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REVIEW & INTERPRETATION

Benefits of Condensed Tannins in Forage Legumes Fed to Ruminants: Importance of Structure, Concentration, and Diet Composition

Irene Mueller-Harvey,* Giuseppe Bee, Frigga Dohme-Meier, Hervé Hoste, Maarit Karonen, Roland Kölliker, Andreas Lüscher, Vincent Niderkorn, Wilbert F. Pellikaan, Juha-Pekka Salminen, Leif Skøt, Lydia M.J. Smith, Stig M. Thamsborg, Paul Totterdell, Ian Wilkinson, Andrew R. Williams, Blasius N. Azuhnwi, Nicolas Baert, Anja Grosse Brinkhaus, Giuseppe Copani, Olivier Desrues, Chris Drake, Marica Engström, Christos Fryganas, Marion Girard, Nguyen T. Huyen, Katharina Kempf, Carsten Malisch, Marina Mora-Ortiz, Jessica Quijada, Aina Ramsay, Honorata M. Ropiak, and Garry C. Waghorn

ABSTRACT

Condensed tannins (CTs) account for up to 20% of the dry matter in forage legumes used as ruminant feeds. Beneficial animal responses to CTs have included improved growth, milk and wool production, fertility, and reduced methane emissions and ammonia volatilization from dung or urine. Most important is the ability of such forages to combat the effects of gastrointestinal parasitic nematodes. Inconsistent animal responses to CTs were initially attributed to concentration in the diet, but recent research has highlighted the importance of their molecular structures, as well as concentration, and also the composition of the diet containing the CTs. The importance of CT structural traits cannot be underestimated. Interdisciplinary research is the key to unraveling the relationships between CT traits and bioactivities and will enable future on-farm exploitation of these natural plant compounds. Research is also needed to provide plant breeders with guidelines and screening tools to optimize CT traits, in both the forage and the whole diet. In addition, improvements are needed in the competitiveness and agronomic traits of CT-containing legumes and our understanding of options for their inclusion in ruminant diets. Farmers need varieties that are competitive in mixed swards and have predictable bioactivities. This review covers recent results from multidisciplinary research on sainfoin (*Onobrychis* Mill. spp.) and provides an overview of current developments with several other tanniniferous forages. Tannin chemistry is now being linked with agronomy, plant breeding, animal nutrition, and parasitology. The past decade has yielded considerable progress but also generated more questions—an enviable consequence of new knowledge!

I. Mueller-Harvey, C. Drake, C. Fryganas, A. Ramsay, and H.M. Ropiak, School of Agriculture, Policy and Development, Univ. of Reading, PO Box 236, Reading RG6 6AT, UK; G. Bee, F. Dohme-Meier, A.G. Brinkhaus, and M. Girard, Agroscope-Posieux, CP 64, 1725 Posieux, Switzerland; H. Hoste, B.N. Azuhnwi, and J. Quijada, INRA, UMR 1225 IHAP, BP 87614, Toulouse 31076, France; M. Karonen, J.-P. Salminen, N. Baert, and M. Engström, Dep. of Chemistry, Univ. of Turku, 20014 Turku, Finland; R. Kölliker, Molecular Plant Breeding, ETH Zürich, 8092 Zürich, Switzerland; A. Lüscher and C. Malisch, Forage Production and Grassland Systems, Agroscope, 8046 Zürich, Switzerland; V. Niderkorn and G. Copani, INRA UMR1213 Herbivores, 63122 St-Genès-Champanelle, France; W.F. Pellikaan and N.T. Huyen, Wageningen Univ. and Research, Animal Nutrition Group, PO Box 338, 6700 AH Wageningen, the Netherlands; L. Skøt, IBERS, Aberystwyth SY23 3EB, UK; L.M.J. Smith and M. Mora-Ortiz, NIAB, Huntingdon Rd., Cambridge CB3 0LE, UK; S.M. Thamsborg, A.R. Williams, and O. Desrues, Dep. of Veterinary Disease Biology, Univ. of Copenhagen, Dyrlægevej 100, Frederiksberg C 1870, Denmark; P. Totterdell and I. Wilkinson, Cotswold Seeds, London Rd., Moreton-in-Marsh GL56 0JQ, UK; K. Kempf, Molecular Ecology, Agroscope, 8046 Zürich, Switzerland; G.C. Waghorn, Institute of Veterinary and Biomedical Sciences, Massey Univ., Private Bag 11222, Palmerston North 4474, New Zealand. Received 19 July 2017. Accepted 18 Oct. 2017. *Corresponding author (i.mueller-harvey@reading.ac.uk). Assigned to Associate Editor Jose Dubeux.

Abbreviations: BSA, bovine serum albumin; CT, condensed tannin; mDP, mean degree of polymerization; MYB, myeloblastosis; NIRS, near-infrared reflectance spectroscopy; PC, procyanidin; PD, prodelphinidin; PEG, polyethylene glycol; RuBisCo, ribulose-1,5-bisphosphate carboxylase/oxygenase; SSR, simple sequence repeat.

Despite the widespread occurrence of condensed tannins (CTs) in the plant kingdom (Fig. 1), there are still large gaps in our knowledge that continue to challenge plant breeding, animal science, and analytical chemistry; progress in all of these disciplines is needed in order understand the mechanisms that underpin their actions and to fully exploit their benefits.

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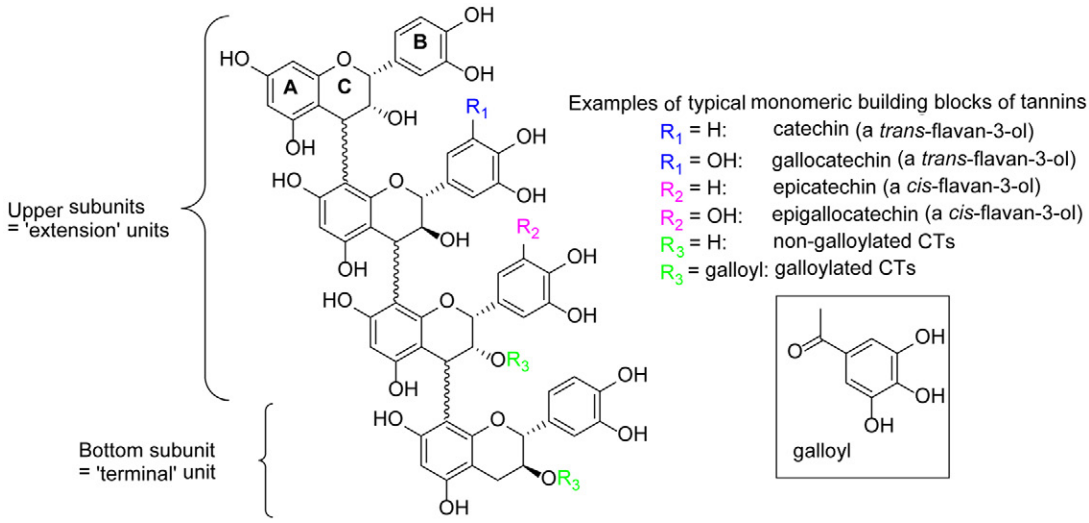

Fig. 1. Example of a condensed tannin (CT) molecule that consists of four flavan-3-ol subunits (= monomeric building blocks). Procyanidins are composed of catechin or epicatechin and prodelphinidins of gallocatechin or epigallocatechin subunits (for further details, see Zeller, 2019).

This review focuses on progress achieved during the past decade and considers the following topics:

1. bioactive CTs in plants in general and forage legumes in particular;
2. intra- and interspecies variations in CT contents and composition, which will be termed CT traits from here on;
3. effects of agronomic, harvesting, and processing practices on CT efficacies;
4. effects on animal health, nutrition, product quality, and environmental emissions; and
5. characteristics useful for plant breeders and tools for selecting or engineering forages with novel CT traits.

This review also summarizes results from a multidisciplinary research consortium that focused on sainfoin (Fig. 2). In this project, agronomists and plant breeders assembled germplasm collections of sainfoin (*Onobrychis* Mill. spp.), which is a traditional forage legume in Europe, and identified molecular markers and strategies for weed control. Ruminant nutritionists studied various accessions for their in vitro fermentation characteristics and in vivo feeding trials, nitrogen balances, and the quality of meat and dairy products. Parasitologists explored the antiparasitic properties of a wide range of CT traits, and chemists developed tools for analyzing the CTs in sainfoin pellets, silages, and digesta.

The background to this work was the pioneering research in New Zealand that first studied fresh sainfoin (*Onobrychis viciifolia* Scop.) and *Lotus* L. spp. in relation to legume bloat and nutritive value (Reid et al., 1974). Sainfoin research included measurements of the protein-binding characteristics of CTs (Fig. 1) (Jones and Mangan, 1977) and aspects of nitrogen digestion in sheep (Egan and Ulyatt, 1980), but problems with sainfoin persistence in swards diverted attention to *Lotus* species. Initial research by Barry and colleagues focused on big trefoil (*Lotus pedunculatus* Cav.), examining effects of CT concentration (Barry et al., 1986), whereas Waghorn et al. (1987) demonstrated the beneficial effects of CTs in birdsfoot trefoil (*Lotus corniculatus* L. var. *corniculatus*) on absorption of essential amino acids from the intestine. Other forages were also evaluated in New Zealand, including sulla (*Hedysarum coronarium* L.; Stienezen et al., 1996) and dock (*Rumex obtusifolius* L.; Waghorn and Jones, 1989), but the importance of CT composition, in addition to concentration, was demonstrated more recently (Waghorn et al., 1997). Subsequent research included CT effects on livestock parasites and greenhouse gas emissions, but financial support decreased because of a lack of competitiveness of tanniniferous forages, especially in fertile soils. It also became apparent that detailed chemical characterization was required to elucidate mechanisms of action, because the two *Lotus* species differed in their biological effects and tannin types. The question was: were the CTs responsible for these different biological effects?

This review seeks to provoke discussion on how to progress this area of research, how to optimize the

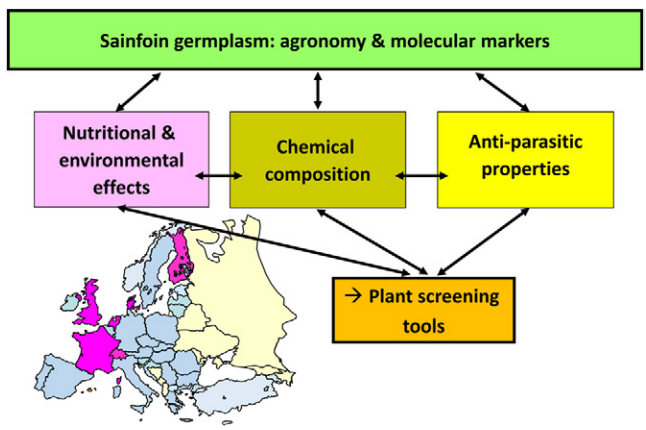

Fig. 2. The LegumePlus project—an interdisciplinary European Union-funded research and training network on sainfoin (http://legumeplus.eu).

bioactivity of CT forages, and how to develop on-farm applications. It will also consider whether farmers might require plants with different CT traits for either antiparasitic or nutritional purposes. The reader is also referred to the accompanying article by Zeller (2019) for a detailed description of CT structures and the review by Tedeschi et al. (2014) for a mechanistic model describing the overall interactions between CTs and ruminants.

ROLES OF TANNINS IN PLANTS AND CHALLENGES TO HARNESSING THEIR BENEFITS FOR LIVESTOCK PRODUCTION

The reader of the tannin literature is repeatedly reminded that tannins are "secondary" plant metabolites and provide a defense against herbivory (Barbehenn and Constabel, 2011; Agrawal et al., 2012; Lattanzio et al., 2012). However, herbivores comprise a range of species, from insects to ruminants, and have distinctly different gut systems. Although tannins can account for antiherbivory effects in insects (Salminen and Karonen, 2011), ruminant behavior suggests that their herbivory effect is marginal because forages containing CTs are consumed, and selection is often in preference to grasses (Waghorn, 2008). Within plants, leaves are selected in preference to stems despite higher CT concentrations, even when CTs accounted for ~20% of the dry matter in erect canary clover [*Dorycnium rectum* (L.) Ser.] leaf (Waghorn and Molan, 2001). Current thinking suggests that these secondary metabolites provide plants with a plasticity that can support their development and interaction with the environment (Bidel et al., 2010; Neilson et al., 2013; Mouradov and Spangenberg, 2014). This concept, that CTs may have multiple and interrelated functions, is now gaining traction. For example, Chen et al. (2014) showed that the temperatures experienced by the mother plant elicit CT pathways that can pass information to the next generation. In addition, the flavan-3-ol monomers, which are precursors of CTs, appear to play a vital role in protecting chromosomes during periods of high cell activity, but not during dormancy or drought stress (Feucht et al., 2013). It is important to appreciate that a multitude of different CT compounds exist (Khanbabaee and van Ree, 2001; Hümmer and Schreier, 2008; Salminen and Karonen, 2011; Zeller, 2019), and that their synthesis in the plant kingdom has not converged on a single structure. These findings suggest that the function of CTs deserves a closer look in plant and crop science.

From an animal's perspective, when dietary CT concentrations are too high, or protein concentrations are too low, as in tropical environments where grasses may have little nitrogen and tree leaves may have high CT concentrations, CTs can be antinutritional (Cooper et al., 1988). The benefits of CTs have been demonstrated in only a few CT-containing feeds with ruminant animals (e.g., sheep, goats, and cattle; Mueller-Harvey, 2006; Waghorn, 2008). The nutritional benefits include improved growth, milk yields, fertility, and tolerance to some intestinal parasites and arise from protection of dietary protein from excessive fermentation in the rumen. Other benefits include bloat prevention, which is associated with tannins reducing the stability of a foam that traps ruminal fermentation gases, and antiparasitic effects against ruminant and nonruminant gastrointestinal parasites (Waghorn, 2008; Kingston-Smith et al., 2010; Terrill et al., 2012; Wang et al., 2012; Hoste et al., 2015, 2016; MacAdam and Villalba, 2015).

Although plants synthesize many different tannin types, this review will focus on CTs, as these are of particular interest in forage legumes and several other pasture plants. Hydrolyzable tannins are not considered here, although evidence is emerging that some may exert similarly useful bioactive effects (Baert et al., 2016; Engström et al., 2016; Bee et al., 2017).

A major impetus for research and utilization of CT forage legumes by livestock producers has been the drive to reduce bloat, to improve farm profitability, to control parasites, and to reduce greenhouse gas and ammonia emissions (Kingston-Smith et al., 2010; Wang et al., 2012; McCaslin et al., 2014; Hoste et al., 2015). Nutritional responses to CTs have been variable, and this has led to contradictory reports about their benefits (Min et al., 2003; Mueller-Harvey, 2006; Waghorn, 2008). This is not surprising given the complexity of plant CTs, their impact when forages are fed as a sole diet or as a dietary component, and their interactions with feed components, host tissues, and the microbiome, plus the effects stemming from the animal's nutrient requirements and parasitism. A concerted multidisciplinary research approach is required to harness the full potential of CTs for livestock production (Mueller-Harvey, 2006; Waghorn, 2008). However, obtaining funding for such a wide-ranging set of topics has been challenging and reflects common barriers to interdisciplinary research, such as narrowly focused funding goals and short funding timelines. Progress in the study of complex plant–livestock systems is inherently slow, and CTs also pose interesting analytical and experimental challenges. Taken together, these facts account for the relatively slow progress in identifying the relationships between CT traits and bioactivities.

CHALLENGES AND OPPORTUNITIES POSED BY PLANTS WITH DIFFERENT CONDENSED TANNIN TYPES

Research on plant CTs has come a long way since the early literature described these compounds of mysterious composition and function as "accidents of [plant] metabolism" or metabolic "waste products" that served to

support the primary plant metabolism (Haslam, 1981). It is now well established that CT synthesis is under genetic control (Szczyglowski and Stougaard, 2008; Scioneaux et al., 2011; Cheynier et al., 2013; Escaray et al., 2014) and that expression depends on the plant species and plant parts (Larkin et al., 1997; Gebrehiwot et al., 2002; Abeynayake et al., 2012; Ferreyra et al., 2012; Hancock et al., 2012; Verdier et al., 2012; Cheynier et al., 2013; Harding et al., 2013; Mouradov and Spangenberg, 2014; Pérez-Díaz et al., 2014; Zhou et al., 2015; Zhu et al., 2015; Chezem and Clay, 2016).

Chemotaxonomic surveys on the distribution of CTs in plants in general and of forage legumes in particular have found that CT compositions tend to follow distinct biosynthetic patterns in terms of their flavan-3-ol subunit composition and polymer sizes, which are described in terms of mean degree of polymerization (mDP). Plants with procyanidin (PC)-type CTs are much more widespread than plants with prodelphinidin (PD)-type CTs (Fig. 1), but many more plant species contain PC/PD mixtures (Porter, 1988; Mueller-Harvey, 2006; Sivakumaran et al., 2006; Mechineni et al., 2014; Laaksonen et al., 2015; Quijada et al., 2015; Hoste et al., 2016; Ropiak et al., 2016b). Most plant CTs have *cis*-flavan-3-ol subunits, especially as extension units (Fig. 1), whereas CTs with predominantly *trans*-flavan-3-ol subunits in extension units are relatively rare (Porter, 1988; Hernes and Hedges, 2004; Klongsiriwet et al., 2013). We have also observed other trends in the composition of CTs in forage legumes, which are illustrated with examples from a few plants that are "specialists" in producing particular CT types:

- CTs that comprise PCs tend to be mixtures of oligomers and smaller polymers (e.g., cocoa [*Theobroma cacao* L.] bean CTs with mDP values of 2–5).

- CTs that comprise PDs are usually mixtures of larger polymers (e.g., sericea lespedeza [*Lespedeza cuneata* (Dum. Cours.) G. Don] CTs with mDP values of 10–30).

However, exceptions exist, as lime tree flowers (*Tilia* L. spp.) and some varieties of cider apples (*Malus domestica* Borkh.) have PCs with higher mDP values of 8 and ~100, respectively (Guyot et al., 2001a; Ropiak et al., 2017).

Other sources of special CT types include:

- Leaves from several willow (*Salix* L. spp.) accessions and black currant (*Ribes nigrum* L.) have high proportions of PCs and PDs with *trans*-flavan-3-ol subunits, respectively (Porter, 1988).

- Tea [*Camellia sinensis* (L.) Kuntze] leaves and shea (*Vitellaria paradoxa* C. F. Gaertn.) nuts are unusual in having high proportions of galloylated flavan-3-ol monomers and galloylated low molecular weight PDs (Henning et al., 2003; Ramsay et al., 2016).

- Water dock (*Rumex hydrolapathum* Huds.) roots and persimmon (*Diospyros kaki* Thunb.) fruits contain highly galloylated smaller PCs (mDP = 6; galloylation = 52%; Ropiak et al., 2016b) and larger PDs (mDP = 26; galloylation = 72%; Li et al., 2010), respectively.

- An entire series of oligomeric and polymeric PC xylosides are present in birch (*Betula pendula* Roth) bark (Liimatainen et al., 2012).

Not surprisingly, these biosynthetic patterns can generate contradictory and confounding effects when attempting to align CT structure with bioactivity (Laaksonen et al., 2015; Hixson et al., 2016). However, the particular CTs in these 'specialist' plants can provide unique opportunities for research, because it would be very difficult to separate sufficient quantities of a particular CT type from the complex CT mixtures, which are typical of most plants, for laboratory or in vitro studies. This problem is illustrated by the CT mixtures in different sainfoin, sericea lespedeza and *Lotus* accessions (Table 1), in which the PC/PD ratios ranged from 84:16 to 3:97, *cis*-/*trans*-flavan-3-ol ratios from 90:10 to 66:34, and mDP values from of 12 to 84 (Meagher et al., 2004; Stringano et al., 2012; Azuhnwi et al., 2013a; Mechineni et al., 2014).

