mar 2020].

DEFRA (2020) *Biosecurity and Preventing Welfare Impacts in Poultry and Captive Birds*, available: https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attach

732 ment_data/file/877359/biosecurity-poultry-guide.pdf [accessed 18 May 2020].

733 Denis, M., Wedlock, D.N., Lacy-Hulbert, S.J., Hillerton, J.E., Buddle, B.M. (2009) 'Vaccines
734 against bovine mastitis in the New Zealand context: What is the best way forward?',
735 *New Zealand Veterinary Journal*, 57(3), 132–140, available:
736 https://doi.org/10.1080/00480169.2009.36892.

737 DHSC (2019) UK 5-Year Action Plan for Antimicrobial Resistance 2019 to 2024 - GOV.UK
738 [online], available: https://www.gov.uk/government/publications/uk-5-year-action-
739 plan-for-antimicrobial-resistance-2019-to-2024 [accessed 15 May 2020].

740 Doherty, E., Burgess, S., Mitchell, S., Wall, R. (2018) 'First evidence of resistance to
741 macrocyclic lactones in Psoroptes ovis sheep scab mites in the UK', *Veterinary Record*,
742 182(4), 106, available: https://www.scopus.com/inward/record.uri?eid=2-s2.0-
743 85044077888&doi=10.1136%2Fvr.104657&partnerID=40&md5=f297d561bf802b52dd9
744 0dd3e5ea6e609.

745 Dorado-García, A., Mevius, D.J., Jacobs, J.J.H., Van Geijlswijk, I.M., Mouton, J.W., Wagenaar,
746 J.A., Heederik, D.J. (2016) 'Quantitative assessment of antimicrobial resistance in
747 livestock during the course of a nationwide antimicrobial use reduction in the
748 Netherlands', *Journal of Antimicrobial Chemotherapy*, 71(12), 3607–3619, available:
749 https://doi.org/10.1093/jac/dkw308.

750 European Parliament (2019) Veterinary Medicines: Fighting Antibiotic Resistance | News |
751 European Parliament [online], available:
752 https://www.europarl.europa.eu/news/en/headlines/society/20181018STO16580/vet
753 erinary-medicines-fighting-antibiotic-resistance [accessed 14 May 2020].

754 Fairbrother, J.M., Nadeau, É. (2006) 'Escherichia coli: on-farm contamination of animals',
755 *Rev. sci. tech. Off. int. Epiz*, 25(2), 555–569, available:
756 http://www.microbionet.com.au/vtectable.htm [accessed 18 May 2020].

757 FAO (2016) The FAO Action Plan on Antimicrobial Resistance 2016-2020 | Global Forum on
758 Food Security and Nutrition (FSN Forum) [online], available:
759 http://www.fao.org/fsnforum/resources/fsn-resources/fao-action-plan-antimicrobial-
760 resistance-2016-2020 [accessed 19 May 2020].

761 Fegan, N., Vanderlinde, P., Higgs, G., Desmarchelier, P. (2004) 'Quantification and
762 prevalence of Salmonella in beef cattle presenting at slaughter', *Journal of Applied*
763 *Microbiology*, 97(5), 892–898, available: https://doi.org/10.1111/j.1365-
764 2672.2004.02380.x.

765 Fthenakis, G.C., Mavrogianni, V.S., Gallidis, E., Papadopoulos, E. (2015) 'Interactions
766 between parasitic infections and reproductive efficiency in sheep', *Veterinary*
767 *Parasitology*, 208(1), 56–66, available:
768 http://www.sciencedirect.com/science/article/pii/S0304401714006517.

769 Fujiwara, M., Haskell, M.J., Macrae, A.I., Rutherford, K.M.D. (2018) 'Survey of dry cow
770 management on UK commercial dairy farms.', *The Veterinary record*, 183(9), 297,
771 available:
772 http://ovidsp.ovid.com/ovidweb.cgi?T=JS&PAGE=reference&D=med13&NEWS=N&AN=
773 29907660.

