

# *RXR ligands negatively regulate thrombosis and hemostasis*

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# RXR Ligands Negatively Regulate Thrombosis and Hemostasis

Amanda J. Unsworth, Gagan D. Flora, Parvathy Sasikumar, Alexander P. Bye, Tanya Sage, Neline Kriek, Marilena Crescente, Jonathan M. Gibbins

**Objective**—Platelets have been found to express intracellular nuclear receptors including the retinoid X receptors (RXR $\alpha$  and RXR $\beta$ ). Treatment of platelets with ligands of RXR has been shown to inhibit platelet responses to ADP and thromboxane A2; however, the effects on responses to other platelet agonists and the underlying mechanism have not been fully characterized.

**Approach and Results**—The effect of 9-cis-retinoic acid, docosahexaenoic acid and methoprene acid on collagen receptor (glycoprotein VI [GPVI]) agonists and thrombin-stimulated platelet function; including aggregation, granule secretion, integrin activation, calcium mobilization, integrin  $\alpha$ IIb $\beta$ 3 outside-in signaling and thrombus formation in vitro and in vivo were determined. Treatment of platelets with RXR ligands resulted in attenuation of platelet functional responses after stimulation by GPVI agonists or thrombin and inhibition of integrin  $\alpha$ IIb $\beta$ 3 outside-in signaling. Treatment with 9-cis-retinoic acid caused inhibition of thrombus formation in vitro and an impairment of thrombosis and hemostasis in vivo. Both RXR ligands stimulated protein kinase A activation, measured by VASP S157 phosphorylation, that was found to be dependent on both cAMP and nuclear factor  $\kappa$ -light-chain-enhancer of activated B cell activity.

**Conclusions**—This study identifies a widespread, negative regulatory role for RXR in the regulation of platelet functional responses and thrombus formation and describes novel events that lead to the upregulation of protein kinase A, a known negative regulator of many aspects of platelet function. This mechanism may offer a possible explanation for the cardioprotective effects described in vivo after treatment with RXR ligands.

**Visual Overview**—An online [visual overview](#) is available for this article. (*Arterioscler Thromb Vasc Biol*. 2017;37:812-822. DOI: 10.1161/ATVBAHA.117.309207.)

**Key Words:** atherosclerosis ■ eczema ■ platelet activation ■ thrombin

Many intracellular nuclear receptors are expressed in human platelets that negatively regulate platelet function when stimulated by their endogenous ligands.<sup>1-12</sup> Retinoid X receptors (RXR) ( $\alpha$ ,  $\beta$ , and  $\gamma$ ) regulate transcription and expression of specific genes involved in cell proliferation, differentiation, and lipid metabolism and are activated by retinoids and vitamin A derivatives.<sup>13-17</sup> RXR ligands have been found to exert cardioprotective effects by reducing atherosclerosis in apolipoprotein E knockout mice,<sup>18</sup> and treatment with RXR agonists has also been shown to sensitize cells to insulin and rescue hyperglycemia and hyperinsulinemia in type 2 diabetes mellitus mouse models, highlighting their potential role as therapeutics for the treatment of type 2 diabetes mellitus.<sup>19</sup>

Several recent studies have identified acute, nongenomic roles for several members of the nuclear hormone superfamily including RXR.<sup>1,3-5,11,20,21</sup> Treatment with nuclear receptor ligands is associated with cardioprotective effects, whereas ligands for RXR are associated with reduced atherosclerosis

and inflammation,<sup>18</sup> which may be because of regulation of platelet activity. 9-cis-retinoic acid (9-cis-RA), which is marketed as Alitretinoin, is used for the treatment of Kaposi sarcoma and chronic hand eczema, and decreased blood clotting is listed as one of its side effects.<sup>22,23</sup>

Treatment of platelets with RXR ligands has been shown to inhibit ADP and thromboxane A2 receptor-induced platelet activation via inhibition of Gq.<sup>5</sup> However, effects of RXR ligands on platelet responses to other receptor agonists (such as thrombin and low concentrations of collagen and other glycoprotein VI [GPVI] receptor agonists) and integrin  $\alpha$ IIb $\beta$ 3 function are unknown, and the antithrombotic effects of RXR agonists have not be studied.