An alternative approach is to use CTs from "CT specialist plants." The already "pure" groups of either PCs or PDs, having either *cis*- or *trans*-flavan-3-ol stereochemistry, can be isolated from these specialist plants, separated in the laboratory into mDP variants, and used to explore the bioactivities of different PC/PD ratios, *cis*-/*trans*-flavan-3-ol ratios, and polymer sizes (Brown et al., 2017). Currently, this is the most straightforward approach to structure-activity studies, because chemical synthesis of CTs is even more challenging.

Another aspect of CT composition that is poorly researched concerns the galloylated CTs. This is a group of particularly potent antioxidants (Fig. 1; Li et al., 2010) that appear to possess strong antiparasitic, nematocidal, and antimicrobial activities but have received little attention for their nutritional or health effects (Brunet and Hoste 2006; Ropiak et al., 2016a). *Acacia nilotica* (L.) Delile leaves, carob (*Ceratonia siliqua* L.) pods, grape (*Vitis vinifera* L.) seeds, persimmon fruits, lentisk (*Pistacia lentiscus* L.) leaves, shea nuts, tea leaves, and *Rumex* L. spp. are good sources of galloylated CTs (Self et al., 1986; Henning et al., 2003; Papagiannopoulos et al., 2004; Spencer et al., 2007; Li et al., 2010; Rodríguez-Pérez et al., 2013; Derksen et

al., 2014; Ramsay et al., 2016; Ropiak et al., 2016b; our unpublished data).

PROGRESS IN TANNIN ANALYSIS

Several new techniques have been developed recently for analyzing CT mixtures. As Zeller (2019) has addressed this topic in detail, only a few additional techniques are described below. An important constraint in the quest for valid tannin data is the requirement for high purity standards for quantitation, which means that the CT concentration and purity of the standards needs to be assessed by CT-specific methods such as thiolysis (Gea et al., 2011; Grabber et al., 2013; Williams et al., 2014a) or nuclear magnetic resonance spectroscopy (Zeller et al., 2015a). We emphasize that the widely used elution of plant extracts with 70% aqueous acetone from Sephadex LH-20 columns for tannin "purification" can lead to CT concentrations as low as ~13 g CTs 100 g^{-1} "purified sample" (Williams et al., 2014b). Their use would overestimate CT concentrations, so additional steps are required to increase their purity (Stringano, 2011; Fryganas, 2016; Brown et al., 2017; Ropiak et al., 2017). It is also essential to use CT mixtures that are specific to the plant species being investigated, because CT composition affects ultraviolet-visible absorption maxima and reaction yields that result from the hydrochloric acid (HCl)-butanol-acetone and thiolysis assays (Grabber et al., 2013; Krueger et al., 2013; Engström et al., 2014; Hixson et al., 2015; Ropiak et al., 2016b; Wang et al., 2016). We emphasize the unsuitability of commercially available CTs from quebracho [*Schinopsis quebracho-colorado* (Schltdl.) F. A. Barkley & T. Mey.] as a standard because these have 5-deoxy-flavan-3-ol subunits, which give particularly low reaction yields that lead to overestimation of CT concentrations (Schofield et al., 2001; Rautio et al., 2007), and tannic acid is even less appropriate, as it contains none of the CT flavan-3-ol subunits.

Additional challenges include variation in extractability of CTs. Some can be extracted with water or aqueous methanol, and others require aqueous acetone, but many CTs are tightly bound to the plant matrix and cannot be extracted with these solvents. By using techniques that only measure the easily extractable CTs, researchers may risk missing a large fraction (Table 2) that

Table 1. Condensed tannin (CT) concentrations (g/100 g dry matter) and compositions of forage legumes and selected pasture plants (Note: these studies used various CT assays that were applied either to whole plants, isolated extracts, or purified CT fractions).

Plant species	CT concentration	PC/PD ratio†	mDP‡	References
	g 100 g^{-1} dry matter			
Erect canary clover [*Dorycnium rectum* (L.) Ser.]	15–20	5:95–17:83	10–127	Sivakumaran et al., 2004
Sericea lespedeza [*Lespedeza cuneata* (Dum. Cours.) G. Don]	6–13	3:97	33	Mechineni et al., 2014
Panicledleaf ticktrefoil [*Desmodium paniculatum* (L.) DC.]	22	nd§	nd	Pawelek et al., 2008
Sulla (*Hedysarum coronarium* L.)	2–12	11:89–27:73	3–46	Tibe et al., 2011
Big trefoil (*Lotus pedunculatus* Cav.)	5–10	16:84–20:80	2–44	Meagher et al., 2004; Sivakumaran et al., 2006
Sainfoin (*Onobrychis viciifolia* Scop.)	1–9	5:95–50:50	12–84	Berard et al., 2011; Stringano et al., 2012; Azuhnwi et al., 2013a; Malisch et al., 2015
Prairie clover (*Dalea purpurea* Vent.)	4–9	nd	nd	Berard et al., 2011
Birdsfoot trefoil (*Lotus corniculatus* L. var. *corniculatus*)	0–5	60:40– 84:16	9	Meagher et al., 2004; Sivakumaran et al., 2006; Berard et al., 2011; Grabber et al., 2014

† PC/PD, procyanidin:prodelphinidin ratio.

‡ mDP, mean degree of polymerization.

§ nd, not determined.

Table 2. Overview of techniques for determining extractable or unextractable condensed tannins (CTs) and their composition.

Analytical technique†	Extractable CTs	Unextractable CTs	Information on CT composition	Literature
HCl-butanol-acetone	X	X	Limited	Grabber et al., 2013
Thiolysis, phloroglucinolysis	X	X	X	Guyot et al., 2001b; Gea et al., 2011; Hixson et al., 2015, 2016; Ramsay et al., 2016
NMR	X	X	X	Grabber et al., 2013; Zeller et al., 2015a
NIRS	X	X	X	Petersen et al., 1991; Larkin et al., 1997; Dykes et al., 2014; Grabber et al., 2014; Klongsiriwet 2016
MALDI-TOF MS	X	–	X	Stringano et al., 2011; Feliciano et al., 2012; Krueger et al., 2013
UPLC-MS/MS	X	–	X	Engström et al., 2014

† NMR, nuclear magnetic resonance; NIRS, near-infrared reflectance spectroscopy; MALDI TOF MS, matrix-assisted laser desorption ionization–time-of-flight mass spectrometry; UPLC-MS/MS, ultra-performance liquid chromatography tandem mass spectrometry (for further information, see Zeller, 2019).

may have potentially important bioactivities (Gea et al., 2011; Pérez-Jiménez and Lluís Torres 2011; Cheynier et al., 2015; Hixson et al., 2016; our unpublished data). The impact of extractable vs. unextractable CTs on ruminant nutrition and health has not yet been determined.

Thiolysis or phloroglucinolysis (analytical degradation of CTs with thiols or phloroglucinol) can be used to determine the composition of CTs by depolymerization, enabling characterization of the flavan-3-ol subunits (Zeller, 2019). The use of thiolysis to analyze CTs in whole plant material (removing the need for extraction) was first reported by Guyot et al. (2001b) for apple residues and then adapted to sainfoin and food samples (Hellström and Mattila, 2008; Gea et al., 2011). However, quantification remains problematic, and higher CT yields have been reported in some samples with the HCl-butanol-acetone assay than with the thiolysis or phloroglucinolysis assays. Low values have been linked to oxidative processes affecting reaction yields (Hixson et al., 2015; Klongsiriwet, 2016; Brillouet et al., 2017; Desrues et al., 2017).

However, thiolysis can also generate up to threefold higher yields than the HCl-butanol-acetone assay (Drake and Mueller-Harvey, unpublished data, 2016); these particular CTs had high degrees of galloylation (Fig. 1), and this demonstrates the variation in reactivity of CTs and yield of assayable end products. These inconsistencies illustrate the challenge of determining the amount and type of CTs in forage material and reinforce the case for using more than one method for analyzing CTs, and for using plant-specific CT standards (see above). Recent observations also suggest that the optimum time for the in situ thiolysis is affected by the plant species. Some samples yielded more consistent CT parameters if thiolysis was performed for 2 to 3 h rather than 1 h, as suggested by Gea et al. (2011). Clearly, there is no one silver bullet for CT analysis, and methods need to be evaluated and adapted for a particular plant species and research objective.

Infrared spectroscopy techniques are of particular interest to plant breeders due to their speed and suitability for screening large numbers of samples; near-infrared reflectance spectroscopy (NIRS) can be used in the laboratory, whereas visible–near-infrared and shortwave-infrared spectroscopy have been used for field screening (Lehmann et al., 2015). Near-infrared reflectance spectroscopy has potential for measuring not only CT concentrations but also PC/PD and *cis-*/*trans*-flavan-3-ol ratios (Fig. 3) (Petersen et al., 1991; Larkin et al., 1997; Dykes et al., 2014; Grabber et al., 2014; Klongsiriwet, 2016; Mueller-Harvey et al., unpublished data, 2010). Sample analysis by NIRS is rapid but requires robust calibrations that are based on laboratory analyses. Once calibrated, a single NIRS scan can also generate a large amount of information on other nutritional parameters, such as fiber, protein, soluble carbohydrate,

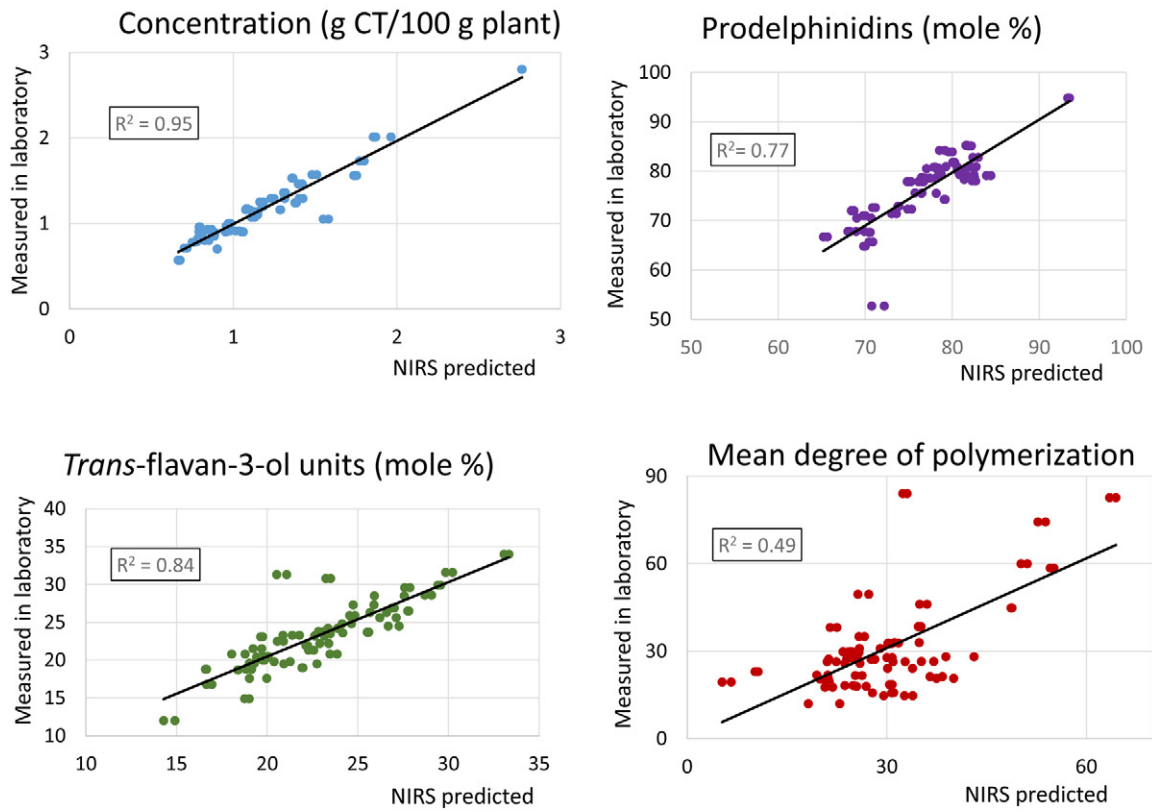

Fig. 3. Near-infrared reflectance spectroscopy (NIRS) for predicting condensed tannin (CT) composition of sainfoin plants that had been analyzed by thiolysis (Gea et al., 2011; Mueller-Harvey et al., 2011).

lignin, dry matter, and ash contents, as well as predicted digestibility and gross energy (Givens et al., 2000).

TANNIN VARIATION IN GERMPLASM COLLECTIONS AND POTENTIAL FOR TRAIT-DIRECTED PLANT BREEDING

Alignment of CT composition with function offers opportunities for exploiting their bioactivities, and germplasm collections offer a rich source of CT variation (Hayot Carbonero et al., 2011; Klongsiriwet, 2016). Concentrations of CTs vary greatly not only between plant species but also between accessions (Larkin et al., 1997; Mosjidis 2001; Sivakumaran et al., 2004; Gruber et al., 2008; Häring et al., 2008; Lorenz et al., 2010; Grabber et al., 2015; Hixson et al., 2016). Table 1 lists the variation in forage plants: birdsfoot trefoil tends to have the lowest (<5 g 100 g^{-1} dry matter) and sericea lespedeza and erect canary clover the highest CT concentrations (6– 20 g 100 g^{-1} dry matter).

The CT traits can also differ markedly between accessions, and between parts of the same plant (Springer et al., 2002). Examples are PC/PD ratios in sainfoin stem CTs of ~50:50 and in sainfoin leaf CTs of ~10:90 (Malisch et al., 2015). Although alfalfa (*Medicago sativa* L.) seed coats have CTs with mDP values of 4 to 7 and a PC/PD ratio of 93:7 (Koupai-Abyazani et al., 1993), only the smaller PC dimers and trimers have been detected in engineered alfalfa leaves (Hancock et al., 2012). This means that plant breeding can target CT composition and concentration, which is important because these traits have been linked to different bioactivities (see below).

Both concentration and composition can change with season (Theodoridou et al., 2011; Grabber et al., 2015; Muir et al., 2017), but accession differences tend to be much larger (Springer et al., 2002; Stringano et al., 2012). Importantly, environment did not affect the ranking of the CT traits of a few sainfoin accessions (i.e., there was no genotype × environment interaction) (Azuhnwi et al., 2013a ; Malisch et al., 2016); this demonstrates that there are opportunities for trait-directed breeding of new cultivars.

All enzymes involved in the biosynthesis of the CT building blocks (i.e., flavan-3-ols) have been identified, apart from the elusive final condensing enzyme(s) (Harding et al., 2013). Two genes and several myeloblastosis (MYB) transcription factors (i.e., proteins with myeloblastosis DNA-binding domains that regulate CT synthesis) are responsible for the production of two of the flavan-3-ols (i.e., catechin and epicatechin; Ferreyra et al., 2012; Cheynier et al., 2013; Zhu et al., 2015; Chezem and Clay, 2016), but the genes and transcription factors for the other flavan-3-ols await identification. The MYB transcription factors from barrelclover (*Medicago truncatula* Gaertn.) and rabbitfoot clover (*Trifolium arvense* L.) have been expressed in alfalfa and white clover (*Trifolium repens* L.), leading to stable and heritable CT production in their shoots and leaves (Hancock et al., 2012, 2014; Verdier et al., 2012; Albert, 2015). These developments provide a pathway for introducing CTs into the leaves of alfalfa and white clover. It would be interesting to explore whether the CTs that are already expressed in white clover flowers can be expressed in the leaves.

It should also be possible to alter the CT composition through conventional crossing experiments. Scioneaux et al. (2011) showed that CT composition, especially the average polymer size (the mDP value), in *Populus* L. was controlled by genetics and that environment (location) and season (months) had only a small effect. In addition, interspecies hybridization can generate plants with novel CT traits, as demonstrated with narrowleaf trefoil (*Lotus tenuis* Waldst. & Kit. ex Willd.) × birdsfoot trefoil hybrids (Escaray et al., 2014).

Despite considerable advances in plant science, genomic resources for forage legumes are still scarce, especially for CT-containing forages (Szczyglowski and Stougaard, 2008; Hayot Carbonero, 2011; Zarrabian et al., 2013; De Vega et al., 2015; Mora-Ortiz, 2015). These are necessary to harness the potential benefits of CTs, and for breeding of new varieties with improved agronomic, nutritional, and antiparasitic traits. The European and Asian sainfoin germplasm is very diverse in terms of morphology, anatomy, drought resistance, CT traits, and genetic polymorphism (Hayot Carbonero, 2011; Zarrabian et al., 2013; Malisch et al., 2015, 2016; Mora-Ortiz, 2015; Kempf et al., 2016, 2017; Mora-Ortiz et al., 2016; Kölliker et al., 2017). For example, anatomical trait analysis (xylem/phloem ratio, vessel and sieve tube diameters, and phloem width) suggested that large differences exist in water and solute transport mechanisms between accessions, which are important for drought resistance (Zarrabian et al., 2013).

Molecular markers are helpful in the analysis of genetic diversity, mapping and quantitative trait loci analysis, and genomics-assisted breeding. Next-generation sequencing technology is facilitating the identification and use of molecular markers in plant genetics and breeding. RNA-sequencing technology is an efficient way of obtaining sequence information of all the genes that are expressed in a given plant tissue, and it can also be mined for molecular marker polymorphisms. The first such library of expressed genes in sainfoin was obtained from five accessions using this technology (Mora-Ortiz et al., 2016). Annotation of the expressed genes in the library allowed identification of 59 genes involved in the CT biosynthesis pathway (Mora-Ortiz et al., 2016). It also provided the platform for identifying >3800 simple sequence repeat (SSR) markers and 77,000 single-nucleotide polymorphism markers (Mora-Ortiz et al., 2016). Phylogenetic analysis revealed that sainfoin is closely related to red clover (*Trifolium pratense* L.) and barrelclover. Some of the SSR markers

were used to assess the genetic diversity of European sainfoin accessions representing cultivars and noncultivars (ecotypes, landraces) (Kempf et al., 2016). These molecular markers are now available as tools for further genetic and genomic research. They were used to study inbreeding and self-fertilization in sainfoin. In a pilot study, one marker locus was identified that could explain up to 12% of the variation in CT composition (i.e., PC/PD ratio; Kempf et al., 2017). We anticipate that the sequence and molecular marker information now available (Kempf et al., 2016; Mora-Ortiz et al., 2016) can be used to assist breeding of novel sainfoin cultivars with CT traits that are optimized for animal nutrition and health.

AGRONOMY, WEED CONTROL, HARVESTING, AND PROCESSING OF TANNINIFEROUS FORAGE LEGUMES

There is currently considerable global interest in harnessing the benefits of CTs in forage legumes to support the sustainability agenda of agriculture; birdsfoot trefoil, sericea lespedeza, and prairie clover (*Dalea purpurea* Vent.) have been evaluated in the United States, Canada, New Zealand, and Switzerland (Mosjidis 2001; Waghorn 2008; Häring et al., 2008; Berard et al., 2011; Li et al., 2014; Grabber et al., 2015), sulla in Australia (de Koning et al., 2003, 2010; Heuzé et al., 2015) and sainfoin in Canada and Europe (Häring et al., 2008; Hayot Carbonero et al., 2011; Malisch et al., 2015; Bhattarai et al., 2016). The choice of which forage to grow will depend on the climate, soil, environment, and farming practices. The performance of these forages has been tested in pure stands and in combinations with partner species to increase the overall forage quantity and quality (Häring et al., 2008; de Koning et al., 2010; Hayot-Carbonero et al., 2011; Döring et al., 2013; Lüscher et al., 2014; Wang et al., 2015).