774 García-Álvarez, L., Holden, M.T.G., Lindsay, H., Webb, C.R., Brown, D.F.J., Curran, M.D.,
775 Walpole, E., Brooks, K., Pickard, D.J., Teale, C., Parkhill, J., Bentley, S.D., Edwards, G.F.,
776 Girvan, E.K., Kearns, A.M., Pichon, B., Hill, R.L.R., Larsen, A.R., Skov, R.L., Peacock, S.J.,
777 Maskell, D.J., Holmes, M.A. (2011) 'Meticillin-resistant Staphylococcus aureus with a
778 novel mecA homologue in human and bovine populations in the UK and Denmark: A
779 descriptive study', *The Lancet Infectious Diseases*, 11(8), 595–603, available:
780 https://www.scopus.com/inward/record.uri?eid=2-s2.0-
781 79960696292&doi=10.1016%2FS1473-3099%2811%2970126-
782 8&partnerID=40&md5=9db93b84170865f3669ff411a8b9175f.

783 Geurden, T., Chartier, C., Fanke, J., di Regalbono, A.F., Traversa, D., von Samson-
784 Himmelstjerna, G., Demeler, J., Vanimisetti, H.B., Bartram, D.J., Denwood, M.J. (2015)
785 'Anthelmintic resistance to ivermectin and moxidectin in gastrointestinal nematodes of
786 cattle in Europe.', *International journal for parasitology. Drugs and drug resistance*,
787 5(3), 163–171, available:
788 http://ovidsp.ovid.com/ovidweb.cgi?T=JS&PAGE=reference&D=med11&NEWS=N&AN=
789 26448902.

790 Glover, M., Clarke, C., Nabb, L., Schmidt, J. (2017) 'Anthelmintic efficacy on sheep farms in

south-west England.', *The Veterinary record*, 180(15), 378, available: http://ovidsp.ovid.com/ovidweb.cgi?T=JS&PAGE=reference&D=med13&NEWS=N&AN=28167646.

Granados-Chinchilla, F., Rodríguez, C. (2017) 'Tetracyclines in Food and Feedingstuffs: From Regulation to Analytical Methods, Bacterial Resistance, and Environmental and Health Implications', *Journal of analytical methods in chemistry*, 2017, 1315497, available: https://pubmed.ncbi.nlm.nih.gov/28168081.

Heymann, D., Ross, E. (2019) Preserve the Effectiveness of Antibiotics with a Global Treaty | Chatham House [online], *Chatham House*, available: https://www.chathamhouse.org/expert/comment/preserve-effectiveness-antibiotics-global-treaty [accessed 19 May 2020].

Heymann, D.L. (2006) 'Resistance to Anti-Infective Drugs and the Threat to Public Health', *Cell*, 124(4), 671–675, available: http://www.sciencedirect.com/science/article/pii/S009286740600184X.

Higham, L.E., Deakin, A., Tivey, E., Porteus, V., Ridgway, S., Rayner, A.C. (2018) 'A survey of dairy cow farmers in the United Kingdom: knowledge, attitudes and practices surrounding antimicrobial use and resistance.', *The Veterinary record*, 183(24), 746, available: http://ovidsp.ovid.com/ovidweb.cgi?T=JS&PAGE=reference&D=prem&NEWS=N&AN=30413678.

Horseman, S. V, Whay, H.R., Huxley, J.N., Bell, N.J., Mason, C.S. (2013) 'A survey of the on-farm treatment of sole ulcer and white line disease in dairy cattle.', *Veterinary journal (London, England : 1997)*, 197(2), 461–467, available: http://ovidsp.ovid.com/ovidweb.cgi?T=JS&PAGE=reference&D=med9&NEWS=N&AN=23602930.

Hyde, R.M., Remnant, J.G., Bradley, A.J., Breen, J.E., Hudson, C.D., Davies, P.L., Clarke, T., Critchell, Y., Hylands, M., Linton, E., Wood, E., Green, M.J. (2017) 'Quantitative analysis of antimicrobial use on British dairy farms.', *The Veterinary record*, [Comment in: Vet Rec. 2017 Dec 23;181(25):681-682; PMID: 29263291 [https://www.ncbi.nlm.nih.gov/pubmed/29263291]], 181(25), 683, available:

821 http://ovidsp.ovid.com/ovidweb.cgi?T=JS&PAGE=reference&D=med13&NEWS=N&AN=
822 29263292.

823 Jones-Diette, J.S., Brennan, M.L., Cobb, M., Doit, H., Dean, R.S. (2016) 'A method for
824 extracting electronic patient record data from practice management software systems
825 used in veterinary practice', *BMC Veterinary Research*, 12(1), 239, available:
826 https://doi.org/10.1186/s12917-016-0861-y.