Cocultivation of Legumes and Companion Crops

Cocultivation of forage legumes with companion crops can deliver higher total yields (Nyfeler et al., 2009; Döring et al., 2013; Finn et al., 2013; Mora-Ortiz, 2015; Hunt et al., 2016; Malisch et al., 2017), more nitrogen fixed per hectare (Nyfeler et al., 2011; Vasileva and Ilieva, 2016) and improved dry matter and nitrogen digestibilities of coensiled mixtures (Wang et al., 2007), but success depends on the agronomic compatibility of the species, appropriate treatments, and development of new sainfoin cultivars with a more competitive canopy (Mora-Ortiz, 2015; Mora-Ortiz and Smith, 2016; Kölliker et al., 2017; Malisch et al., 2017).

Traditional mixtures for sainfoin establishment in the United Kingdom have included grasses such as timothy (*Phleum* L.) and meadow fescue [*Schedonorus pratensis* (Huds.) P. Beauv.] or undersowing with spring barley (*Hordeum* L.) as companions (Liu et al., 2008; Mora-Ortiz, 2015; Mora-Ortiz and Smith, 2016). However, new investigations showed that chicory (*Cichorium intybus* L.)—which also has antiparasitic properties—and oat (*Avena sativa* L.) can be cocultivated with sainfoin for a short period (i.e., two agronomic cycles; Mora-Ortiz, 2015; Mora-Ortiz and Smith, 2016), but chicory was found to be a very aggressive partner and to suppress sainfoin (Häring et al., 2008). North American researchers have also explored oat and alfalfa as companion crops for irrigated birdsfoot trefoil and sainfoin (Wang et al., 2015; Hunt et al., 2016), and this involved developing a new sainfoin population for cocultivation with alfalfa. The novelty of this work lies in the fact that this new sainfoin population has good competitiveness against alfalfa compared with previous sainfoin accessions, where seedlings first produced a long taproot and could become outgrown by weeds and many companion species (Hayot Carbonero et al., 2011).

Weed Control

Weed control in sainfoin can be achieved (Mora-Ortiz, 2015; Mora-Ortiz and Smith, 2016; Malisch et al., 2017) through appropriate choice of partner species, sowing densities, and cutting frequencies and can lead to stable sainfoin percentages (i.e., ~40% of the sward was sainfoin), which suffice to reduce the incidence of bloat (Wang et al., 2006; Malisch et al., 2017). This was also demonstrated for several other legume species in a Pan-European experiment (Finn et al., 2013; Connolly et al., 2017; Suter et al., 2017). Other strategies for weed control can include application of preemergence, postemergence, and maintenance herbicides (Sheldrick and Thomson, 1982; Moyer et al., 1990; Frame et al., 1998; Amiri et al., 2013; Mora-Ortiz, 2015). In the case of sainfoin, weed suppression is directly correlated with sainfoin establishment and yields.

Current understanding of CT expression in sainfoin suggests that it should be possible to breed new forage legumes with good yields and consistent CT profiles, which is important, as farmers need varieties with predictable CT traits.

Effects of Drought on Yield and Condensed Tannin Content in Sainfoin Accessions

However, instead of using irrigation to boost yield, others have focused on exploiting the taproot of sainfoin, as sainfoin can remain productive on dry, marginal soils and can continue to grow during unseasonably dry weather. To test this drought tolerance, 30 sainfoin accessions were subjected to drought for 18 wk, with the mean soil water potential of the upper 40 cm being below −2 MPa, and were compared with the dry matter yields of rainfed controls that received additional irrigation when the soil water potential was below −0.6 MPa. The results were also compared with

six other forage species (legumes, nonleguminous forbs, and grasses) of known drought tolerance. This showed that some sainfoin accessions were as tolerant under severe drought as alfalfa, and a few accessions even exceeded its drought tolerance. Most sainfoin accessions also had lower drought losses than chicory (Malisch et al., 2014). In addition, the CT traits were assessed under drought and control conditions for five of these accessions. Although CT composition was hardly affected by drought, the CT concentrations increased at the vegetative, but not the reproductive, stage (Malisch et al., 2016).

Moreover, there were no interactions between drought and accession for CT traits, and it remains to be seen whether the ranking of sainfoin accessions according to their CTs is maintained across other environments (Malisch et al., 2016). Another study with 100 sainfoin genotypes from 10 ecotypes showed that drought tolerance was, however, correlated with leaf proline content (Irani et al., 2015). This correlation is in accordance with previous studies, and although the cause-and-effect relationship between proline and drought tolerance is not yet fully understood, proline can act as an osmoprotectant, thus stabilizing membranes and maintaining cell turgor. Additionally, there is some indication that it might contribute to upregulation of drought tolerance genes (Szabados and Savouré, 2010; Per et al., 2017). Therefore, the current understanding of CT expression in sainfoin suggests that it should be possible to breed new cultivars with good yields, drought tolerance, and consistent CT profiles, which is important, as farmers need forage legumes with predictable CT traits.

Grazing vs. Preservation

Legume swards expressing CTs can be grazed safely, as they are nonbloating (Wang et al., 2012; MacAdam and Villalba, 2015), but optimal use of CT forages would most likely be as a substitute for existing legumes in mixed swards, such as white clover with ryegrass (*Lolium* L.). This is because dry matter yields are greater from grasses than from legumes (subject to adequate nitrogen availability), and because farmers in temperate climates have identified forage species (including grasses) that enable profitable farming. In these situations, the CT concentration in the diet is diluted by the contribution of the CT-free component of the sward (Waghorn and Shelton, 1997), so high CT concentrations of the bioactive CT legume might be most useful, as the CTs in one forage can affect digestion of protein in the companion forage (Waghorn and Jones, 1989).

Several CT forages possess antiparasitic properties, which are of special interest for sheep and goat grazing systems (Hoste et al., 2010), or when there is anthelmintic resistance (Terrill et al., 2012). Condensed tannins may also provide benefits when immunity of animals is low (i.e., around parturition or weaning), but the need to combat parasitism at such times does not necessarily coincide with the availability of fresh CT forages. Therefore, bioactivity needs to be maintained when processing CT forages into hay, pellets, or silages. Conserved forages allow out-of-season feeding, but may, especially in the case of pellets (Girard et al., 2016b), also offer opportunities for standardizing and optimizing CT traits and enable transportation to other regions.

Processing of CT forages into hay, pellets, or silages can have a marked effect on CT concentration and extractability (Fig. 4) and appears to increase protein-bound, and possibly covalently linked, CTs (Terrill et al., 1997; Minnée et al., 2002; Lorenz et al., 2010; Vernhet et al., 2011; Ramsay et al., 2015; Girard, 2016; Huang et al., 2016). Ensiling sainfoin or sulla reduced ammonia production, improved silage quality, and protected plant protein during fermentation, which improved its nutritional value relative to forages without CTs (Niezen et al., 1998b; Lorenz et al., 2010). There is a need for feeding trials to assess the biological significance of unextractable CTs in terms of ruminal or intestinal digestion and efficacy against parasitic nematodes. These questions could perhaps be addressed through experiments that explore accession differences in terms of unextractable CTs.

ELUCIDATING RELATIONSHIPS BETWEEN CONDENSED TANNIN STRUCTURES AND ANTIPARASITIC EFFECTS

Parasitism imposes a considerable nutritional penalty on animals, and therefore controlling the parasite burden will indirectly benefit the nutritional status of animals. This is the reason for noting that CT forages can be used for nutraceutical purposes, which refers to a combined action of nutritional and antiparasitic benefits that include anthelmintic (Terrill et al., 2012; Hoste et al., 2015, 2016) and anticoccidial effects (Kommuru et al., 2014; Saratsis et al., 2016). Most evidence of anthelmintic effects of CTs stems from in vitro assays, but some in vivo feeding trials

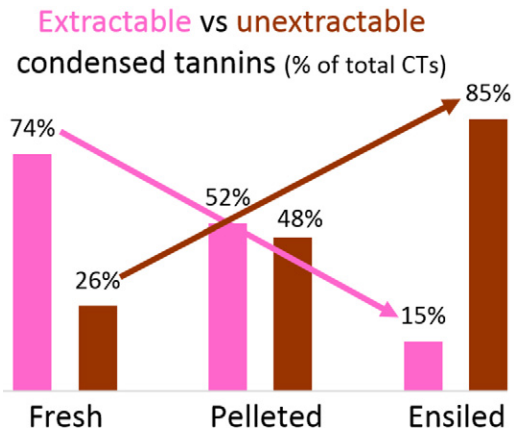

Fig. 4. Average percentages of extractable and unextractable condensed tannins (CTs) in fresh, pelleted, and ensiled sainfoin, sulla, and birdsfoot trefoil (Minnée et al., 2002; Lorenz et al., 2010; Ramsay et al., 2015).

under controlled experimental conditions have also yielded promising results. It is therefore timely to summarize the emerging trends and CT structure-activity relationships, which were obtained with a range of in vitro assays, to guide future feeding trials.

Antiparasitic Effects of Prodelphinidins and Galloylated Condensed Tannins

Condensed tannins with high molar percentages of PDs and galloylation have given good anthelmintic effects in vitro with parasite larvae (L). These include inhibition of L3 migration and L4 motility with *Ascaris suum*, as well as L1 feeding inhibition, adult motility assays of *Ostertagia ostertagi* and *Cooperia oncophora*, and larval exsheathment inhibition of *Haemonchus contortus* and *Trichostrongylus colubriformis* of L3 larvae (Brunet and Hoste 2006; Quijada et al., 2015; Desrues et al., 2016a; Ramsay et al., 2016). These findings could explain the excellent in vivo results with sheep or goats that were obtained when feeding sericea lespedeza (these CTs consist of almost pure PDs with high mDP values), big trefoil, sainfoin, sulla plants, or hazelnut (*Corylus* L.) peels (high PD percentage) and lentisk browse (galloylated compounds) (Landau et al., 2010; Rodríguez-Pérez et al., 2013; Hoste et al., 2015). These in vivo trials found better host resilience and lower fecal egg counts of *H. contortus*, *Teladorsagia circumcincta*, and *T. colubriformis* and of a mixed gastrointestinal nematode infection, which were attributed to lower worm fecundity (Niezen et al., 1995, 1998a; Paolini et al., 2005; Landau et al., 2010), and also lower *Eimeria* oocyst counts (Kommuru et al., 2014). Control of *T. colubriformis* was reflected in higher liveweight gains by lambs fed sulla than alfalfa (Niezen et al., 1995).

Antiparasitic Effects of the Mean Degree of Condensed Tannin Polymerization

The average size of CTs in a polymeric mixture (mDP value) can also affect their antiparasitic activity, with larger polymers being more potent against exsheathment of *H. contortus* L3 and larval feeding of *O. ostertagi* and *C. oncophora* L1 larvae in vitro (Quijada et al., 2015; Desrues et al., 2016a). However, PD-rich plants tend to have CTs with high mDP values and high CT concentrations (Kommuru et al., 2014; Laaksonen et al., 2015; Mueller-Harvey et al., unpublished data, 2016), confounding our understanding of CT structure-activity relationships. The characteristics of CTs responsible for antiparasitic effects need to be understood to identify optimal feed sources.

Apart from the examples above, very few other plants with high molar PD or galloyl percentages or high mDP values (>15) have been evaluated in vivo to determine their efficacy against gastrointestinal parasites. The plant kingdom remains a rich and underexplored resource of such promising CTs (Table 3), and it would be timely to test these in vitro results by feeding plants, browse, or agroindustrial byproducts with differing CT traits to parasitized livestock. As traits vary across accession and environment, it is important to analyze the dietary CTs to establish their relationships with antiparasitic effects. This would enable guidelines to be formulated for development of new plant cultivars for optimal bioactivities.

One other aspect also needs investigation: are CT traits that are optimal for antiparasitic effects compatible with nutritional benefits, or would farmers require plants with different CTs for either antiparasitic or nutritional purposes? Given the encouraging results with sericea lespedeza and panicledleaf ticktrefoil [*Desmodium paniculatum* (L.) DC.], especially in parasitized animals, anthelmintic and nutritional benefits may not be mutually exclusive (Terrill et al., 2012; Cherry et al., 2014), but other options could include sacrificing nutrition for a short period, enabling an appropriate period of CT feeding to achieve parasite control.

Mechanisms of Antiparasitic Condensed Tannin Action

A key question concerns the mechanisms by which CTs exert their effects. It is thought that the ability of CTs

Table 3. Plant materials containing condensed tannins (CTs) with structural characteristics that are of interest for conferring antiparasitic activities.

CT traits	Plant species and part	Reference
Prodelphinidin-rich CTs	Peels of hazelnut (*Corylus avellana* L.) seeds; leaves of Persian ironwood [*Parrotia persica* (DC.) C.A. Mey.], London plane (*Platanus* × *hispanica* Mill. ex Münchh. [*occidentalis* × *orientalis*]), black or red currant (*Ribes nigrum* L., *R. rubrum* L.) bushes, black locust (*Robinia pseudoacacia* L.); sainfoin (*Onobrychis viciifolia* Scop.), sulla (*Hedysarum coronarium* L.), sericea lespedeza [*Lespedeza cuneata* (Dum. Cours.) G. Don] plants; white clover (*Trifolium repens* L.) flowers; erect canary clover [*Dorycnium rectum* (L.) Ser.]	Sivakumaran et al., 2004; Tibe et al., 2011; Mechineni et al., 2014; Hoste et al., 2016; Ropiak et al., 2016b
Galloylated CTs	Shea (*Vitellaria paradoxa* C. F. Gaertn.) nuts, persimmon (*Diospyros kaki* Thunb.) fruits, lentisk (*Pistacia lentiscus* L.) leaves, carob (*Ceratonia siliqua* L.) fruits, grape (*Vitis vinifera* L.) seeds, great water dock (*Rumex hydrolapathum* Huds.) roots, dock (*Rumex obtusifolius* L.) leaves	Papagiannopoulos et al., 2004; Spencer et al., 2007; Li et al., 2010; Rodríguez-Pérez et al., 2013; Ramsay et al., 2016; Ropiak et al., 2016b
High mean degree of polymerization values	Erect canary clover plant, persimmon fruits, apple (*Malus domestica* Borkh. spp., cider cultivars); leaves of Persian ironwood, London plane, black currant, black locust; white clover flowers; sericea lespedeza plant	Guyot et al., 2001a; Sivakumaran et al., 2004; Li et al., 2010; Mechineni et al., 2014; Hoste et al., 2016; Ropiak et al., 2016b

to suppress gastrointestinal nematodes stems from their ability to bind with proteins (Hoste et al., 2012). Thus, CTs could act by inhibiting key parasite enzymes, and indeed inhibition of parasite glutathione-*S*-transferases, which play an important role in the detoxification of xenobiotic compounds, has been confirmed in in vitro experiments (Hansen et al., 2016). Prodelphinidins have more phenolic groups capable of forming hydrogen bonds with proteins than PCs (Fig. 1), but it has also been established that the mDP value is the most important factor for CT protein aggregation and precipitation (Zeller et al., 2015b; Ropiak et al., 2017). Taken together, these observations could explain why PDs, which generally have larger mDP values than PCs, have better anthelmintic properties (Hoste et al., 2016; Kommuru et al., 2014, 2015).

By using electron microscopy, a number of studies have revealed physical deformations of *H. contortus* adult worms that had been exposed to CTs in either in vitro experiments or collected after feeding sericea lespedeza or sainfoin plants or tzalam [*Lysiloma latisiliquum* (L.) Benth.] leaves (Martínez-Ortíz-de-Montellano et al., 2013; Kommuru et al., 2015). The shriveled surfaces and plaque formations around orifices could account for inhibition of feeding and lowering of fecundity and are thought to stem from CTs interacting directly with proteins on parasite surfaces (Ropiak et al., 2016a). Scanning and transmission electron microscopy detected not only external but also internal damage to the cuticle and sensilla of the lip region of young and adult *Caenorhabditis elegans* by different CT types, and to the cuticle and underlying tissue (i.e., muscle cells) and intestinal cells of *H. contortus* L3 larvae and adults and of *T. colubriformis* L3 larvae by sainfoin and tzalam CTs (Brunet et al., 2011; Martínez-Ortíz-de-Montellano et al., 2013; Ropiak et al., 2016a). Given the low uptake (bioavailability) of polymeric CTs in mammalian tissues in comparison with monomeric flavonoids (Li and Hagerman, 2013), it would be worth exploring whether the internal tissue damage arises from the CTs or from other compounds that may be present in plant extracts (Brunet and Hoste, 2006; Williams et al., 2015; Klongsiriwet et al., 2015; Desrues et al., 2016a; Mengistu et al., 2017). It also raises the question whether mixtures of CTs plus co-occurring smaller plant compounds (such as quercetin, luteolin, cinnamaldehyde, etc.) can cause both external and internal damage, or whether internal damage may be due to a disruption of nematode metabolism, in turn causing necrosis of cells and tissues. Condensed tannins together with a flavone (luteolin), a flavonol (quercetin), or cinnamaldehyde can act synergistically against parasitic nematodes in vitro (Barrau et al., 2005; Klongsiriwet et al., 2015; Hoste et al., 2016; Ropiak et al., 2016a). These in vitro findings are yet to be tested in feeding trials using combinations of feeds with different CTs and such monomeric compounds, but such combinations should hopefully lead to future on-farm applications.

Tissue and Host Responses to Condensed Tannins

In addition to direct anthelmintic effects against parasites, more work is warranted on how CTs may influence host responses to parasites. Niezen et al. (2002) measured higher antibody titers against antigens to adult *T. circumcincta* and adult and larval *T. colubriformis* when lambs were fed with sulla compared with alfalfa. This may be a result of increased intestinal flow of proteins and amino acids associated with ruminal protection of protein degradation, which can contribute to the host's ability to maintain growth, immune function, and regulation of worm populations (Ríos-De Álvarez et al., 2008; Ramírez-Restrepo et al., 2010; Hoste et al., 2012). Moreover, recent in vitro studies have also indicated that CTs can directly modulate the activity of immune cells such as $\gamma\delta$ T cells and dendritic cells, potentially enhancing the host's innate immune response (Tibe et al., 2012; Williams et al., 2016, 2017). Similar to other bioactivity studies, immune-modulating activity in vitro is highly dependent on CT size, with mDP >6 eliciting a stronger response than CTs with mDP <6, and flavan-3-ol monomers have little or no effect (Williams et al., 2016, 2017).

Effects of Fermentation on Condensed Tannin Activity

Work has started on determining how fermentation affects CT concentrations and bioactivities. According to results from the HCl-butanol-acetone or thiolysis assays, fermentation can reduce "apparent" CT concentrations by 30 or 85%, respectively, in silages (Mena et al., 2015; Ramsay et al., 2015) and the gut (Desrues et al., 2017; Quijada et al., 2018). However, despite these apparent losses, sericea lespedeza and sainfoin silage extracts and silages still exerted anthelmintic effects in vitro by inhibiting the exsheathment of *H. contortus* L3 larvae and in vivo by lowering adult worm burden and fecal egg counts (Heckendorn et al., 2006; Manolaraki, 2011; Terrill et al., 2016). This may either be due to sufficient quantities of undegraded CTs or hydrolysis of anthelmintic flavonoids from inactive glycosides (Manolaraki, 2011), or to protein-bound CTs surviving ruminal fermentation and exerting anthelmintic activity in the abomasum, where a lower pH facilitates dissociation of the CT-protein complex (Jones and Mangan, 1977). A high activity against *O. ostertag*i nematodes of the abomasum contrasted with no activity against intestinal *Cooperia oncophora* nematodes and was associated with 2.3% CTs (g 100 g^{-1} dry matter by thiolysis) in the abomasum vs. 0.02% CTs in the intestine (Desrues et al., 2016a, 2016b, 2017). However, both nematode species were affected by CTs in the in vitro larval feeding inhibition assay (Desrues et al., 2016a).

EFFECTS ON RUMINAL FERMENTATION AND CONSEQUENCES FOR RUMINANT NUTRITION AND GREENHOUSE GAS EMISSIONS

Effects on Nutrition and Dietary Protein Use

Nutritional effects of CTs are currently understood in general terms, and information concerning the ways that CTs affect specific processes are only now being defined by determining the CT traits of forages used for in vivo and in vitro measurements and their effects on the microbiome (Grosse Brinkhaus et al., 2016, 2017). Binding to dietary proteins and reduction in rumen proteolysis, resulting in higher proportions of dietary protein passing to the small intestine, have been measured (Waghorn, 2008), but details and mechanisms are not understood. The findings by Kariuki and Norton (2008) indicated that the release of dietary protein between the abomasum and terminal ileum was correlated with the protein precipitation capacity of CTs, which depends on the structures of both the CTs and the proteins (Hagerman and Butler, 1981; Dobreva et al., 2012). The measurements of Wang et al. (1996) are especially important in this regard, because they showed that in sheep fed birdsfoot trefoil with polyethylene glycol (PEG, which inactivated the CTs), ~80% of amino acid absorption occurred in the first half of the intestine, but the entire length of the intestine was required for absorption when CTs were active. Hence, the unknowns concerning CT interactions with rumen function, microbial growth, and intestinal absorption need to be evaluated in terms of dietary CT traits.

The current evidence suggests that only four forage legumes–birdsfoot trefoil, sainfoin, sulla and crownvetch [*Securigera varia* (L.) Lassen]–support higher ruminant growth rate or milk yield when fed as a sole diet, compared with CT-free diets (Burns et al., 1972; Waghorn et al., 1990, 1997; Mueller-Harvey, 2006; Rochfort et al., 2008; Waghorn, 2008; Patra and Saxena 2010; MacAdam et al., 2011; Naumann et al., 2013; Piluzza et al., 2014; MacAdam and Villalba 2015). However, other CT forage legumes or some CT browse species may be beneficial when fed as part of a diet (e.g., lotuses; Ayres et al., 2006). A universal consequence of dietary CTs is a reduction in urinary nitrogen excretion (because of reduced rumen proteolysis) and an increase in fecal excretion of nitrogen (Mueller-Harvey, 2006; Waghorn, 2008).

Evaluation of nutritional benefits can be complicated, especially if both feed quality (digestibility) and voluntary feed intake are important. Variations in intake are likely to confound comparisons of digestibility, but digestible matter intake is a recognized indicator of performance. There are a number of ways that the effects of CTs can be determined, and several studies (Table 4) have fed a CT forage to two groups of animals, with one receiving daily doses (or intraruminal infusions) of PEG to bind and deactivate the CTs. This ensures that the majority of the diet is the same and enables the effect of the CTs to be evaluated. An option used by some researchers to compare a CT diet with a similar non-CT diet is fraught with difficulties. No species are optimal for comparisons, and any differences in composition, digestion, or intake will compromise the evaluation of the CTs. It is also important to distinguish the CT effect from a legume effect when evaluating a CT forage fed with grasses, for example, and if the animals are parasitized, any benefits of CTs could arise from direct or indirect impacts on the parasites (Hoste et al., 2015, 2016).

Furthermore, because CTs reduce the digestion of protein in the rumen and over the entire gastrointestinal tract (Waghorn, 2008), it is unlikely that any nutritional benefits will result when dietary crude protein is insufficient. Benefits are more likely when dietary protein is in excess of requirements. However, if in the presence of enough protein (or amino acids) other nutrients are limiting (e.g., energy intake or phosphorus), providing additional protein will not improve production (Waghorn, 2008; Pagán-Riestra et al., 2010). Hence, the methods by which animals are fed and their physiological state (e.g., lactating, growing, or at maintenance) when evaluating the nutritional effects of CTs can contribute to inconsistencies in findings. In addition, comparative measures of digestion may be confounded by variation in intakes, because increasing intakes may reduce digestibility (Tyrrell and Moe, 1975), but on other occasions have no effect (Hammond et al., 2013).

A number of studies with sainfoin and birdsfoot trefoil have also yielded contradictory results, often with lower or no production benefits measured when compared with CT-free controls (Thomson et al., 1971; Waghorn et al., 1997; Theodoridou et al., 2010; Aufrère et al., 2013; Azuhnwi et al., 2013b; Copani et al., 2016; Girard et al., 2016a, 2016b). In addition to the constraints mentioned above (dietary crude protein concentration, or whether other nutrients are limiting production), variations in CT traits within germplasms may also affect animal responses (Grabber et al., 2015). For example, the UK Hampshire Common and Cotswold Common sainfoin accessions had higher PC/PD ratios (>30:70) than the continental European Visnovsky accession (<19:81) (Stringano, 2011; Stringano et al., 2012) and could have accounted for these contradictory reports. Birdsfoot trefoil has delivered good growth rates in the United States and New Zealand, and the PC/PD ratios were ~80:20 (Meagher et al., 2004; MacAdam and Villalba, 2015). However, when two sainfoin cultivars with PC/PD ratios of 24:76 (Visnovsky) and 37:63 (Perly) were fed to lambs infected with *H. contortus*, concentrations of essential amino acids in plasma were higher than when both diets had

Table 4. Variation of nutritional effects achieved with different sainfoin accessions and feeding regimes in comparison with two trefoil species and crownvetch forage legumes.

Plant species†	PC/PD‡	mDP§	Nutritional effects	References
Sainfoin accessions:				
Visnovsky (dried or ensiled)	19:81–24:76	16–29	Lambs: lower apparent organic matter and fiber digestibility, increased plasma concentration of essential amino acids, no effect on body N retention	Scharenberg et al., 2007; Stringano et al., 2012; Azuhnwi et al., 2013b
Perly (pellets, 20% of basal diet)	ND	ND	No effect on N retention in dairy cows	Grosse Brinkhaus et al., 2016
Perly (ensiled with timothy grass)	ND	ND	Lower growth rate of lambs than on red clover mixtures	Copani et al., 2016
Perly (ensiled)	ND	ND	Condensed tannins (CTs) had no effect on N retention compared with the +polyethylene glycol (PEG) control	Theodoridou et al., 2012
Shoshone (grazed)	ND	ND	Comparable weight gains by beef calves grazing sainfoin and alfalfa diets	Maughan et al., 2014; Villalba et al., 2015
Renumex (grazed)	ND	ND	Growth rate and slaughter data were the same for sainfoin- and alfalfa-fed lambs	Karnezos et al., 1994
No details (fresh forage)	ND	ND	CTs in sulla, but not in sainfoin, improved amino acid absorption compared with the +PEG control	Bermingham et al., 2001
Zeus/Esparcette (ensiled)	ND	ND	Dairy cows: higher milk yield compared with grass silage	Huyen et al., 2016b
UK accessions (Sombourne, Hampshire, Cotswold Common)	29:71–33:67	12–32	Farmers report high growth rates of lambs fed UK sainfoin accessions (but no accession details were provided in feeding trial)	Thompson et al., 1971; Stringano et al., 2012
Fakir-cut and stall-fed	ND	ND	Positive effect on N retention compared with rye grass and clover, greater recycling and degradation of urea, improved utilization of recycled N	Egan and Ulyatt, 1980
Lotus spp.				
Big trefoil (Maku–indoor trials)	20:80	ND	Reduced growth rate, lower apparent digestibility of essential amino acids	Waghorn et al., 1997; Min et al., 2003; Meagher et al., 2004; Waghorn, 2008
Birdsfoot trefoil (Goldie [indoor trials], Norcen, Oberhaunstadter [grazed])	84:16	ND	Enhanced absorption of essential amino acids, positive effects on livestock production (increased cattle and sheep growth, cow and sheep milk yield, sheep fertility and wool growth)	Waghorn and Shelton, 1997; Waghorn et al., 1997; Meagher et al., 2004; Waghorn, 2008; MacAdam et al., 2011; MacAdam and Villalba, 2015
Crownvetch				
Chemung, Penngift (field cured, wind-rowed, baled)	26:74	>13	Larger daily gain of cattle and sheep under grazing or stall feeding compared with alfalfa or sericea lespedeza.	Burns et al., 1972, 1977 (Note: our CT data are unpublished and not from the plants that were fed)

† With information, if available, on variety or accession and method of feeding.

‡ PC/PD, procyanidin:prodelphinidin ratio; ND, not determined.

§ mDP, mean degree of polymerization.

been treated with PEG to inactivate CTs (Azuhnwi et al., 2013b); it is not known whether the higher plasma concentrations were a direct result of CTs on protein digestion or an anthelmintic effect against *H. contortus*. Variable results could be investigated by considering CT traits in conjunction with diet composition (e.g., protein, fiber, water-soluble carbohydrate, starch, and amino acid contents, as these can be affected by environment; Grabber et al., 2015), forage harvesting, or preservation methods. This means that nutritional evaluations of CT plants need to be undertaken under documented and controlled conditions, with information on harvesting and preservation methods (grazed, dried as hay or pellets, or ensiled).

Another potentially important issue could be how animals are fed, because CT traits also vary within plants. In Spain, it is traditional to take the first sainfoin cut as hay and then leave animals to graze the regrowth (Dr S.F. Demdoum, personal communication, 2010). Under light stocking regimes in the United Kingdom, sheep will only eat the sainfoin tops (flowers and younger leaves) and leave older leaves and stems intact (Mueller-Harvey, personal observation). Therefore, feeding whole plants as pellets, hay, or silage could result in forages with very different nutritive values than grazed forages, and also because conservation affects CT concentrations; these facts could be another reason for the contradictory results mentioned above.

One of the very few feeding trials that compared two birdsfoot trefoil cultivars with similar nutritional composition—apart from their CT concentrations—found that the 'Maitland' cultivar (3.5 g CTs 100 g^{-1} dry matter) achieved better nitrogen retention in sheep than the 'Empire' cultivar (0.5 g CTs 100 g^{-1} dry matter) (Waghorn et al., 1987). Greater retention of dietary protein suggested that the Maitland CTs protected more soluble protein from digestion in the

rumen but did not interfere with protein digestion and absorption in the abomasum and small intestine. More research is needed to establish the precise fate of dietary protein in the abomasum and intestine in the presence of CTs. Estimates based on their potentially beneficial protein protection effects suggest that if alfalfa had just 1% CTs in its dry matter, this could achieve a 12% increase in net returns for US dairy farmers (McCaslin et al., 2014).

It is unfortunate that few studies describe the composition of CTs in plants that have been fed to ruminants, and the LegumePlus program has attempted to address this issue by encouraging communication and collaboration among chemists and plant and animal scientists. However, we still do not know which CT traits plant breeders should be targeting (i.e., dietary CT concentration, PC/PD ratio, or mDP values or a combination of these) to increase livestock production and product quality. Apart from a few in vitro studies (Azuhnwi et al., 2013b; Hatew et al., 2016; Huyen, 2016; Huyen et al., 2016b), no feeding trials have attempted to resolve the relationships between animal production and CT traits. Interdisciplinary research is needed to uncover the mechanisms by which CTs exert their effects on rumen (microbial) digestion and utilization of dietary nutrients, and this will require compositional (rather than colorimetric) analyses of the dietary CTs.

The size of CTs is the key parameter that controls CT protein aggregation and precipitation. This has been illustrated using bovine serum albumin (BSA) and gelatin, where aggregation increased markedly as the mDP values increased from 3 to 8 (corresponding to CT molecular weights of ~1000 to 2400 Da), but there were only small differences in the efficacies of larger CTs with mDP >9 to aggregate the proteins (Ropiak et al., 2017). It would be worth testing whether CT size also affects the mechanism by which CTs interact with dietary or endogenous animal proteins, as Zeller et al. (2015b) showed that relatively more alfalfa protein was precipitated by CTs than BSA protein, which could be due to the major leaf protein ribulose-1,5-bisphosphate carboxylase/oxygenase (RuBisCo) being much larger (560 kDa) than BSA (67 kDa). It will be necessary to determine if CTs with different mDP values affect dietary protein degradation and digestion, especially as the major proteins in forages (RuBisCo), beans (albumins, prolamins), and intestinal gut tissue and mucoproteins have quite different structures, which will influence their interactions with CTs (Hagerman and Butler, 1981). As an aside, we have included bean proteins in this context, as several papers from the 1960s and 1970s on the nutritional effects of dietary tannins originated from feeding CT-containing beans (Jansman, 1993; Lowry et al., 1996). It would therefore be timely to review some of these results in the light of new knowledge on CT and protein structures.

Effects on the Quality of Animal Products

There are some effects of sainfoin diets on milk and meat quality and on the animal's physiological response to dietary CTs. Cattle fed a grass–sainfoin (mixture of Zeus and Esparcette accessions) silage or sheep grazed on big trefoil ('Maku') partitioned energy towards protein synthesis, rather than lipid synthesis, compared with grass–corn (*Zea mays* L.) silage and white clover diets, respectively (Purchas and Keogh, 1984; Huyen et al., 2016a). Feeding dairy cows with sainfoin pellets lowered milk and blood urea concentrations compared with alfalfa and birdsfoot trefoil pellets (Girard et al., 2016b; Grosse Brinkhaus et al., 2016) because of the reduction in rumen proteolysis and ammonia absorption.

Sainfoin and birdsfoot trefoil diets reduced bacterial biohydrogenation in the rumen, increased unsaturated fatty acid contents in milk, cheese, and meat products, and reduced indole and skatole in lamb meat (Priolo et al., 2005; Schreurs et al., 2007; MacAdam and Villalba, 2015; Girard et al., 2016a, 2016b; Huyen, 2016; Huyen et al., 2016a). Skatole and indole were associated with "fecal" flavor characteristic of pasture-fed products and originated from amino acid degradation by *Clostridium aminophilum* (Attwood et al., 2006), which was relatively sensitive to CTs (Sivakumaran et al., 2004). More recently, Campidonico et al. (2016) reported that CTs and polyphenol oxidases in a sainfoin–red clover silage mixture generated additive effects that increased the intramuscular unsaturated fatty acid contents of lambs compared with a pure grass diet.

Effects on Nitrogen and Methane Emissions

The reduction in urinary nitrogen and increase in fecal nitrogen excretion seem to be universal consequences of dietary CTs fed to ruminants and are important because a greater proportion of nitrogen is lost from urine than feces. Thus, CTs can improve soil nitrogen status, lower emissions of the potent greenhouse gas nitrous oxide (N_2O), and lessen nitrogen leachate into waterways and groundwater (Kingston-Smith et al., 2010; Theodoridou et al., 2010). A shift from urinary to fecal nitrogen could redue nitrogen losses by 25% and achieve savings on nitrogen fertilizers based on preliminary estimates from the Integrated Farm System model for dairy farms (Zeller and Grabber, 2015). Other opportunities for reducing the environmental impact of nitrogen emissions from ruminant livestock include the use of galloylated CTs and epigallocatechin gallate (a galloylated flavan-3-ol monomer), because they are urease inhibitors (Huynh-Ba et al., 1994; Powell et al., 2011; Takeuchi et al., 2014). Urease inhibition reduces ammonia emissions from urine and subsequent N_2O production (Kingston-Smith et al., 2010). Grape seeds and some agroindustrial residues are sources of galloylated CTs and epigallocatechin gallate (Li et al., 2010; Lee et al., 2014; Ramsay et al., 2016), and

application to the barn floor could lessen ammonia and N_2O emissions from intensive systems.

It is clear that CTs can provide important benefits to ruminant farming; however, high dietary concentrations or CTs with the "wrong" compositional traits will lower the digestion and utilization of dietary protein and absorption of essential amino acids by ruminants (Min et al., 2003). The challenge is to identify which CT traits are best able to enhance utilization of dietary protein to improve animal production, environmental sustainability, and profitability for farmers.

CONCLUSIONS AND POSSIBLE DIRECTIONS FOR THE FUTURE

Condensed tannins are the fourth largest group of secondary plant metabolites in the plant kingdom and provide opportunities for breeding forage legumes with novel CT traits. Research on CT-containing feeds has the potential to improve ruminant health by preventing bloat and mitigating effects of parasitism, as well as lowering environmental footprints and improving the sustainability of food quality and production for consumers (Tedeschi et al., 2014). Innovative molecular approaches have enabled alfalfa and white clover to express PC dimers and trimers in their foliages (Hancock et al., 2012, 2014; Verdier et al., 2012; McCaslin et al., 2014). Apart from research by Mosjidis and colleagues, who selected for low CT concentration and grazing-tolerant sericea lespedeza for cattle production (Mosjidis, 2001), there has been hardly any plant breeding for enhanced CT composition; however, progress is becoming feasible, as new genomic data and molecular markers for CTs have been obtained (Kempf et al., 2016; Mora-Ortiz et al., 2016). Alternatively, cultivars with specific CT traits could also be obtained by conventional selection, focusing on the compositional CT differences that already exist between plant species and between, but also within, accessions and plant parts.

This review has highlighted that interdisciplinary research is essential for developing new forages with desirable CT traits and bioactivities, and that it requires well-coordinated inputs from plant scientists, chemists, animal nutritionists, and parasitologists. Such collaborations have succeeded in identifying the large variation in CT traits and their in vitro nutritional and parasitological effects that pertain to the germplasms of sainfoin and birdsfoot trefoil (Stringano et al., 2012; Grabber et al., 2014; Malisch et al., 2015), which grow in temperate climates. Such variation is likely to exist also in other species (e.g., sericea lespedeza and prairie clover, which grow in warm humid and colder regions, respectively; Mosjidis, 2001; Berard et al., 2011). It would be timely to explore these in vitro results by feeding CT forage legumes with varying CT traits to develop robust targets and tools for plant breeding.

Researchers must become aware that most colorimetric assays are not appropriate for determining CT concentrations or compositions. For example the *p*-dimethylaminocinnamaldehyde and HCl-vanillin reagents detect CTs, but also their monomeric flavan-3-ol precursors, and the Folin–Ciocalteu reagent detects all phenolic groups, whether in monomeric flavonoids, proteins, or condensed or hydrolyzable tannins (Schofield et al., 2001). These problems are made worse by the use of inappropriate CT material to construct calibration curves for analysis of CT concentrations, as mentioned above (Grabber et al., 2013; Krueger et al., 2013). The fact that a multitude of methods or standards have been chosen to measure CTs has prevented comparisons of published CT concentrations from feeding trials and the setting of optimum thresholds for CT traits. Appropriate CT analysis is crucial to progressing this field.

Laboratory studies have probed the impact of CT concentration, polymer size, and PC/PD ratios on ruminal fermentation and antiparasitic effects. The time has come for feeding trials with selected forages of similar nutritional compositions but different CT traits to test these in vitro results and establish the in vivo nutritional and antiparasitic effects associated with contrasting CT traits. This will also require comparison of forages that have been grazed or processed into hay, pellets, or silages, because CTs become less extractable on processing, but the underlying mechanisms and biological significance of these changes are yet to be explored. Such studies will help to optimize dietary protein utilization and energy partitioning and reduce the environmental footprint of livestock production.