827 Jones, J., Pearson, R., Jeckel, S. (2012) 'HELMINTH CONTROL Suspected anthelmintic
828 resistance to macrocyclic lactones in lambs in the UK', *VETERINARY RECORD*, 170(2),
829 59-U96.

830 Kamaludeen, J., Graham-Brown, J., Stephens, N., Miller, J., Howell, A., Beesley, N.J.,
831 Hodgkinson, J., Learmount, J., Williams, D. (2019) 'Lack of efficacy of triclabendazole
832 against Fasciola hepatica is present on sheep farms in three regions of England, and
833 Wales.', *The Veterinary record*, 184(16), 502, available:
834 http://ovidsp.ovid.com/ovidweb.cgi?T=JS&PAGE=reference&D=prem&NEWS=N&AN=3
835 0824600.

836 Kaminsky, R., Gauvry, N., Schorderet Weber, S., Skripsky, T., Bouvier, J., Wenger, A.,
837 Schroeder, F., Desaules, Y., Hotz, R., Goebel, T., Hosking, B.C., Pautrat, F., Wieland-
838 Berghausen, S., Ducray, P. (2008) 'Identification of the amino-acetonitrile derivative
839 monepantel (AAD 1566) as a new anthelmintic drug development candidate',
840 *Parasitology research*, 103(4), 931–939, available:
841 https://pubmed.ncbi.nlm.nih.gov/18594861.

842 Klous, G., Huss, A., Heederik, D.J.J., Coutinho, R.A. (2016) 'Human-livestock contacts and
843 their relationship to transmission of zoonotic pathogens, a systematic review of
844 literature', *One health (Amsterdam, Netherlands)*, 2, 65–76, available:
845 https://pubmed.ncbi.nlm.nih.gov/28616478.

846 Landers, T.F., Cohen, B., Wittum, T.E., Larson, E.L. (2012) 'A review of antibiotic use in food
847 animals: perspective, policy, and potential', *Public health reports (Washington, D.C. :
848 1974)*, 127(1), 4–22, available: https://www.ncbi.nlm.nih.gov/pubmed/22298919.

849 Learmount, J., Stephens, N., Boughtflower, V., Barrecheguren, A., Rickell, K. (2016) 'The
850 development of anthelmintic resistance with best practice control of nematodes on

commercial sheep farms in the UK.', *Veterinary parasitology*, 229, 9–14, available: http://ovidsp.ovid.com/ovidweb.cgi?T=JS&PAGE=reference&D=med12&NEWS=N&AN=27809985.

Lekagul, A., Tangcharoensathien, V., Yeung, S. (2019) 'Patterns of antibiotic use in global pig production: A systematic review', *Veterinary and Animal Science*, 7, 100058, available: http://www.sciencedirect.com/science/article/pii/S2451943X18302473.

Lima, E., Lovatt, F., Davies, P., Kaler, J. (2019) 'Using lamb sales data to investigate associations between implementation of disease preventive practices and sheep flock performance.', *Animal : an international journal of animal bioscience*, 1–9, available: http://ovidsp.ovid.com/ovidweb.cgi?T=JS&PAGE=reference&D=medp&NEWS=N&AN=31094306.

Little, P.R., Hodge, A., Maeder, S.J., Wirtherle, N.C., Nicholas, D.R., Cox, G.G., Conder, G.A. (2011) 'Efficacy of a combined oral formulation of derquantel-abamectin against the adult and larval stages of nematodes in sheep, including anthelmintic-resistant strains.', *Veterinary parasitology*, 181(2–4), 180–193, available: http://ovidsp.ovid.com/ovidweb.cgi?T=JS&PAGE=reference&D=med7&NEWS=N&AN=21684691.

MacFadyen, A.C., Fisher, E.A., Costa, B., Cullen, C., Paterson, G.K. (2018) 'Genome analysis of methicillin resistance in Macrococcus caseolyticus from dairy cattle in England and Wales.', *Microbial genomics*, 4(8), available: http://ovidsp.ovid.com/ovidweb.cgi?T=JS&PAGE=reference&D=prem&NEWS=N&AN=29916803.

Mahood, Q., Van Eerd, D., Irvin, E. (2014) 'Searching for grey literature for systematic reviews: Challenges and benefits', *Research Synthesis Methods*, 5(3), 221–234.