We also need answers to the following questions: what effects do CTs exert on intestinal cells in ruminants and nonruminants in terms of nutrient absorption and cell signaling cascades? What are the mechanisms by which CTs protect dietary protein from rumen degradation and affect amino acid absorption from the intestine (e.g., in the presence of CTs, amino acid absorption takes place across the entire intestine; however, in the absence of CTs, absorption occurs in the first third of the intestine; Wang et al., 1996)? What are the effects on the ruminal or colonic microbiomes? How does the impact of CTs on recycling of urea-nitrogen modify the utilization of energy and protein in dairy cows, and what is the origin of higher fecal nitrogen outputs in the presence of CTs? We need to establish the fate and bioactivity of CTs during ensiling and digestion, their mechanisms of action against parasites along the digestive tract, and their effects on the in vivo immune response and establish relationships with CT concentration and composition. This review has also highlighted the need for analytical methods and CT standards that are fit for purpose so that published CT values can be compared between research groups and experiments.

Although considerable progress has been achieved over recent years, questions remain on how to translate the research results into practice, such as:

- How should we use CT forages? Is it better to graze forages with low CT concentrations and use high-CT forages as supplements?
- Can high-CT and CT-free forages be grazed together in strips, and could this improve utilization of dietary protein?
- Can the competitiveness and persistency of CT-containing legumes be increased so they can be sown together with other crops and achieve successful weed suppression?
- What are the ecological implications of intake and selectivity by different ruminant species on plant persistence in the field?
- Does an optimum CT concentration and composition exist that can deliver nutritional plus antiparasitic effects? According to Cherry et al. (2014), it may be possible to achieve both, or do we need forages with different CT compositions (i.e., cultivars that either improve protein utilization or possess antiparasitic effects)?
- Will increased use of CT forages generate resistance of parasites to CTs, and how can feeding regimes mitigate against this?
- Is it best to use a short-term supply of high-CT forages for reducing parasite burdens at strategic times (e.g., pregnancy, parturition, weaning), or should a longer-term supply of low-CT forages be used to boost the immune response?
- What agronomic, harvesting, or processing measures can best ensure that CT plants deliver consistent results? Which cultivars should be grazed and processed into hay, pellets, or silage, and at what times of the year?

From these questions we need to develop practical solutions through collaboration with farmers and veterinarians. Producers and consumers alike are looking for sustainable innovations that produce high-quality foods profitably while also maintaining soil fertility and the quality of our environment.

Conflict of Interest

The authors declare that there is no conflict of interest.

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References

Abeynayake, S.W., S. Panter, R. Chapman, T. Webster, S. Rochfort, A. Mouradov, and G. Spangenberg. 2012. Biosynthesis of proanthocyanidins in white clover flowers: Cross talk within the flavonoid pathway. Plant Physiol. 158:666–678. doi:10.1104/pp.111.189258

Agrawal, A.A., A. Hastings, M.T.J. Johnson, J.L. Maron, and J.-P. Salminen. 2012. Insect herbivores drive real-time ecological and evolutionary change in plant populations. Science 338:113–116.

Albert, N.W. 2015. Subspecialization of R2R3-MYB repressors for anthocyanin and proanthocyanidin regulation in forage legumes. Front. Plant Sci. 6:1165. doi:10.3389/fpls.2015.01165

Amiri, S., H. Karimmojeni, M.M. Majidi, and A. Boromand. 2013. Evaluation of post-emergence herbicides to control weeds of newly planted sainfoin (*Onobrychis sativa*). Plant Knowledge J. 2:145–149.

Attwood, G., D. Li, D. Pacheco, and M. Tavendale. 2006. Production of indolic compounds by rumen bacteria isolated from grazing ruminants. J. Appl. Microbiol. 100:1261–1271. doi:10.1111/j.1365-2672.2006.02896.x

Aufrère, J., M. Dudilieu, D. Andueza, C. Poncet, and R. Baumont. 2013. Mixing sainfoin and lucerne to improve the feed value of legumes fed to sheep by the effect of condensed tannins. Animal 7:82–92. doi:10.1017/S1751731112001097

Ayres, J.F., M.J. Blumenthal, L.A. Lane, and J.W. O'Connor. 2006. Birdsfoot trefoil (*Lotus corniculatus*) and greater lotus (*Lotus uliginosus*) in perennial pastures in eastern Australia 2. Adaptation and applications of lotus-based pasture. Aust. J. Exp. Agric. 46:521–534. doi:10.1071/EA04242

Azuhnwi, B.N., B. Boller, F. Dohme-Meier, H.D. Hess, M. Kreuzer, E. Stringano, and I. Mueller-Harvey. 2013a. Exploring variation in proanthocyanidin composition and content of sainfoin (*Onobrychis viciifolia*). J. Sci. Food Agric. 93:2102–2109. doi:10.1002/jsfa.6119

Azuhnwi, B.N., H. Hertzberg, Y. Arrigo, A. Gutzwiller, H.D. Hess, I. Mueller-Harvey, et al. 2013b. Investigation of sainfoin (*Onobrychis viciifolia*) cultivar differences on nitrogen balance and fecal egg count in artificially infected lambs. J. Anim. Sci. 91:2343–2354. doi:10.2527/jas.2012-5351

Baert, N., W.F. Pellikaan, M. Karonen, and J.-P. Salminen. 2016. A study of the structure-activity relationship of oligomeric ellagitannins on ruminal fermentation *in vitro*. J. Dairy Sci. 99:8041–8052. doi:10.3168/jds.2016-11069

Barbehenn, R.V., and C.P. Constabel. 2011. Tannins in plant-herbivore interactions. Phytochemistry 72:1551–1565. doi:10.1016/j.phytochem.2011.01.040

Barrau, E., N. Fabre, I. Fouraste, and H. Hoste. 2005. Effect of bioactive compounds from sainfoin (*Onobrychis viciifolia* Scop.) on the in vitro larval migration of *Haemonchus contortus*: Role of tannins and flavonol glycosides. Parasitology 131:531–538. doi:10.1017/S0031182005008024

Barry, T.N., T.R. Manley, and S.J. Duncan. 1986. The role of condensed tannins in the nutritional value of *Lotus pedunculatus* for sheep. 4. Sites of carbohydrate and protein digestion as influenced by dietary reactive tannin concentrations. Br. J. Nutr. 55:123–137. doi:10.1079/BJN19860016

Bee, G., P. Silacci, S. Ampuero-Kragten, M. Čandek-Potokar, A.L. Wealleans, J. Litten-Brown, J.-P. Salminem, and I. Mueller-Harvey. 2017. Hydrolysable tannin-based diet rich in gallotannins has a minimal impact on pig performance but significantly reduces salivary and bulbo-urethral gland size. Animal 11:1617–1625. doi:10.1017/S1751731116002597

Bhattarai, S., B. Coulman, and B. Biligetu. 2016. Sainfoin (*Onobrychis viciifolia* Scop.): Renewed interest as a forage legume for western Canada. Can. J. Plant Sci. 96:748–756. doi:10.1139/cjps-2015-0378

Berard, N.C., Y. Wang, K.M. Wittenberg, D.O. Krause, B.E. Coulman, T.A. McAllister, and K.H. Ominski. 2011. Condensed tannin concentrations found in vegetative and mature forage legumes grown in western Canada. Can. J. Plant Sci. 91:669–675. doi:10.4141/cjps10153

Bermingham, E.N., K.J. Hutchinson, D.K. Revell, I.M. Brookes, and W.C. McNabb. 2001. The effect of condensed tannins in sainfoin (*Onobrychis viciifolia*) and sulla (*Hedysarum coronarium*) on the digestion of amino acids in sheep. Proc. N.Z. Soc. Anim. Prod. 61:116–119.

Bidel, L.P.R., M. Coumans, Y. Baissae, P. Doumas, and C. Jay-Allemand. 2010. Biological activity of phenolics in plant cells. In: C. Santos-Buelga, M.T. Escribano-Bailon, and V. Lattanzio, editors, Recent advances in polyphenol research. Vol. 2. Blackwell Publishing, Oxford, UK. p. 163–205.

Brillouet, J.-M., H. Fulcrand, S. Carrillo, L. Rouméas, and C. Romieu. 2017. Isolation of native proanthocyanidins from grapevine (*Vitis vinifera*) and other fruits in aqueous buffer. J. Agric. Food Chem. 65:2895–2901. doi:10.1021/acs.jafc.6b05561

Brown, R.H., I. Mueller-Harvey, W.E. Zeller, L. Reinhardt, E. Stringano, A. Gea, et al. 2017. Facile purification of milligram to gram quantities of condensed tannins according to mean degree of polymerization and flavan-3-ol subunit composition. J. Agric. Food Chem. 65:8072–8082. doi:10.1021/acs.jafc.7b03489

Brunet, S., I. Fourquaux, and H. Hoste. 2011. Ultrastructural changes in the third-stage, infective larvae of ruminant nematodes treated with sainfoin (*Onobrychis viciifolia*) extract. Parasitol. Int. 60:419–424. doi:10.1016/j.parint.2010.09.011

Brunet, S., and H. Hoste. 2006. Monomers of condensed tannins affect the larval exsheathment of parasitic nematodes of ruminants. J. Agric. Food Chem. 54:7481–7487. doi:10.1021/jf0610007

Burns, J.C., W.A. Cope, and E.R. Barrick. 1977. Cow and calf performance, per hectare productivity, and persistence of crownvetch under grazing. Agron. J. 69:77–81. doi:10.2134/agronj1977.00021962006900010020x

Burns, J.C., R.D. Mochrie, and W.A. Cope. 1972. Responses of dairy heifers to crownvetch, sericea lespedeza, and alfalfa forages. Agron. J. 64:193–195. doi:10.2134/agronj1972.00021962006400020020x

Campidonico, L., P.G. Toral, A. Priolo, G. Luciano, B. Valenti, G. Hervás, et al. 2016. Fatty acid composition of ruminal digesta and longissimus muscle from lambs fed silage mixtures including red clover, sainfoin, and timothy. J. Anim. Sci. 94:1550–1560. doi:10.2527/jas.2015-9922

Chen, M., D.R. MacGregor, A. Dave, H. Florance, K. Moore, K. Paszkiewicz, et al. 2014. Maternal temperature history activates Flowering Locus T in fruits to control progeny dormancy according to time of year. Proc. Natl. Acad. Sci. USA 111:18787–18792. doi:10.1073/pnas.1412274111

Cherry, N.M., M. Bullinger, B.D. Lambert, J.P. Muir, T.W. Whitney, J.E. Miller, and J.T. Sawyer. 2014. Feeding panicled tick-clover to growing goats reduces *Haemonchus contortus* infection without negative effects on growth. J. Appl. Anim. Nutr. 2:e15. doi:10.1017/jan.2014.9

Cheynier, V., G. Bomte, K.M. Davies, V. Lattanzio, and S. Martens. 2013. Plant phenolics: Recent advances on their biosynthesis, genetics and ecophysiology. Plant Physiol. Biochem. 72:1–20. doi:10.1016/j.plaphy.2013.05.009

Cheynier, V., F.A. Tomas-Barberan, and K. Yoshida. 2015. Polyphenols: From plants to a variety of food and nonfood uses. J. Agric. Food Chem. 63:7589–7594. doi:10.1021/acs.jafc.5b01173

Chezem, W.R., and N.K. Clay. 2016. Regulation of plant secondary metabolism and associated specialized cell development by MYBs and bHLHs. Phytochemistry 131:26–43. doi:10.1016/j.phytochem.2016.08.006

Connolly, J., M.-T. Sebastià, L. Kirwan, J.A. Finn, R. Llurba, M. Suter, et al. 2017. Weed suppression greatly increased by plant diversity in intensively managed grasslands: A continental-scale experiment. J. Appl. Ecol. (in press). doi:10.1111/1365-2664.12991

Cooper, S.M., N. Owen-Smith, and J.P. Bryant. 1988. Foliage acceptability to browsing ruminants in relation to seasonal changes in the leaf chemistry of woody plants in a South African savanna. Oecologia 75:336–342.

Copani, G., V. Niderkorn, F. Anglard, A. Quereuil, and C. Ginane. 2016. Silages containing bioactive forage legumes: A promising protein-rich feed source for growing lambs. Grass Forage Sci. 71:622–631. doi:10.1111/gfs.12225

de Koning, C., D. Lloyd, S. Hughes, D. McLachlan, G. Crocker, S. Boschma, and A. Craig. 2003. Solutions for a better environment. In: Proceedings of the 11th Australian Agronomy Conference, Geelong, VIC. 2–6 Feb. 2003. [CD]. Aust. Soc. Agron.

de Koning, C., P. Schutz, and J. Howie. 2010. Sulla (*Hedysarum coronarium*) production sown with cover crops. In: Food security from sustainable agriculture: Proceedings of the 15th Australian Agronomy Conference, Lincoln, New Zealand. Nov. 2010. Aust. Soc. Agron. http://www.regional.org.au/au/asa/2010/pastures-forage/dryland-perennials/6987_dekoningct.htm (accessed 22 Nov 2016).

De Vega, J., S. Ayling, M.J. Hegarty, D. Kudrna, J.L. Goicoechea, Å. Ergon, et al. 2015. Red clover (*Trifolium pratense* L.) draft genome provides a platform for trait improvement. Sci. Rep. 5:17394. doi:10.1038/srep17394

Derksen, A., A. Hensel, W. Hafezi, F. Herrmann, T.J. Schmidt, C. Ehrhardt, et al. 2014. 3-*O*-galloylated procyanidins from *Rumex acetosa* L. inhibit the attachment of influenza A virus. PLoS One 9:e110089. doi:10.1371/journal.pone.0110089

Desrues, O., C. Fryganas, H.M. Ropiak, I. Mueller-Harvey, H.L. Enemark, and S.M. Thamsborg. 2016a. Impact of chemical structure of flavanol monomers and condensed tannins on in vitro anthelmintic activity against bovine nematodes. Parasitology 143:444–454. doi:10.1017/S0031182015001912

Desrues, O., I. Mueller-Harvey, W.F. Pellikaan, H.L. Enemark, and S.M. Thamsborg. 2017. Condensed tannins in the gastrointestinal tract of cattle after sainfoin (*Onobrychis viciifolia*) intake and their possible relationship with anthelmintic effects. J. Agric. Food Chem. 65:1420–1427. doi:10.1021/acs.jafc.6b05830

Desrues, O., M. Peña-Espinoza, T.V.A. Hansen, H.L. Enemark, and S.M. Thamsborg. 2016b. Anti-parasitic activity of pelleted sainfoin (*Onobrychis viciifolia*) against *Ostertagia ostertagi* and *Cooperia oncophora* in calves. Parasit. Vectors 9:329. doi:10.1186/s13071-016-1617-z

Dobreva, M.A., E. Stringano, R.A. Frazier, R.J. Green, and I. Mueller-Harvey. 2012. Interaction of sainfoin (*Onobrychis viciifolia*) condensed tannins and proteins. In: V. Lattanzio, N. Mulinacci, P. Pinelli, A. Romani, editors, Proceedings of the 26th International Conference on Polyphenols, Florence, Italy. 23–26 Jul 2012. Stabilimento Grafico Rindi, Prato, Italy. p. 151–152.

Döring, T.F., J.A. Baddeley, R. Brown, R. Collins, O. Crowley, S. Cuttle, et al. 2013. Using legume-based mixtures to enhance the nitrogen use efficiency and economic viability of cropping systems. Agric. Hort. Dev. Board. https://www.ruralbusinessschool.org.uk/sites/default/files/attachments/legume_link.pdf (accessed 30 Sept. 2017).

Dykes, L., L. Hoffmann, Jr., O. Portillo-Rodriguez, W.L. Rooney, and L.W. Rooney. 2014. Prediction of total phenols, condensed tannins, and 3-deoxyanthocyanidins in sorghum grain using near-infrared (NIR) spectroscopy. J. Cereal Sci. 60:138–142. doi:10.1016/j.jcs.2014.02.002

Egan, A.R., and M.J. Ulyatt. 1980. Quantitative digestion of fresh herbage by sheep. VI. Utilization of nitrogen in five herbages. J. Agric. Sci. 94:47–56. doi:10.1017/S002185960002788X

Engström, M.T., M. Karonen, J.R. Ahern, N. Baert, B. Payré, H. Hoste, and J.-P. Salminen. 2016. Chemical structures of plant hydrolyzable tannins reveal their in vitro activity against egg hatching and motility of *Haemonchus contortus* nematodes. J. Agric. Food Chem. 64:840–851. doi:10.1021/acs.jafc.5b05691

Engström, M.T., M. Pälijärvi, C. Fryganas, J.H. Grabber, I. Mueller-Harvey, and J.-P. Salminen. 2014. Rapid qualitative and quantitative analyses of proanthocyanidin oligomers and polymers by UPLC-MS/MS. J. Agric. Food Chem. 62:3390–3399. doi:10.1021/jf500745y

Escaray, F.J., V. Passeri, F.M. Babuin, F. Marco, P. Carrasco, F. Damiani, et al. 2014. *Lotus tenuis* × *L. corniculatus* interspecific hybridization as a means to breed bloat-safe pastures and gain insight into the genetic control of proanthocyanidin biosynthesis in legumes. BMC Plant Biol. 14:40. doi:10.1186/1471-2229-14-40

Feliciano, R.P., C.G. Krueger, D. Shanmuganayagam, M.M. Vestling, and J.D. Reed. 2012. Deconvolution of matrix-assisted laser desorption/ionization time-of-flight mass spectrometry isotope patterns to determine ratios of A-type to B-type interflavan bonds in cranberry proanthocyanidins. Food Chem. 135:1485–1493. doi:10.1016/j.foodchem.2012.05.102

Ferreyra, M.L.F., S.P. Rius, and P. Casati. 2012. Flavonoids: Biosynthesis, biological functions, and biotechnological applications. Front. Plant Sci. 3:222. doi:10.3389/fpls.2012.00222

Feucht, W., D. Treutter, H. Dithmar, and J. Polster. 2013. Loss of nuclear flavanols during drought periods in *Taxus baccata*. Plant Biol. (Berlin, Ger.) 15:462–470. doi:10.1111/j.1438-8677.2012.00661.x

Finn, J.A., L. Kirwan, J. Connolly, M.T. Sebastià, À. Helgadóttir, O.H. Baadshaug, et al. 2013. Ecosystem function enhanced by combining four functional types of plant species in intensively managed grassland mixtures: A 3-year continental-scale field experiment. J. Appl. Ecol. 50:365–375. doi:10.1111/1365-2664.12041

Frame, J., J. Charlton, and A. Laidlaw. 1998. Temperate forage legumes. CAB Int., Wallingford, UK.

Fryganas, C. 2016. Investigations into the chemical analysis and bioactivity of plant proanthocyanidins to support sustainable livestock farming. Ph.D. diss., Univ. of Reading, UK.

Gea, A., E. Stringano, R.H. Brown, and I. Mueller-Harvey. 2011. In situ analysis and structural elucidation of sainfoin (*Onobrychis viciifolia*) tannins for high throughput germplasm screening. J. Agric. Food Chem. 59:495–503. doi:10.1021/jf103609p

Gebrehiwot, L., P.R. Beuselinck, and C.A. Roberts. 2002. Seasonal variations in condensed tannin concentration of three *Lotus* species. Agron. J. 94:1059–1065. doi:10.2134/agronj2002.1059

Girard, M. 2016. Bioactive compounds in forage legumes: Structural changes during conservation, their fate along the digestive tract and their potential to impact ruminant products. Ph.D. diss., ETH Zurich, Zurich, Switzerland.