McArthur, C.L., Bartley, D.J., Shaw, D.J., Matthews, J.B. (2011) 'Assessment of ivermectin efficacy against gastrointestinal nematodes in cattle on four Scottish farms.', *The Veterinary record*, 169(25), 658, available: http://ovidsp.ovid.com/ovidweb.cgi?T=JS&PAGE=reference&D=med7&NEWS=N&AN=21984566.

Mellor, K.C., Petrovska, L., Thomson, N.R., Harris, K., Reid, S.W.J., Mather, A.E. (2019)

881 'Antimicrobial Resistance Diversity Suggestive of Distinct Salmonella Typhimurium
882 Sources or Selective Pressures in Food-Production Animals.', *Frontiers in microbiology*,
883 10, 708, available:
884 http://ovidsp.ovid.com/ovidweb.cgi?T=JS&PAGE=reference&D=prem&NEWS=N&AN=3
885 1031720.

886 Mills, H.L., Turner, A., Morgans, L., Massey, J., Schubert, H., Rees, G., Barrett, D., Dowsey, A.,
887 Reyher, K.K. (2018) 'Evaluation of metrics for benchmarking antimicrobial use in the UK
888 dairy industry.', *The Veterinary record*, [Comment in: Vet Rec. 2018 Dec
889 22;183(24):752; PMID: 30573584 [https://www.ncbi.nlm.nih.gov/pubmed/30573584]],
890 182(13), 379, available:
891 http://ovidsp.ovid.com/ovidweb.cgi?T=JS&PAGE=reference&D=medc&NEWS=N&AN=2
892 9476032.

893 Mitchell, E.S.E., Hunt, K.R., Wood, R., McLean, B. (2010) 'Anthelmintic resistance on sheep
894 farms in Wales.', *The Veterinary record*, 166(21), 650–652, available:
895 http://ovidsp.ovid.com/ovidweb.cgi?T=JS&PAGE=reference&D=med7&NEWS=N&AN=2
896 0495166.

897 Moher, D., Shamseer, L., Clarke, M., Ghersi, D., Liberati, A., Petticrew, M., Shekelle, P.,
898 Stewart, L.A., Group, P.-P. (2015) 'Preferred reporting items for systematic review and
899 meta-analysis protocols (PRISMA-P) 2015 statement', *Systematic Reviews*, 4(1), 1,
900 available: https://doi.org/10.1186/2046-4053-4-1.

901 Morgan, E.R., Hosking, B.C., Burston, S., Carder, K.M., Hyslop, A.C., Pritchard, L.J.,
902 Whitmarsh, A.K., Coles, G.C. (2012) 'A survey of helminth control practices on sheep
903 farms in Great Britain and Ireland.', *Veterinary journal (London, England : 1997)*,
904 [Comment in: Vet J. 2012 Jul;193(1):2-3; PMID: 22749119
905 [https://www.ncbi.nlm.nih.gov/pubmed/22749119]], 192(3), 390–397, available:
906 http://ovidsp.ovid.com/ovidweb.cgi?T=JS&PAGE=reference&D=med8&NEWS=N&AN=2
907 1908211.

908 Mphahlele, M., Molefe, N., Tsotetsi-Khambule, A., Oriel, T. (2019) 'Anthelmintic Resistance
909 in Livestock', in *Helminthiasis*, IntechOpen.

910 Mueller-Doblies, D., Speed, K.C.R., Kidd, S., Davies, R.H. (2018) 'Salmonella Typhimurium in

livestock in Great Britain - trends observed over a 32-year period.', *Epidemiology and infection*, 146(4), 409–422, available: http://ovidsp.ovid.com/ovidweb.cgi?T=JS&PAGE=reference&D=med13&NEWS=N&AN=29415790.

Nieuwhof, G.J., Bishop, S.C. (2005) 'Costs of the major endemic diseases of sheep in Great Britain and the potential benefits of reduction in disease impact', *Animal Science*, 81(1), 23–29, available: https://www.cambridge.org/core/article/costs-of-the-major-endemic-diseases-of-sheep-in-great-britain-and-the-potential-benefits-of-reduction-in-disease-impact/C1E2B560AB5FA568CECCAF6F8B23160A.

O'Kane, H., Ferguson, E., Kaler, J., Green, L. (2017) 'Associations between sheep farmer attitudes, beliefs, emotions and personality, and their barriers to uptake of best practice: The example of footrot.', *Preventive veterinary medicine*, 139(Pt B), 123–133, available: http://ovidsp.ovid.com/ovidweb.cgi?T=JS&PAGE=reference&D=med13&NEWS=N&AN=27371994.

O'Neill, J. (2016) *TACKLING DRUG-RESISTANT INFECTIONS GLOBALLY: FINAL REPORT AND RECOMMENDATIONS THE REVIEW ON ANTIMICROBIAL RESISTANCE CHAIRED BY JIM O'NEILL*, available: https://amr-review.org/sites/default/files/160518_Final paper_with cover.pdf [accessed 11 Jun 2019].

OIE (2016) AMR: OIE - World Organisation for Animal Health [online], available: https://www.oie.int/en/for-the-media/amr/ [accessed 19 May 2020].

Padiyara, P., Inoue, H., Sprenger, M. (2018) 'Global Governance Mechanisms to Address Antimicrobial Resistance', *Infectious diseases*, 11, 1178633718767887–1178633718767887, available: https://www.ncbi.nlm.nih.gov/pubmed/29686487.

Page, S.W., Gautier, P. (2012) 'Use of antimicrobial agents in livestock.', *Revue Scientifique et Technique - Office International des Épizooties*, 31(1), 145–188.

Paterson, G.K., Larsen, J., Harrison, E.M., Larsen, A.R., Morgan, F.J., Peacock, S.J., Parkhill, J., Zadoks, R.N., Holmes, M.A. (2012) 'First detection of livestock-associated meticillin-resistant Staphylococcus Aureus CC398 in bulk tank milk in the United Kingdom, January to July 2012', *Eurosurveillance*, 17(50), available:

941 https://www.scopus.com/inward/record.uri?eid=2-s2.0-
942 84871507033&partnerID=40&md5=6e15836f3790e1a697d80a06b8905d0e.

943 Paterson, G.K., Morgan, F.J.E., Harrison, E.M., Peacock, S.J., Parkhill, J., Zadoks, R.N., Holmes,
944 M.A. (2014) 'Prevalence and properties of mecC methicillin-resistant Staphylococcus
945 aureus (MRSA) in bovine bulk tank milk in Great Britain.', *The Journal of antimicrobial*
946 *chemotherapy*, 69(3), 598–602, available:
947 http://ovidsp.ovid.com/ovidweb.cgi?T=JS&PAGE=reference&D=med10&NEWS=N&AN=
948 24155057.

949 Piasecki, J., Waligora, M., Dranseika, V. (2018) 'Google Search as an Additional Source in
950 Systematic Reviews', *Science and Engineering Ethics*, 24(2), 809–810.

951 Randall, L., Heinrich, K., Horton, R., Brunton, L., Sharman, M., Bailey-Horne, V., Sharma, M.,
952 McLaren, I., Coldham, N., Teale, C., Jones, J. (2014) 'Detection of antibiotic residues and
953 association of cefquinome residues with the occurrence of Extended-Spectrum beta-
954 Lactamase (ESBL)-producing bacteria in waste milk samples from dairy farms in England
955 and Wales in 2011.', *Research in veterinary science*, 96(1), 15–24, available:
956 http://ovidsp.ovid.com/ovidweb.cgi?T=JS&PAGE=reference&D=med10&NEWS=N&AN=
957 24314891.

958 RODRIGUEZ, A., PANGLOLI, P., RICHARDS, H.A., MOUNT, J.R., DRAUGHON, F.A.N.N. (2006)
959 'Prevalence of Salmonella in Diverse Environmental Farm Samples', *Journal of Food*
960 *Protection*, 69(11), 2576–2580, available: https://doi.org/10.4315/0362-028X-
961 69.11.2576.

962 Rossi, G., De Leo, G.A., Pongolini, S., Natalini, S., Zarenghi, L., Ricchi, M., Bolzoni, L. (2017)
963 'The Potential Role of Direct and Indirect Contacts on Infection Spread in Dairy Farm
964 Networks', *PLOS Computational Biology*, 13(1), e1005301, available:
965 https://doi.org/10.1371/journal.pcbi.1005301.

966 RUMA (2019) *Targets Task Force: Two Years On*, available: https://www.ruma.org.uk/wp-
967 content/uploads/2019/10/RUMA-TTF-update-2019-two-years-on-FULL-REPORT.pdf
968 [accessed 3 Mar 2020].