Girard, M., F. Dohme-Meier, P. Silacci, S. Ampuero Kragten, M. Kreuzer, and G. Bee. 2016a. Forage legumes rich in condensed tannins may increase n-3 fatty acid levels and sensory quality of lamb meat. J. Sci. Food Agric. 96:1923–1933. doi:10.1002/jsfa.7298

Girard, M., F. Dohme-Meier, D. Wechsler, D. Goy, M. Kreuzer, and G. Bee. 2016b. Ability of 3 tanniferous forage legumes to modify quality of milk and Gruyère-type cheese. J. Dairy Sci. 99:205–220. doi:10.3168/jds.2015-9952

Givens, D.I., E. Owen, and A.T. Adesogan. 2000. Current procedures, future requirements and the need for standardisation. In: D.I. Givens, E. Owen, R.F.E. Axford, and H. Omed, editors, Forage evaluation in ruminant nutrition. CABI Publishing, Wallingford, UK. p. 449–474. doi:10.1079/9780851993447.0449

Grabber, J., W.E. Zeller, and I. Mueller-Harvey. 2013. Acetone enhances the direct analysis of procyanidin- and prodelphinidin-based condensed tannins in *Lotus* species by the butanol-HCl-iron assay. J. Agric. Food Chem. 61:2669–2678. doi:10.1021/jf304158m

Grabber, J.H., W.K. Coblentz, H. Riday, T.C. Griggs, D.H. Min, J.W. MacAdam, and K.A. Cassida. 2015. Protein and dry-matter degradability of European- and Mediterranean-derived birdsfoot trefoil cultivars grown in the colder continental USA. Crop Sci. 55:1356–1364. doi:10.2135/cropsci2014.09.0659

Grabber, J.H., H. Riday, K.A. Cassida, T.C. Griggs, D.H. Min, and J.W. MacAdam. 2014. Yield, morphological characteristics, and chemical composition of European- and Mediterranean-derived birdsfoot trefoil cultivars grown in the colder continental United States. Crop Sci. 54:1893–1901. doi:10.2135/cropsci2013.09.0644

Grosse Brinkhaus, A., G. Bee, P. Silacci, M. Kreuzer, and F. Dohme-Meier. 2016. Effect of exchanging *Onobrychis viciifolia* and *Lotus corniculatus* for *Medicago sativa* on ruminal fermentation and nitrogen turnover in dairy cows. J. Dairy Sci. 99:4384–4397. doi:10.3168/jds.2015-9911

Grosse Brinkhaus, A., U. Wyss, Y. Arrigo, M. Girard, G. Bee, J.O. Zeitz, et al. 2017. In vitro ruminal fermentation characteristics and utilisable crude protein supply of sainfoin and birdsfoot trefoil silages and their mixtures with other legumes. Animal 11:580–590. doi:10.1017/S1751731116001816

Gruber, M., B. Skadhauge, M. Yu, A. Muir, and K. Richards. 2008. Variation in morphology, plant habit, proanthocyanidins, and flavonoids within a *Lotus* germplasm collection. Can. J. Plant Sci. 88:121–132. doi:10.4141/P06-158

Guyot, S., N. Marnet, and J.-F. Drilleau. 2001a. Thiolysis-HPLC characterization of apple procyanidins covering a large range of polymerization states. J. Agric. Food Chem. 49:14–20. doi:10.1021/jf000814z

Guyot, S., N. Marnet, P. Sanoner, and J.F. Drilleau. 2001b. Direct thiolysis on crude apple materials for high-performance liquid chromatography characterization and quantification of polyphenols in cider apple tissues and juices. Methods Enzymol. 335:57–70. doi:10.1016/S0076-6879(01)35231-X

Hagerman, A.E., and L.G. Butler. 1981. The specificity of proanthocyanidin-protein interactions. J. Biol. Chem. 256:4444–4497.

Hammond, K.J., J.L. Burke, J.P. Koolaard, S. Muetzel, C.S. Pinares-Patiño, and G.C. Waghorn. 2013. Effects of feed intake on enteric methane emissions from sheep fed fresh white clover (*Trifolium repens*) and perennial ryegrass (*Lolium perenne*) forages. Anim. Feed Sci. Technol. 179:121–132. doi:10.1016/j.anifeedsci.2012.11.004

Hancock, K., V. Collette, E. Chapman, K. Hanson, S. Temple, R. Moraga, and J. Caradus. 2014. Progress towards developing bloat-safe legumes for the farming industry. Crop Pasture Sci. 65:1107–1113. doi:10.1071/CP13308

Hancock, K.R., V. Collette, K. Fraser, M. Greig, H. Xue, K. Richardson, et al. 2012. Expression of the R2R3-MYB transcription factor TaMYB14 from *Trifolium arvense* activates proanthocyanidin biosynthesis in the legumes *Trifolium repens* and *Medicago sativa*. Plant Physiol. 159:1204–1220. doi:10.1104/pp.112.195420

Hansen, T.V.A., C. Fryganas, N. Acevedo, L.R. Carballo, S.M. Thamsborg, I. Mueller-Harvey, and A.R. Williams. 2016. Proanthocyanidins inhibit *Ascaris suum* glutathione-S-transferase activity and increase susceptibility of larvae to levamisole *in vitro*. Parasitol. Int. 65:336–339. doi:10.1016/j.parint.2016.04.001

Harding, S.A., L.-J. Xue, L. Du, B. Nyamdari, R.L. Lindroth, R. Sykes, et al. 2013. Condensed tannin biosynthesis and polymerization synergistically condition carbon use, defense, sink strength and growth in *Populus*. Tree Physiol. 34:1240–1251. doi:10.1093/treephys/tpt097

Häring, D.A., A. Scharenberg, F. Heckendorn, F. Dohme, A. Lüscher, V. Maurer, et al. 2008. Tanniferous forage plants: Agronomic performance, palatability and efficacy against parasitic nematodes in sheep. Renew. Agric. Food Syst. 23:19–29. doi:10.1017/S1742170507002049

Haslam, E. 1981. Vegetable tannins. In: E.E. Conn, editor, The biochemistry of plants. Vol. 7. Academic Press, New York. p. 527–556.

Hatew, B., E. Stringano, I. Mueller-Harvey, W.H. Hendriks, C. Hayot Carbonero, L.M.J. Smith, and W.F. Pellikaan. 2016. Impact of variation in structure of condensed tannins from sainfoin (*Onobrychis viciifolia*) on in vitro ruminal methane production and fermentation characteristics. J. Anim. Physiol. Anim. Nutr. 100:348–360. doi:10.1111/jpn.12336

Hayot Carbonero, C. 2011. Sainfoin (*Onobrychis viciifolia*), a forage legume with great potential for sustainable agriculture, an insight on its morphological, agronomical, cytological and genetic characterisation. Ph.D. diss., Manchester, UK. https://www.escholar.manchester.ac.uk/api/datastream?publicationPid=uk-ac-man-scw:120238&datastreamId=FULL-TEXT.PDF (accessed 18 Nov. 2017).

Hayot Carbonero, C., I. Mueller-Harvey, T.A. Brown, and L. Smith. 2011. Sainfoin (*Onobrychis viciifolia*): A beneficial forage legume. Plant Genet. Resour. 9:70–85. doi:10.1017/S1479262110000328

Heckendorn, F., D.A. Häring, V. Maurer, J. Zinsstag, W. Langhans, and H. Hertzberg. 2006. Effect of sainfoin (*Onobrychis viciifolia*) silage and hay on established populations of *Haemonchus contortus* and *Cooperia curticei* in lambs. Vet. Parasitol. 142:293–300. doi:10.1016/j.vetpar.2006.07.014

Hellström, J.K., and P.H. Mattila. 2008. HPLC determination of extractable and unextractable proanthocyanidins in plant materials. J. Agric. Food Chem. 56:7617–7624. doi:10.1021/jf801336s

Henning, S.M., C. Fajardo-Lira, H.W. Lee, A.A. Youssefian, V.L.W. Go, and D. Heber. 2003. Catechin content of 18 teas and a green tea extract supplement correlates with the antioxidant capacity. Nutr. Cancer 45:226–235. doi:10.1207/S15327914NC4502_13

Hernes, P.J., and J.I. Hedges. 2004. Tannin signatures of barks, needles, leaves, cones, and wood at the molecular level. Geochim. Cosmochim. Acta 68:1293–1307. doi:10.1016/j.gca.2003.09.015

Heuzé, V., G. Tran, and F. Lebas. 2015. Sulla *(Hedysarum coronarium)*. Feedipedia, a programme by Inst. Natl. Rech. Agron., Ctr. Coop. Int. Rech. Agron. Dév., Assoc. Fran. Zootech., and FAO. http://www.feedipedia.org/node/292 (acccessed 18 Nov. 2017).

Hixson, J.L., K.A. Bindon, and P.A. Smith. 2015. Evaluation of direct phloroglucinolysis and colorimetric depolymerization assays and their applicability for determining condensed tannins in grape marc. J. Agric. Food Chem. 63:9954–9962. doi:10.1021/acs.jafc.5b04207

Hixson, J.L., J.L. Jacobs, E.N. Wilkes, and P.A. Smith. 2016. Survey of the variation in grape marc condensed tannin composition and concentration and analysis of key compositional factors. J. Agric. Food Chem. 64:7076–7086. doi:10.1021/acs.jafc.6b03126

Hoste, H., C. Martinez-Ortiz-De-Montellano, F. Manolaraki, S. Brunet, N. Ojeda-Robertos, I. Fourquaux, et al. 2012. Direct and indirect effects of bioactive tannin-rich tropical and temperate legumes against nematode infections. Vet. Parasitol. 186:18–27. doi:10.1016/j.vetpar.2011.11.042

Hoste, H., S. Sotiraki, S.Y. Landau, F. Jackson, and I. Beveridge. 2010. Goat nematode interactions: Think differently! Trends Parasitol. 26:376–381. doi:10.1016/j.pt.2010.04.007

Hoste, H., J.F.J. Torres-Acosta, J. Quijada, I. Chan-Perez, M.M. Dakheel, D.S. Kommuru, et al. 2016. Interactions between nutrition and infections with *Haemonchus contortus* and related gastrointestinal nematodes in small ruminants. In: R.B. Gasser and G. von Samson-Himmelstjerna, editors, Advances in parasitology. Vol. 93. *Haemonchus contortus* and Haemonchosis: Past, present and future trends. Academic Press, New York. p. 239–351.

Hoste, H., J.F.J. Torres-Acosta, C.A. Sandoval-Castro, I. Mueller-Harvey, S. Sotiraki, H. Louvandini, et al. 2015. Tannin containing legumes as a model for nutraceuticals against digestive parasites in livestock. Vet. Parasitol. 212:5–17. doi:10.1016/j.vetpar.2015.06.026

Huang, Q.Q., L. Jin, Z. Xu, S. Acharya, T.A. McAllister, T.M. Hu, et al. 2016. Effects of conservation method on condensed tannin content, ruminal degradation, and in vitro intestinal digestion of purple prairie clover (*Dalea purpurea* Vent.). Can. J. Anim. Sci. 96:524–531. doi:10.1139/cjas-2016-0006

Hümmer, A., and P. Schreier. 2008. Analysis of proanthocyanidins. Mol. Nutr. Food Res. 52:1381–1398. doi:10.1002/mnfr.200700463

Hunt, S.R., J.W. MacAdam, and T.C. Griggs. 2016. Seeding rate, oat companion crop, and planting season effects on irrigated organic birdsfoot trefoil stands in the mountain West United States. Crop Sci. 56:463–473. doi:10.2135/cropsci2015.04.0240

Huyen, N.T. 2016. Sainfoin (*Onobrychis viciifolia*): A forgotten crop for dairy cows with future potential. Ph.D. diss., Wageningen Univ., Wageningen, the Netherlands.

Huyen, N.T., O. Desrues, S.J.J. Alferink, T. Zandstra, M.W.A. Verstegen, W.H. Hendriks, and W.F. Pellikaan. 2016a. Inclusion of sainfoin (*Onobrychis viciifolia*) silage in dairy cow rations affects nutrient digestibility, nitrogen utilization, energy balance, and methane emissions. J. Dairy Sci. 99:3566–3577. doi:10.3168/jds.2015-10583

Huyen, N.T., C. Fryganas, G. Uittenbogaard, I. Mueller-Harvey, M.W.A. Verstegen, W.H. Hendriks, and W.F. Pellikaan. 2016b. Structural features of condensed tannins affect *in vitro* ruminal methane production and fermentation characteristics. J. Agric. Sci. 154:1474–1487. doi:10.1017/S0021859616000393

Huynh-Ba, T., T. Nguyen, G. Philippossian, and A. Vanstraceele. 1994. Anti-urease cosmetic or dermatological composition. US Patent CA2101234 A1. Date issued: 4 February.

Irani, S., M.M. Majidi, A. Mirlohi, M. Karami, and M. Zargar. 2015. Response to drought stress in sainfoin: Within and among ecotype variation. Crop Sci. 55:1868–1880. doi:10.2135/cropsci2014.07.0481

Jansman, A.J.M. 1993. Tannins in faba beans (*Vicia faba* L.). Ph.D. diss., Wageningen Univ., Wageningen, the Netherlands.

Jones, W.T., and J.L. Mangan. 1977. Complexes of the condensed tannins of sainfoin (*Onobrychis viciifolia* Scop.) with Fraction 1 leaf protein and with submaxillary mucoprotein, and their reversal by polyethylene glycol and pH. J. Sci. Food Agric. 28:126–136. doi:10.1002/jsfa.2740280204

Kariuki, I.W., and B.W. Norton. 2008. The digestion of dietary protein bound by condensed tannins in the gastro-intestinal tract of sheep. Anim. Feed Sci. Technol. 142:197–209. doi:10.1016/j.anifeedsci.2007.08.006

Karnezos, T.P., A.G. Matches, and C.P. Brown. 1994. Spring lamb production on alfalfa, sainfoin, and wheatgrass pastures. Agron. J. 86:497–502. doi:10.2134/agronj1994.00021962008600030008x

Kempf, K., C.S. Malisch, C. Grieder, F. Widmer, and R. Kölliker. 2017. Marker-trait association analysis for agronomic and compositional traits in sainfoin (*Onobrychis viciifolia*). Genet. Mol. Res. 16:Gmr16019483. doi:10.4238/gmr16019483

Kempf, K., M. Mora-Ortiz, L.M.J. Smith, R. Kölliker, and L. Skøt. 2016. Characterization of novel SSR markers in diverse sainfoin (*Onobrychis viciifolia*) germplasm. BMC Genet. 17:124. doi:10.1186/s12863-016-0431-0

Khanbabaee, K., and T. van Ree. 2001. Tannins: Classification and definition. Nat. Prod. Rep. 18:641–649. doi:10.1039/b101061l

Kingston-Smith, A.H., J.E. Edwards, S.A. Huws, E.J. Kim, and M. Abberton. 2010. Plant-based strategies towards minimising 'livestock's long shadow'. Proc Nutr. Soc. 69:613–620. doi:10.1017/S0029665110001953

Klongsiriwet, C. 2016. Phytochemical analysis of a willow germplasm collection and its pharmacological activities. PhD diss., Univ. of Reading, Reading, UK.

Klongsiriwet, C., A. Karp, S. Hanley, R. Barnes, L. Falchero, and I. Mueller-Harvey. 2013. Near infrared spectroscopy (NIRS) for screening condensed tannins in a willow germplasm collection. Planta Med. 79:PJ26. doi:10.1055/s-0033-1352230

Klongsiriwet, C., J. Quijada, A.R. Williams, I. Mueller-Harvey, E.M. Williamson, and H. Hoste. 2015. Synergistic inhibition of *Haemonchus contortus* exsheathment by flavonoid monomers and condensed tannins. Int. J. Parasitol. Drugs Drug Resist. 5:127–134. doi:10.1016/j.ijpddr.2015.06.001

Kölliker, R., K. Kempf, C.S. Malisch, and A. Lüscher. 2017. Promising options for improving performance and proanthocyanidins of the forage legume sainfoin (*Onobrychis viciifolia* Scop.). Euphytica 213:179. doi:10.1007/s10681-017-1965-6

Kommuru, D.S., T. Barker, S. Desai, J.M. Burke, A. Ramsay, I. Mueller-Harvey, et al. 2014. Use of pelleted sericea lespedeza (*Lespedeza cuneata*) for natural control of coccidia and gastrointestinal nematodes in weaned goats. Vet. Parasitol. 204:191–198. doi:10.1016/j.vetpar.2014.04.017

Kommuru, D.S., N.C. Whitley, J.E. Miller, J.A. Mosjidis, J.M. Burke, S. Gujja, et al. 2015. Effect of sericea lespedeza leaf meal pellets on adult female *Haemonchus contortus* in goats. Vet. Parasitol. 207:170–175. doi:10.1016/j.vetpar.2014.11.008

Koupai-Abyazani, M.R., J.J. McCallum, A.D. Muir, G.L. Lees, B.A. Bohm, G.H.N. Towers, and M.Y. Gruber. 1993. Purification and characterization of a proanthocyanidin polymer from seed of alfalfa (*Medicago sativa* Cv. Beaver). J. Agric. Food Chem. 41:565–569. doi:10.1021/jf00028a012

Krueger, C.G., J.D. Reed, R.P. Feliciano, and A.B. Howell. 2013. Quantifying and characterizing proanthocyanidins in cranberries in relation to urinary tract health. Anal. Bioanal. Chem. 405:4385–4395. doi:10.1007/s00216-013-6750-3

Laaksonen, O.A., J.-P. Salminen, L. Makila, H.P. Kallio, and B. Yang. 2015. Proanthocyanidins and their contribution to sensory attributes of black currant juices. J. Agric. Food Chem. 63:5373–5380. doi:10.1021/acs.jafc.5b01287

Landau, S., H. Azaizeh, H. Muklada, T. Glasser, E.D. Ungar, H. Baram, et al. 2010. Anthelmintic activity of *Pistacia lentiscus* foliage in two Middle Eastern breeds of goats differing in their propensity to consume tannin-rich browse. Vet. Parasitol. 173:280–286. doi:10.1016/j.vetpar.2010.07.006

Larkin, P.J., G.J. Tanner, R.G. Joseph, and W.M. Kelman. 1997. Modifying condensed tannin content in plants. In: J.G. Buchanan-Smith, L.D. Bailey, and P. McCaughey, editors, Proceedings of the 18th International Grasslands Congress, Winnipeg, MB, and Saskatoon, SK. 9–18 June 1997. Int. Grassl. Congr., Lexington, KY. p. 167–178. http://www.internationalgrasslands.org/files/igc/publications/1997/iii-167.pdf (accessed 30 Sept. 2017).

Lattanzio, V., A. Cardinali, and V. Linsalata. 2012. Plant phenolics: A biochemical and physiological perspective. In: V. Cheynier, P. Sarni-Manchado, and S. Quideau, editors, Recent advances in polyphenol research. Vol. 3. John Wiley & Sons, Chichester, UK. p. 1–39.

Lee, L.-S., S.-H. Kim, Y.-B. Kim, and Y.-C. Kim. 2014. Quantitative analysis of major constituents in green tea with different plucking periods and their antioxidant activity. Molecules 19:9173–9186. doi:10.3390/molecules19079173

Lehmann, J.R.K., A. Große-Stoltenberg, M. Römer, and J. Oldeland. 2015. Field spectroscopy in the VNIR-SWIR region to discriminate between Mediterranean native plants and exotic-invasive shrubs based on leaf tannin content. Remote Sens. 7:1225–1241. doi:10.3390/rs70201225

Li, C., R. Leverence, J.D. Trombley, S. Xu, J. Yang, Y. Tian, et al. 2010. High molecular weight persimmon (*Diospyros kaki* L.) proanthocyanidin: A highly galloylated, A-linked tannin with an unusual flavonol terminal unit, myricetin. J. Agric. Food Chem. 58:9033–9042. doi:10.1021/jf102552b

Li, M., and A.E. Hagerman. 2013. Interactions between plasma proteins and naturally occurring polyphenols. Curr. Drug Metab. 14:432–445. doi:10.2174/1389200211314040006

Li, Y., A.D. Iwaasa, Y. Wang, L. Jin, G. Han, and M. Zhao. 2014. Condensed tannins concentration of selected prairie legume forages as affected by phenological stages during two consecutive growth seasons in western Canada. Can. J. Plant Sci. 94:817–826. doi:10.4141/cjps2013-234

Liimatainen, J., M. Karonen, and J. Sinkkonen. 2012. Procyanidin xylosides from the bark of *Betula pendula*. Phytochemistry 76:178–183. doi:10.1016/j.phytochem.2012.01.008

Liu, Z., G.P.F. Lane, and W.P. Davies. 2008. Establishment and production of common sainfoin (*Onobrychis viciifolia* Scop.) in the UK. 2. Effects of direct sowing and undersowing in spring barley on sainfoin and sainfoin-grass mixtures. Grass Forage Sci. 63:242–248. doi:10.1111/j.1365-2494.2008.00629.x

Lorenz, M.M., T. Eriksson, and P. Udén. 2010. Effect of wilting, silage additive, PEG treatment and tannin content on the distribution of N fractions after ensiling of three different sainfoin (*Onobrychis viciifolia*) varieties. Grass Forage Sci. 65:175–184. doi:10.1111/j.1365-2494.2010.00736.x

Lüscher, A., I. Mueller-Harvey, J.F. Soussana, R.M. Rees, and J.L. Peyraud. 2014. Potential of legume-based grassland-livestock systems in Europe. Grass Forage Sci. 69:206–228. doi:10.1111/gfs.12124

MacAdam, J.W., and J.J. Villalba. 2015. Beneficial effects of temperate forage legumes that contain condensed tannins. Agriculture 5:475–491. doi:10.3390/agriculture5030475

MacAdam, J.W., R.E. Ward, T.C. Griggs, B.R. Min, and G.E. Aiken. 2011. Case study: Average daily gain and blood fatty acid composition of cattle grazing the nonbloating legumes birdsfoot trefoil and cicer milkvetch in the Mountain West. Prof. Anim. Sci. 27:574–583. doi:10.15232/S1080-7446(15)30542-8

Malisch, C., B. Studer, D. Suter, J.-P. Salminen, and A. Lüscher. 2014. Phenotypic variation among and within thirty accessions of *Onobrychis viciifolia* examined under climate change scenarios. In: Proceedings of the ASA, CSSA, and SSSA International Annual Meeting, Long Beach, CA. 2–5 Nov. 2014. ASA, CSSA, SSSA, Madison, WI. Paper 125-14. https://scisoc.confex.com/scisoc/2014am/webprogram/Paper88495.html (accessed 18 Nov. 2017).

Malisch, C.S., A. Lüscher, N. Baert, M.T. Engström, B. Studer, C. Fryganas, et al. 2015. Large variability of proanthocyanidin content and composition in sainfoin (*Onobrychis viciifolia*). J. Agric. Food Chem. 63:10234–10242. doi:10.1021/acs.jafc.5b04946

Malisch, C.S., J.-P. Salminen, R. Kölliker, M.T. Engström, D. Suter, B. Studer, and A. Lüscher. 2016. Drought effects on proanthocyanidins in sainfoin (*Onobrychis viciifolia* Scop.) are dependent on the plant's ontogenetic stage. J. Agric. Food Chem. 64:9307–9316. doi:10.1021/acs.jafc.6b02342

Malisch, C.S., D. Suter, B. Studer, and A. Lüscher. 2017. Multifunctional benefits of sainfoin mixtures: Effects of partner species, sowing density and cutting regime. Grass Forage Sci. (in press). doi:10.1111/gfs.12278

Manolaraki, F. 2011. Propriétés anthelminthiques du sainfoin (*Onobrychis viciifoliae*): Analyse des facteurs de variations et du rôle des composés phénoliques impliqués. (In French.) Ph.D. diss., Univ. of Toulouse, Toulouse, France. http://ethesis.inp-toulouse.fr/archive/00001566/01/manolaraki.pdf (accessed 30 Sept. 2017).

Martínez-Ortíz-de-Montellano, C., C. Arroyo-López, I. Fourquaux, J.F.J. Torres-Acosta, C.A. Sandoval-Castro, and H. Hoste. 2013. Scanning electron microscopy of *Haemonchus contortus* exposed to tannin-rich plants under in vivo and in vitro conditions. Exp. Parasitol. 133:281–286. doi:10.1016/j.exppara.2012.11.024

Maughan, B., F.D. Provenza, R. Tansawat, C. Maughan, S. Martini, R. Ward, et al. 2014. Importance of grass-legume choices on cattle grazing behavior, performance, and meat characteristics. J. Anim. Sci. 92:2309–2324. doi:10.2527/jas.2013-7297

McCaslin, M., P. Reisen, and J. Ho. 2014. New strategies for forage quality improvement in alfalfa. In: Proceedings of the 2014 California Alfalfa, Forage, and Grain Symposium, Long Beach, CA, 10–12 Dec. 2014. Univ. of California Coop. Ext., Plant Sci. Dep., Univ. of California, Davis, CA. http://alfalfa.ucdavis.edu/+symposium/proceedings/2014/14CAS07_McCaslin_AlfalfaImprovement.pdf (accessed 30 Sept. 2017).

Meagher, L.P., G. Lane, S. Sivakumaran, M.H. Tavendale, and K. Fraser. 2004. Characterization of condensed tannins from *Lotus* species by thiolytic degradation and electrospray mass spectrometry. Anim. Feed Sci. Technol. 117:151–163. doi:10.1016/j.anifeedsci.2004.08.007

Mechineni, A., D.S. Kommuru, S. Gujja, J.A. Mosjidis, J.E. Miller, J.M. Burke, et al. 2014. Effect of fall-grazed sericea lespedeza (*Lespedeza cuneata*) on gastrointestinal nematode infections, skin and carcass microbial load, and meat quality of growing goats. Vet. Parasitol. 204:221–228. doi:10.1016/j.vetpar.2014.06.002

Mena, P., L. Calani, R. Bruni, and D. Del Rio. 2015. Bioactivation of high-molecular-weight polyphenols by the gut microbiome. In: K.T.D. Rio, editor, Diet-microbe interactions in the gut. Academic Press, San Diego, CA. p. 73–101. doi:10.1016/B978-0-12-407825-3.00006-X

Mengistu, G., H. Hoste, M. Karonen, J.-P. Salminen, W.H. Hendriks, and W.F. Pellikaan. 2017. The in vitro anthelmintic properties of browse plant species against *Haemonchus contortus* is determined by the polyphenol content and composition. Vet. Parasitol. 237:110–116. doi:10.1016/j.vetpar.2016.12.020

Min, B.R., T.N. Barry, G.T. Attwood, and W.C. McNabb. 2003. The effect of condensed tannins on the nutrition and health of ruminants fed fresh temperate forages: A review. Anim. Feed Sci. Technol. 106:3–19. doi:10.1016/S0377-8401(03)00041-5

Minnée, E.M.K., S.L. Woodward, G.C. Waghorn, and P.G. Laboyrie. 2002. The effect of ensiling forage legumes on condensed tannins. Agron. N. Z. 32:117–119.

Mora-Ortiz, M. 2015. Pre-breeding, functional genomics and agronomic improvement of the tanniniferous forage crop *Onobrychis viciifolia*. Ph.D. diss., Univ. of Reading, Reading, UK.

Mora-Ortiz, M., and L. Smith. 2016. Sainfoin: Surprising science behind a forgotten forage. Cotswold Grass Seeds, Moreton-in-Marsh, UK.

Mora-Ortiz, M., M.T. Swain, M.J. Vickers, M.J. Hegarty, R. Kelly, L.M.J. Smith, and L. Skøt. 2016. De-novo transcriptome assembly for gene identification, analysis, annotation, and molecular marker discovery in *Onobrychis viciifolia*. BMC Genomics 17:756. doi:10.1186/s12864-016-3083-6

Mosjidis, J.A. 2001. Forage legume breeding and evaluation at Auburn University in the last 16 years. In: Proceedings of the 56th Southern Pasture and Forage Crop Improvement Conference, Springdale, AR. 21–22 Apr. 2001. Texas A&M Agrilife Ext., College Station. http://agrilife.org/spfcic/annual-proceedings/56th/forage-legume-breeding-and-evaluation/ (accessed 30 Sept. 2017).

Mouradov, A., and G. Spangenberg. 2014. Flavonoids: A metabolic network mediating plants adaptation to their real estate. Front. Plant Sci. 5:620. doi:10.3389/fpls.2014.00620

Moyer, J.R., R. Hironaka, G.C. Kozub, and P. Bergen. 1990. Effect of herbicide treatments on dandelion, alfalfa and sainfoin yields and quality. Can. J. Plant Sci. 70:1105–1113. doi:10.4141/cjps90-133

Mueller-Harvey, I. 2006. Unravelling the conundrum of tannins in animal nutrition and health. J. Sci. Food Agric. 86:2010–2037.

Mueller-Harvey, I., M. Martin-Lorenzo, E. Stringano, R.J. Barnes, J. Oliver, K. Theodoridou, et al. 2011. NIR spectroscopy for predicting the nutritional, anthelmintic and environmental effects of sainfoin. Proceedings of the 8th International Symposium on the Nutrition of Herbivores (ISNH8), 'Herbivores in a Changing World'; Aberystwyth, UK, 6–9 Sep 2011. Adv. Anim. Biosci. 2:265.

Muir, J.P., T.H. Terrill, J.A. Mosjidis, J.-M. Luginbuhl, J.E. Miller, and J.M. Burke. 2017. Season progression, ontogenesis and environment affect *Lespedeza cuneata* herbage condensed tannin, fiber and crude protein content. Crop Sci. 57:515–524. doi:10.2135/cropsci2016.07.0605

Naumann, H.D., J.P. Muir, B.D. Lambert, L.O. Tedeschi, and M.M. Kothmann. 2013. Condensed tannins in the ruminant environment: A perspective on biological activity. J. Agric. Sci. 1:8–20.

Neilson, E.H., J.Q.D. Goodger, I.E. Woodrow, and B. Lindberg Moller. 2013. Plant chemical defense: At what cost? Trends Plant Sci. 18:250–258. doi:10.1016/j.tplants.2013.01.001

Niezen, J.H., W.A.G. Charleston, H.A. Robertson, D. Shelton, G.C. Waghorn, and R. Green. 2002. The effect of feeding sulla (*Hedysarum coronarium*) or lucerne (*Medicago sativa*) on lamb parasite burdens and development of immunity to gastro intestinal nematodes. Vet. Parasitol. 105:229–245. doi:10.1016/S0304-4017(02)00014-6

Niezen, J.H., G.C. Waghorn, and W.A.G. Charleston. 1998a. Establishment and fecundity of *Ostertagia circumcincta* and *Trichostrongylus colubriformis* in lambs fed lotus (*Lotus pedunculatus*) or perennial ryegrass (*Lolium perenne*). Vet. Parasitol. 78:13–21. doi:10.1016/S0304-4017(98)00121-6

Niezen, J.H., T.S. Waghorn, W.A.G. Charleston, and G.C. Waghorn. 1995. Growth and gastrointestinal nematode parasitism in lambs grazing either lucerne (*Medicago sativa*) or sulla (*Hedysarum coronarium*) which contains condensed tannins. J. Agric. Sci. 125:281–284. doi:10.1017/S0021859600084422

Niezen, J.H., G.C. Waghorn, T.B. Lyons, and D.C. Corson. 1998b. The potential benefits of ensiling the forage legume sulla compared with pasture. Proc. N. Z. Grassl. Assoc. 60:105–109.

Nyfeler, D., O. Huguenin-Elie, M. Suter, E. Frossard, J. Connolly, and A. Lüscher. 2009. Strong mixture effects among four species in fertilized agricultural grassland led to persistent and consistent transgressive overyielding. J. Appl. Ecol. 46:683–691. doi:10.1111/j.1365-2664.2009.01653.x

Nyfeler, D., O. Huguenin-Elie, M. Suter, E. Frossard, and A. Lüscher. 2011. Grass-legume mixtures can yield more nitrogen than legume pure stands due to mutual stimulation of nitrogen uptake from symbiotic and non-symbiotic sources. Agric. Ecosyst. Environ. 140:155–163. doi:10.1016/j.agee.2010.11.022

Pagán-Riestra, S., J.P. Muir, B.D. Lambert, L.O. Tedeschi, and L. Redmon. 2010. Phosphorus and other nutrient disappearance from plants containing condensed tannins using the nylon bag technique. Anim. Feed Sci. Technol. 156:19–25. doi:10.1016/j.anifeedsci.2009.12.006

Paolini, V., F. De-La-Farge, F. Prevot, P. Dorchies, and H. Hoste. 2005. Effects of the repeated distribution of sainfoin hay on the resistance and the resilience of goats naturally infected with gastrointestinal nematodes. Vet. Parasitol. 127:277–283. doi:10.1016/j.vetpar.2004.10.015

Papagiannopoulos, M., H.R. Wollseifen, A. Mellenthin, B. Haber, and R. Galensa. 2004. Identification and quantification of polyphenols in carob fruits (*Ceratonia siliqua* L.) and derived products by HPLC-UV-ESI/MSn. J. Agric. Food Chem. 52:3784–3791. doi:10.1021/jf030660y

Patra, A.K., and J. Saxena. 2010. A new perspective on the use of plant secondary metabolites to inhibit methanogenesis in the rumen. Phytochemistry 71:1198–1222. doi:10.1016/j.phytochem.2010.05.010

Pawelek, D.L., J.P. Muir, B.D. Lambert, and R.D. Wittie. 2008. *In sacco* rumen disappearance of condensed tannins, fiber, and nitrogen from herbaceous native Texas legumes in goats. Anim. Feed Sci. Technol. 142:1–16. doi:10.1016/j.anifeedsci.2007.02.014

Per, T.S., N.A. Khan, P.S. Reddy, A. Masood, M. Hasanuzzaman, M.I.R. Khan, and N.A. Anjum. 2017. Approaches in modulating proline metabolism in plants for salt and drought stress tolerance: Phytohormones, mineral nutrients and transgenics. Plant Physiol. Biochem. 115:126–140. doi:10.1016/j.plaphy.2017.03.018

Pérez-Díaz, R., M. Ryngajllo, J. Pérez-Díaz, H. Peña-Cortés, J.A. Casaretto, E. González-Villanueva, and S. Ruiz-Lara. 2014. VvMATE1 and VvMATE2 encode putative proanthocyanidin transporters expressed during berry development in *Vitis vinifera* L. Plant Cell Rep. 33:1147–1159. doi:10.1007/s00299-014-1604-9

Pérez-Jiménez, J., and J. Lluís Torres. 2011. Analysis of nonextractable phenolic compounds in foods: The current state of the art. J. Agric. Food Chem. 59:12713–12724. doi:10.1021/jf203372w

Petersen, J.C., N.S. Hill, and J.A. Mosjidis. 1991. Screening sericea lespedeza germplasm for herbage quality. Agron. J. 83:581–588. doi:10.2134/agronj1991.00021962008300030016x

Piluzza, G., L. Sulas, and S. Bullitta. 2014. Tannins in forage plants and their role in animal husbandry and environmental sustainability: A review. Grass Forage Sci. 69:32–48.

Porter, L.J. 1988. Flavans and proanthocyanidins. In: J.B. Harborne, editor, The flavonoids: Advances in research since 1980. Springer, Boston, MA. p. 21–62. doi:10.1007/978-1-4899-2913-6_2

Powell, J.M., M.J. Aguerre, and M.A. Wattiaux. 2011. Tannin extracts abate ammonia emissions from simulated dairy barn floors. J. Environ. Qual. 40:907–914. doi:10.2134/jeq2010.0492

Priolo, A., M. Bella, M. Lanza, V. Galofaro, L. Biondi, D. Barbagallo, et al. 2005. Carcass and meat quality of lambs fed fresh sulla (*Hedysarum coronarium* L.) with or without polyethylene glycol or concentrate. Small Rumin. Res. 59:281–288. doi:10.1016/j.smallrumres.2005.05.012

Purchas, R.W., and R.G. Keogh. 1984. Fatness of lambs grazed on 'Grasslands Maku' lotus and 'Grasslands Huia' white clover. Proc. N. Z. Soc. Anim. Prod. 44:219–221.

Quijada, J., C. Drake, E. Gaudin, R. El-Korso, H. Hoste, and I. Mueller-Harvey. 2018. Condensed tannin changes along the digestive tract in lambs fed with sainfoin pellets or hazelnut skins. J. Agric. Food Chem. (in press). doi:10.1021/acs.jafc.7b05538

Quijada, J., C. Fryganas, H.M. Ropiak, A. Ramsay, I. Mueller-Harvey, and H. Hoste. 2015. Anthelmintic activities against *Haemonchus contortus* or *Trichostrongylus colubriformis* from small ruminants are influenced by structural features of condensed tannins. J. Agric. Food Chem. 63:6346–6354. doi:10.1021/acs.jafc.5b00831

Ramírez-Restrepo, C.A., A. Pernthaner, T.N. Barry, N. López-Villalobos, R.J. Shaw, W.E. Pomroy, and W.R. Hein. 2010. Characterization of immune responses against gastrointestinal nematodes in weaned lambs grazing willow fodder blocks. Anim. Feed Sci. Technol. 155:99–110. doi:10.1016/j.anifeedsci.2009.10.006

Ramsay, A., C. Drake, A. Grosse Brinkhaus, M. Girard, F. Dohme-Meier, G. Bee, et al. 2015. NaOH enhances extractability and analysis of proanthocyanidins in ensiled sainfoin (*Onobrychis viciifolia*). J. Agric. Food Chem. 63:9471–9479. doi:10.1021/acs.jafc.5b04106

Ramsay, A., A.R. Williams, S.M. Thamsborg, and I. Mueller-Harvey. 2016. Galloylated proanthocyanidins from shea (*Vitellaria paradoxa*) meal have potent anthelmintic activity against *Ascaris suum*. Phytochemistry 122:146–153. doi:10.1016/j.phytochem.2015.12.005

Rautio, P., U.A. Bergvall, M. Karonen, and J.-P. Salminen. 2007. Bitter problems in ecological feeding experiments: Commercial tannin preparations and common methods for tannin quantifications. Biochem. Syst. Ecol. 35:257–262. doi:10.1016/j.bse.2006.10.016

Reid, C.S.W., M. Ulyatt, and J.M. Wilson. 1974. Plant tannins, bloat and nutritive value. Proc. N. Z. Soc. Anim. Prod. 34:82–92.