969 Rutherford, S.-J., Jeckel, S., Ridler, A. (2015) 'Characteristics of sheep flocks affected by
970 Streptococcus dysgalactiae arthritis.', *The Veterinary record*, 176(17), 435, available:

971 http://ovidsp.ovid.com/ovidweb.cgi?T=JS&PAGE=reference&D=med11&NEWS=N&AN=
972 25724543.

973 SCOPS (2019) SCOPS | Sustainable Control of Parasites in Sheep [online], available:
974 https://www.scops.org.uk/ [accessed 14 May 2020].

975 Scott, L., Menzies, P., Reid-Smith, R.J., Avery, B.P., McEwen, S.A., Moon, C.S., Berke, O.
976 (2012) 'Antimicrobial resistance in fecal generic Escherichia coli and Salmonella spp.
977 obtained from Ontario sheep flocks and associations between antimicrobial use and
978 resistance', *Canadian journal of veterinary research = Revue canadienne de recherche*
979 *veterinaire*, 76(2), 109–119, available: https://pubmed.ncbi.nlm.nih.gov/23024453.

980 Shalaby, H.A. (2013) 'Anthelmintics Resistance; How to Overcome it?', *Iranian journal of*
981 *parasitology*, 8(1), 18–32, available: https://www.ncbi.nlm.nih.gov/pubmed/23682256.

982 Stubbings, L., SCOPS (2012) 'Efficacy of macrocyclic lactone treatments in sheep in the UK.',
983 *The Veterinary record*, 170(25), 653, available:
984 http://ovidsp.ovid.com/ovidweb.cgi?T=JS&PAGE=reference&D=med8&NEWS=N&AN=2
985 2730501.

986 Sutherland, I.A., Leathwick, D.M. (2011) 'Anthelmintic resistance in nematode parasites of
987 cattle: a global issue?', *Trends in Parasitology*, 27(4), 176–181, available:
988 http://www.sciencedirect.com/science/article/pii/S1471492210002370.

989 Talan, D.A., Takhar, S.S., Krishnadasan, A., Abrahamian, F.M., Mower, W.R., Moran, G.J.,
990 Group, Emerge.I.D.N.S. (2016) 'Fluoroquinolone-Resistant and Extended-Spectrum β-
991 Lactamase-Producing Escherichia coli Infections in Patients with Pyelonephritis, United
992 States(1)', *Emerging infectious diseases*, 22(9), 1594–1603, available:
993 https://pubmed.ncbi.nlm.nih.gov/27532362.

994 Tang, K.L., Caffrey, N.P., Nóbrega, D.B., Cork, S.C., Ronksley, P.E., Barkema, H.W., Polachek,
995 A.J., Ganshorn, H., Sharma, N., Kellner, J.D., Ghali, W.A. (2017) 'Restricting the use of
996 antibiotics in food-producing animals and its associations with antibiotic resistance in
997 food-producing animals and human beings: a systematic review and meta-analysis',
998 *The Lancet. Planetary health*, 1(8), e316–e327, available:
999 https://www.ncbi.nlm.nih.gov/pubmed/29387833.

1000 Taylor, M.A., Learmount, J., Lunn, E., Morgan, C., Craig, B.H. (2009) 'Multiple resistance to
1001 anthelmintics in sheep nematodes and comparison of methods used for their
1002 detection', *Small Ruminant Research*, 86(1–3), 67–70, available:
1003 https://www.scopus.com/inward/record.uri?eid=2-s2.0-
1004 70450223329&doi=10.1016%2Fj.smallrumres.2009.09.020&partnerID=40&md5=457e8
1005 32e7783938331e22fedf113a4f2.

1006 Thomas, E. (2015) 'Anthelmintic study update – results of the recent WAARD project',
1007 *CATTLE PRACTICE*, 23(2), 377–378.

1008 Thomas, V., De Jong, A., Moyaert, H., Simjee, S., El Garch, F., Morrissey, I., Marion, H., Vallé,
1009 M. (2015) 'Antimicrobial susceptibility monitoring of mastitis pathogens isolated from
1010 acute cases of clinical mastitis in dairy cows across Europe: VetPath results',
1011 *International journal of antimicrobial agents*, 46(1), 13–20, available:
1012 https://www.scopus.com/inward/record.uri?eid=2-s2.0-
1013 84934921255&doi=10.1016%2Fj.ijantimicag.2015.03.013&partnerID=40&md5=948b82
1014 8f582b490d2c9e252da26cfd63.