Ríos-De Álvarez, L., A.W. Greer, F. Jackson, S. Athanasiadou, I. Kyriazakis, and J.F. Huntley. 2008. The effect of dietary sainfoin (*Onobrychis viciifolia*) on local cellular responses to *Trichostrongylus colubriformis* in sheep. Parasitology 135:1117–1124. doi:10.1017/S0031182008004563

Rochfort, S., A.J. Parker, and F.R. Dunshea. 2008. Plant bioactives for ruminant health and productivity. Phytochemistry 69:299–322. doi:10.1016/j.phytochem.2007.08.017

Rodríguez-Pérez, C., R. Quirantes-Pine, N. Amessis-Ouchemoukh, K. Madani, A. Seguar-Carretero, and A. Fernandex-Gutierrez. 2013. A metabolite-profiling approach allows the identification of new compounds from *Pistacia lentiscus* leaves. J. Pharm. Biomed. Anal. 77:167–174. doi:10.1016/j.jpba.2013.01.026

Ropiak, H.M., O. Desrues, A.R. Williams, A. Ramsay, I. Mueller-Harvey, and S.M. Thamsborg. 2016a. Structure-activity relationship of condensed tannins and synergism with *trans*-cinnamaldehyde against *Caenorhabditis elegans*. J. Agric. Food Chem. 64:8795–8805. doi:10.1021/acs.jafc.6b03842

Ropiak, H.M., P. Lachmann, A. Ramsay, R.J. Green, and I. Mueller-Harvey. 2017. Identification of structural features of condensed tannins that affect protein aggregation. PLoS One 12:e0170768. doi:10.1371/journal.pone.0170768

Ropiak, H.M., A. Ramsay, and I. Mueller-Harvey. 2016b. Condensed tannins in extracts from European medicinal plants and herbal products. J. Pharm. Biomed. Anal. 121:225–231. doi:10.1016/j.jpba.2015.12.034

Salminen, J.-P., and M. Karonen. 2011. Chemical ecology of tannins and other phenolics: We need a change in approach. Funct. Ecol. 25:325–338. doi:10.1111/j.1365-2435.2010.01826.x

Saratsis, A., N. Voutzourakis, T. Theodosiou, A. Stefanakis, and S. Sotiraki. 2016. The effect of sainfoin (*Onobrychis viciifolia*) and carob pods (*Ceratonia siliqua*) feeding regimes on the control of lamb coccidiosis. Parasitol. Res. 115:2233–2242. doi:10.1007/s00436-016-4966-9

Scharenberg, A., Y. Arrigo, A. Gutzwiller, U. Wyss, D.H. Hess, M. Kreuzer, and F. Dohme. 2007. Effect of feeding dehydrated and ensiled tanniferous sainfoin (*Onobrychis viciifolia*) on nitrogen and mineral digestion and metabolism of lambs. Arch. Anim. Nutr. 61:390–405. doi:10.1080/17450390701565081

Schofield, P., D.M. Mbugua, and A.N. Pell. 2001. Analysis of condensed tannins: A review. Anim. Feed Sci. Technol. 91:21–40. doi:10.1016/S0377-8401(01)00228-0

Schreurs, N.M., D.M. Marotti, M.H. Tavendale, G.A. Lane, T.N. Barry, N. López-Villalobos, and W.C. McNabb. 2007. Concentration of indoles and other rumen metabolites in sheep after a meal of fresh white clover, perennial ryegrass or *Lotus corniculatus* and the appearance of indoles in the blood. J. Sci. Food Agric. 87:1042–1051. doi:10.1002/jsfa.2804

Scioneaux, A.N., M.A. Schmidt, M.A. Moore, R.L. Lindroth, S.C. Wooley, and A.E. Hagerman. 2011. Qualitative variation in proanthocyanidin composition of *Populus* species and hybrids: Genetics is the key. J. Chem. Ecol. 37:57–70. doi:10.1007/s10886-010-9887-y

Self, R., J. Eagles, G.C. Galletti, I. Mueller-Harvey, R.D. Hartley, A.G.H. Lea, et al. 1986. Fast atom bombardment mass spectrometry of polyphenols (syn. vegetable tannins). Biomed. Environ. Mass Spectrom. 13:449–468. doi:10.1002/bms.1200130902

Sheldrick, R.D., and D.J. Thomson. 1982. Management and utilization of sainfoin (*Onobrychis sativa*). Inf. Leaflet no. 13. Grassl. Res. Inst., Hurley, Berkshire, UK.

Sivakumaran, S., A.L. Molan, L.P. Meagher, B. Kolb, L.Y. Foo, G.A. Lane, et al. 2004. Variation in antimicrobial action of proanthocyanidins from *Dorycnium rectum* against rumen bacteria. Phytochemistry 65:2485–2497. doi:10.1016/j.phytochem.2004.08.046

Sivakumaran, S., W. Rumball, G.A. Lane, K. Fraser, L.Y. Foo, M. Yu, and L.P. Meagher. 2006. Variation of proanthocyanidins in *Lotus* species. J. Chem. Ecol. 32:1797–1816. doi:10.1007/s10886-006-9110-3

Spencer, P., S. Sivakumaran, K. Fraser, L.Y. Foo, G.A. Lane, P.J.B. Edwards, and L.P. Meagher. 2007. Isolation and characterisation of procyanidins from *Rumex obtusifolius*. Phytochem. Anal. 18:193–203. doi:10.1002/pca.967

Springer, T.L., R.L. McGraw, and G.E. Aiken. 2002. Variation of condensed tannins in roundhead lespedeza germplasm. Crop Sci. 42:2157–2160. doi:10.2135/cropsci2002.2157

Stienezen, M., G.C. Waghorn, and G.B. Douglas. 1996. Digestibility and effects of condensed tannins on digestion of sulla (*Hedysarum coronarium*) when fed to sheep. N. Z. J. Agric. Res. 39:215–221. doi:10.1080/00288233.1996.9513180

Stringano, E. 2011. Analysis of sainfoin (*Onobrychis viciifolia*) proanthocyanidins by complementary and newly developed techniques. Ph.D. diss., Univ. of Reading, Reading, UK.

Stringano, E., R. Cramer, W. Hayes, C. Smith, T. Gibson, and I. Mueller-Harvey. 2011. Deciphering the complexity of sainfoin (*Onobrychis viciifolia*) proanthocyanidins by MALDI-TOF mass spectrometry with a judicious choice of isotope patterns and matrices. Anal. Chem. 83:4147–4153. doi:10.1021/ac2003856

Stringano, E., C. Hayot Carbonero, L.M.J. Smith, R.H. Brown, and I. Mueller-Harvey. 2012. Proanthocyanidin diversity in the EU 'HealthyHay' sainfoin (*Onobrychis viciifolia*) germplasm collection. Phytochemistry 77:197–208. doi:10.1016/j.phytochem.2012.01.013

Suter, M., D. Hofer, and A. Lüscher. 2017. Weed suppression enhanced by increasing functional trait dispersion and resource capture in forage ley mixtures. Agric. Ecosyst. Environ. 240:329–339. doi:10.1016/j.agee.2017.01.007

Szabados, L., and A. Savouré. 2010. Proline: A multifunctional amino acid. Trends Plant Sci. 15:89–97. doi:10.1016/j.tplants.2009.11.009

Szczyglowski, K., and J. Stougaard. 2008. *Lotus* genome: Pod of gold for legume research. Trends Plant Sci. 13:515–517. doi:10.1016/j.tplants.2008.08.001

Takeuchi, H., V.T. Trang, N. Morimoto, Y. Nishida, Y. Matsumura, and T. Sugiura. 2014. Natural products and food components with anti-*Helicobacter pylori* activities. World J. Gastroenterol. 20:8971–8978. doi:10.3748/wjg.v20.i27.8971

Tedeschi, L.O., C.A. Ramírez-Restrepo, and J.P. Muir. 2014. Developing a conceptual model of possible benefits of condensed tannins for ruminant production. Animal 8:1095–1105. doi:10.1017/S1751731114000974

Terrill, T.H., J.E. Miller, J.M. Burke, J.A. Mosjidis, and R.M. Kaplan. 2012. Experiences with integrated concepts for the control of *Haemonchus contortus* in sheep and goats in the United States. Vet. Parasitol. 186:28–37. doi:10.1016/j.vetpar.2011.11.043

Terrill, T.H., W.R. Windham, C.S. Hoveland, H.E. Amos, B. Kouakou, and S. Gelaye. 1997. Nitrogen and fiber digestion in sheep fed fresh-frozen and field-dried high- and low-tannin sericea lespedeza. In: J.G. Buchanan-Smith, L.D. Bailey, and P. McCaughey, editors, Proceedings of the 18th International Grasslands Congress, Winnipeg, MB, and Saskatoon, SK. 9–18 June 1997. Int. Grassl. Congr., Lexington, KY. p. 8-11–8-12. http://www.internationalgrasslands.org/files/igc/publications/1997/1-08-011.pdf (accessed 30 Sept. 2017).

Terrill, T.H., E. Griffin, D.S. Kommuru, J.E. Miller, J.A. Mosjidis, M.T. Kearney, and J.M. Burke. 2016. Effect of ensiling on anti-parasitic properties of sericea lespedeza. Am. Forage Grassl. Counc., Berea, KY. http://www.afgc.org/proceedings/2016/TerrillAFGCpaper.pdf (accessed 30 Sept. 2017).

Theodoridou, K., J. Aufrère, D. Andueza, A. Le Morvan, F. Picard, J. Pourrat, and R. Baumont. 2012. Effects of condensed tannins in wrapped silage bales of sainfoin (*Onobrychis viciifolia*) on in vivo and in situ digestion in sheep. Animal 6:245–253. doi:10.1017/S1751731111001510

Theodoridou, K., J. Aufrère, D. Andueza, A. Le Morvan, F. Picard, E. Stringano, et al. 2011. Effect of plant development during first and second growth cycle on chemical composition, condensed tannins and nutritive value of three sainfoin (*Onobrychis viciifolia*) varieties and lucerne. Grass Forage Sci. 66:402–414. doi:10.1111/j.1365-2494.2011.00798.x

Theodoridou, K., J. Aufrère, D. Andueza, J. Pourrat, A. Le Morvan, E. Stringano, et al. 2010. Effects of condensed tannins in fresh sainfoin (*Onobrychis viciifolia*) on *in vivo* and *in situ* digestion in sheep. Anim. Feed Sci. Technol. 160:23–38. doi:10.1016/j.anifeedsci.2010.06.007

Thomson, D.J., D.E. Beever, D.G. Harrison, I.W. Hill, and D.F. Osbourn. 1971. The digestion of dried lucerne (*Medicago sativa*, L) and dried sainfoin (*Onobrychis viciifolia*, Scop) by sheep. Proc. Nutr. Soc. 30:14A.

Tibe, O., L.P. Meagher, K. Fraser, and D.R.K. Harding. 2011. Condensed tannins and flavonoids from the forage legume sulla (*Hedysarum coronarium*). J. Agric. Food Chem. 59:9402–9409. doi:10.1021/jf2014759

Tibe, O., A. Pernthaner, I. Sutherland, L. Lesperance, and D.R.K. Harding. 2012. Condensed tannins from Botswanan forage plants are effective priming agents of γδ T cells in ruminants. Vet. Immunol. Immunopathol. 146:237–244. doi:10.1016/j.vetimm.2012.03.003

Tyrrell, H.F., and P.W. Moe. 1975. Effect of intake on digestive efficiency. J. Dairy Sci. 58:1151–1163. doi:10.3168/jds.S0022-0302(75)84694-7

Vasileva, V., and A. Ilieva. 2016. Changes in some parameters in mixtures of sainfoin with subterranean clover. Int. J. Waste Resour. 6:226. doi:10.4172/2252-5211.1000226

Verdier, J., J. Zhao, I. Torres-Jerez, S. Ge, C. Liu, X. He, et al. 2012. MtPAR MYB transcription factor acts as an on switch for proanthocyanidin biosynthesis in *Medicago truncatula*. Proc. Natl. Acad. Sci. USA 109:1766–1771. doi:10.1073/pnas.1120916109

Vernhet, A., S. Dubascoux, B. Cabane, H. Fulcrand, E. Dubreucq, and C. Poncet-Legrand. 2011. Characterization of oxidized tannins: Comparison of depolymerization methods, asymmetric flow field-flow fractionation and small-angle X-ray scattering. Anal. Bioanal. Chem. 401:1559–1569. doi:10.1007/s00216-011-5076-2

Villalba, J.J., R. Cabassu, and S.A. Gunter. 2015. Forage choice in pasturelands: Influence on cattle foraging behavior and performance. J. Anim. Sci. 93:1729–1740. doi:10.2527/jas.2014-8667

Waghorn, G. 2008. Beneficial and detrimental effects of dietary condensed tannins for sustainable sheep and goat production: Progress and challenges. Anim. Feed Sci. Technol. 147:116–139. doi:10.1016/j.anifeedsci.2007.09.013

Waghorn, G.C., A. John, W.T. Jones, and I.D. Shelton. 1987. Nutritive value of *Lotus corniculatus* L. containing low and medium concentrations of condensed tannins for sheep. Proc. N. Z. Soc. Anim. Prod. 47:25–30.

Waghorn, G.C., and W.J. Jones. 1989. Bloat in cattle 46. The potential of dock (*Rumex obtusifolius*) as an antibloat agent for cattle. N. Z. J. Agric. Res. 32:227–235. doi:10.1080/00288233.1989.10423458

Waghorn, G.C., W.T. Jones, I.D. Shelton, and W.C. McNabb. 1990. Condensed tannins and the nutritive value of herbage. Proc. N. Z. Grassl. Assoc. 51:171–179.

Waghorn, G.C., and A.L. Molan. 2001. Effect of condensed tannins in Dorycnium rectum on its nutritive value and on the development of sheep parasite larvae. Proc. N. Z. Grassl. Assoc. 63:273–278.

Waghorn, G.C., and I.D. Shelton. 1997. Effect of condensed tannins in *Lotus corniculatus* on the nutritive value of pasture for sheep. J. Agric. Sci. 128:365–372. doi:10.1017/S0021859697004218

Waghorn, G.C., J.D. Reed, and L.R. Ndlovu. 1997. Condensed tannins and herbivore nutrition. In: J.G. Buchanan-Smith, L.D. Bailey, and P. McCaughey, editors, Proceedings of the 18th International Grasslands Congress, Winnipeg, MB, and Saskatoon, SK. 9–18 June 1997. Int. Grassl. Congr., Lexington, KY. p. 153–166. http://www.internationalgrasslands.org/files/igc/publications/1997/iii-153.pdf (accessed 30 Sept. 2017).

Wang, Y., L.R. Barbieri, B.P. Berg, and T.A. McAllister. 2007. Effects of mixing sainfoin with alfalfa on ensiling, ruminal fermentation and total tract digestion of silage. Anim. Feed Sci. Technol. 135:296–314. doi:10.1016/j.anifeedsci.2006.07.002

Wang, Y., B.P. Berg, L.R. Barbieri, D.M. Veira, and T.A. McAllister. 2006. Comparison of alfalfa and mixed alfalfa-sainfoin pastures for grazing cattle: Effects on incidence of bloat, ruminal fermentation, and feed intake. Can. J. Anim. Sci. 86:383–392. doi:10.4141/A06-009

Wang, Y., W. Majak, and T.A. McAllister. 2012. Forthy bloat in ruminants: Cause, occurrence, and mitigation strategies. Anim. Feed Sci. Technol. 172:103–114. doi:10.1016/j.anifeedsci.2011.12.012

Wang, Y., T.A. McAllister, and S. Acharya. 2015. Condensed tannins in sainfoin: Composition, concentration, and effects on nutritive and feeding value of sainfoin forage. Crop Sci. 55:13–22. doi:10.2135/cropsci2014.07.0489

Wang, Y., A.P. Singh, W.J. Hurst, J.A. Glinski, H. Koo, and N. Vorsa. 2016. Influence of degree-of-polymerization and linkage on the quantification of proanthocyanidins using 4-dimethylaminocinnamaldehyde (DMAC) assay. J. Agric. Food Chem. 64:2190–2199. doi:10.1021/acs.jafc.5b05408

Wang, Y., G.C. Waghorn, W.C. McNabb, T.N. Barry, M.J. Hedley, and I.D. Shelton. 1996. Effect of condensed tannins in *Lotus corniculatus* upon the digestion of methionine and cysteine in the small intestine of sheep. J. Agric. Sci. 127:413–421. doi:10.1017/S0021859600078576

Williams, A.R., C. Fryganas, A. Ramsay, I. Mueller-Harvey, and S.M. Thamsborg. 2014a. Direct anthelmintic effects of condensed tannins from diverse plant sources against *Ascaris suum*. PLoS One 9:e97053. doi:10.1371/journal.pone.0097053 [erratum: 9:e99738.]

Williams, A.R., C. Fryganas, K. Reichwald, S. Skov, I. Mueller-Harvey, and S.M. Thamsborg. 2016. Polymerization-dependent activation of porcine γδ T-cells by proanthocyanidins. Res. Vet. Sci. 105:209–215. doi:10.1016/j.rvsc.2016.02.021

Williams, A.R., E.J. Klaver, L.C. Laan, A. Ramsay, C. Fryganas, R. Difborg, et al. 2017. Co-operative suppression of inflammatory responses in human dendritic cells by proanthocyanidins and products from the parasitic nematode *Trichuris suis*. Immunology 150:312–328. doi:10.1111/imm.12687

Williams, A.R., A. Ramsay, T.V.A. Hansen, H.M. Ropiak, H. Mejer, P. Nejsum, et al. 2015. Anthelmintic activity of *trans*-cinnamaldehyde and A- and B-type proanthocyanidins derived from cinnamon (*Cinnamomum verum*). Nat. Sci. Rep. 5:14791. doi:10.1038/srep14791

Williams, A.R., H.M. Ropiak, C. Fryganas, O. Desrues, I. Mueller-Harvey, and S.M. Thamsborg. 2014b. Assessment of the anthelmintic activity of medicinal plant extracts and purified condensed tannins against free-living and parasitic stages of *Oesophagostomum dentatum*. Parasit. Vectors 7:518. doi:10.1186/s13071-014-0518-2

Zarrabian, M., M.M. Majidi, and M.H. Ehtemam. 2013. Genetic diversity in a worldwide collection of sainfoin using morphological, anatomical, and molecular markers. Crop Sci. 53:2483–2495. doi:10.2135/cropsci2013.03.0130

Zeller, W.E. 2019. Activity, purification, and analysis of condensed tannins: Current state of affairs and future endeavors. Crop Sci. (in review).

Zeller, W.E., and J.H. Grabber. 2015. Redesigning forages with condensed tannins. Progressive Forage Grower 16:7–8.

Zeller, W.E., A. Ramsay, H.M. Ropiak, C. Fryganas, I. Mueller-Harvey, R.H. Brown, et al. 2015a. ^{1}H-^{13}C HSQC NMR spectroscopy for estimating procyanidin/prodelphinidin and *cis/trans* flavanol ratios of condensed tannin fractions: Correlation with thiolysis. J. Agric. Food Chem. 63:1967–1973. doi:10.1021/jf504743b

Zeller, W.E., M.L. Sullivan, I. Mueller-Harvey, J.H. Grabber, A. Ramsay, C. Drake, and R.H. Brown. 2015b. Protein precipitation behavior of condensed tannins from *Lotus pedunculatus* and *Trifolium repens* with different mean degrees of polymerization. J. Agric. Food Chem. 63:1160–1168. doi:10.1021/jf504715p

Zhou, M., L. Wei, Z. Sun, L. Gao, Y. Meng, Y. Tang, and Y. Wu. 2015. Production and transcriptional regulation of proanthocyanidin biosynthesis in forage legumes. Appl. Microbiol. Biotechnol. 99:3797–3806. doi:10.1007/s00253-015-6533-1

Zhu, Q., S. Sui, X. Lei, Z. Yang, K. Lu, G. Liu, et al. 2015. Ectopic expression of the Coleus R2R3 MYB-type proanthocyanidin regulator gene SsMYB3 alters the flower color in transgenic tobacco. PLoS One 10:e0139392. doi:10.1371/journal.pone.0139392