1015 Træholt Franck, K., Olsen, S., Sönksen, U. (2017) 'DANMAP 2016, RESISTANCE IN HUMAN
1016 CLINICAL BACTERIA, E. coli and K. pneumoniae'.

1017 Velasova, M., Smith, R.P., Lemma, F., Horton, R.A., Duggett, N.A., Evans, J., Tongue, S.C.,
1018 Anjum, M.F., Randall, L.P. (2019) 'Detection of extended-spectrum beta-lactam, AmpC
1019 and carbapenem resistance in Enterobacteriaceae in beef cattle in Great Britain in
1020 2015.', *Journal of applied microbiology*, 126(4), 1081–1095, available:
1021 http://ovidsp.ovid.com/ovidweb.cgi?T=JS&PAGE=reference&D=medl&NEWS=N&AN=3
1022 0693606.

1023 Veldman, K., Wit, B., Pelt, W., Heederik, D., DJ, M. (2017) *MARAN 2017: Monitoring of*
1024 *Antimicrobial Resistance and Antibiotic Usage in Animals in the Netherlands in 2016.*
1025 *Combined with NETHMAP-2017: Consumption of Antimicrobial Agents and*
1026 *Antimicrobial Resistance among Medically Important Bacteria in the Netherlan*.

1027 VMD (2019a) UK One Health Report: Antibiotic Use and Antibiotic Resistance in Animals and
1028 Humans - GOV.UK [online], available:
1029 https://www.gov.uk/government/publications/uk-one-health-report-antibiotic-use-

and-antibiotic-resistance-in-animals-and-humans [accessed 18 May 2020].

VMD (2019b) *UK Veterinary Antibiotic Resistance and Sales Surveillance Report Www.Gov.Uk/Government/Organisations/Veterinary-Medicines-Directorate*, available: www.nationalarchives.gov.uk/doc/open-government-licence/version/3/oremailPSI@nationalarchives.gov.uk.Thispublicationisavailableatwww.gov.uk/government/collections/veterinary-antimicrobial-resistance-and-sales-surveillance. [accessed 3 Mar 2020].

Warner, R.G., Snow, L.C., Cheney, T., Wearing, H., Harris, K., Cook, A.J., Teale, C.J., Coldham, N.G. (2011) 'Identification of risk factors for the prevalence of CTX-M ESBL E. coli on dairy farms in North West England and North Wales', *Cattle Practice*, 19(1), 51, available: https://www.scopus.com/inward/record.uri?eid=2-s2.0-79953712603&partnerID=40&md5=4c304c3b9a8f1649b957eb0f186fe665.

WHO (2019) 'WHO | WHO list of critically important antimicrobials (WHO CIA list)', *WHO*, available: https://www.who.int/foodsafety/areas_work/antimicrobial-resistance/cia/en/ [accessed 18 May 2020].

Woolhouse, M., Ward, M., Bunnik, B. van, Farrar, J. (2015) 'Antimicrobial resistance in humans, livestock and the wider environment', *Philosophical transactions of the Royal Society of London. Series B, Biological sciences*, 370(1670).

Wu, G., Ehricht, R., Mafura, M., Stokes, M., Smith, N., Pritchard, G.C., Woodward, M.J. (2012) 'Escherichia coli isolates from extraintestinal organs of livestock animals harbour diverse virulence genes and belong to multiple genetic lineages.', *Veterinary microbiology*, 160(1–2), 197–206, available: http://ovidsp.ovid.com/ovidweb.cgi?T=JS&PAGE=reference&D=med8&NEWS=N&AN=22766078.

Wu, Z., Sippy, R., Sahin, O., Plummer, P., Vidal, A., Newell, D., Zhang, Q. (2014) 'Genetic diversity and antimicrobial susceptibility of Campylobacter jejuni isolates associated with sheep abortion in the United States and Great Britain.', *Journal of clinical microbiology*, 52(6), 1853–1861, available: http://ovidsp.ovid.com/ovidweb.cgi?T=JS&PAGE=reference&D=medc&NEWS=N&AN=24648552.

Zaytoun, O.M., Vargo, E.H., Rajan, R., Berglund, R., Gordon, S., Jones, J.S. (2011) 'Emergence of Fluoroquinolone-resistant Escherichia coli as Cause of Postprostate Biopsy Infection: Implications for Prophylaxis and Treatment', *Urology*, 77(5), 1035–1041, available: http://www.sciencedirect.com/science/article/pii/S0090429511000513.