This study set out to investigate the mechanism by which RXR ligands regulate platelet function and thrombus formation and explore the potential antiplatelet properties of RXR ligands. We describe that RXR agonists evoke broad inhibition of platelet function, resulting in inhibition of thrombus

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**Nonstandard Abbreviations and Acronyms**

<b>9-cis-RA</b>	9-cis-retinoic acid
<b>IP</b>	prostaglandin I <sub>2</sub>
<b>NF<math>\kappa</math>B</b>	nuclear factor $\kappa$ -light-chain-enhancer of activated B cells
<b>PKA</b>	protein kinase A
<b>PPAR</b>	peroxisome proliferator-activated receptor
<b>RXR</b>	retinoid X receptor
<b>VASP</b>	vasodilator-stimulated phosphoprotein

formation in vitro and in vivo. We report that RXR ligands increase protein kinase A (PKA) activity via both cAMP and nuclear factor  $\kappa$ -light-chain-enhancer of activated B cells (NF $\kappa$ B)-dependent mechanisms that have not been described for the endogenous agonist of any other nuclear receptor.<sup>24</sup>

## Materials and Methods

Materials and Methods are available in the [online-only Data Supplement](#).

## Results

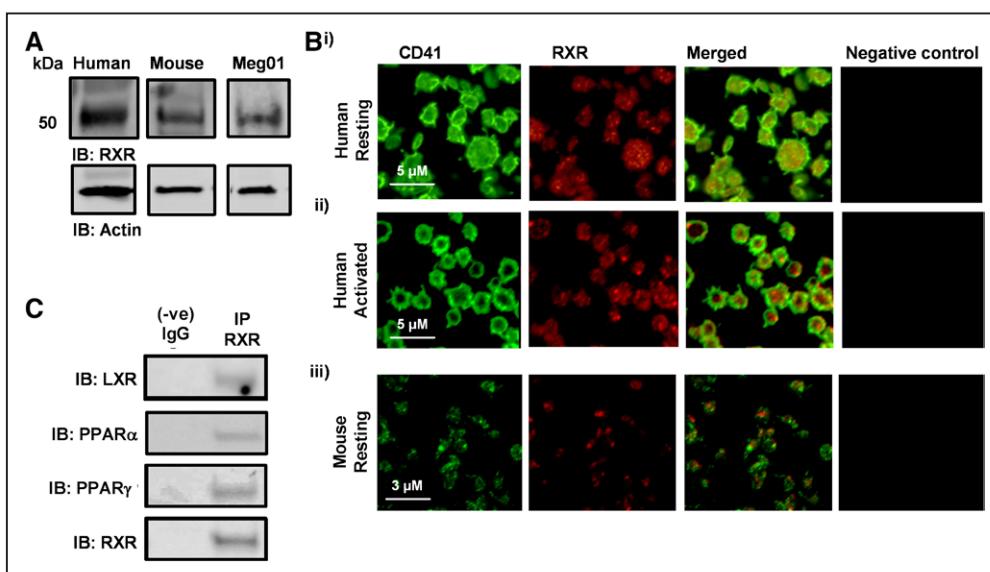
### Expression and Localization of RXR in Platelets

Here, we confirm protein expression of RXR in both human and mouse platelets and Meg01 cells using immunoblot analysis (Figure 1A) and also immunohistochemistry (Figure 1B and 1C). This is further supported by previously described expression of the separate RXR isoforms in both the human and mouse transcriptome and protein expression in human platelets.<sup>5,25</sup> Staining of platelets with antibodies raised against both RXR isoforms

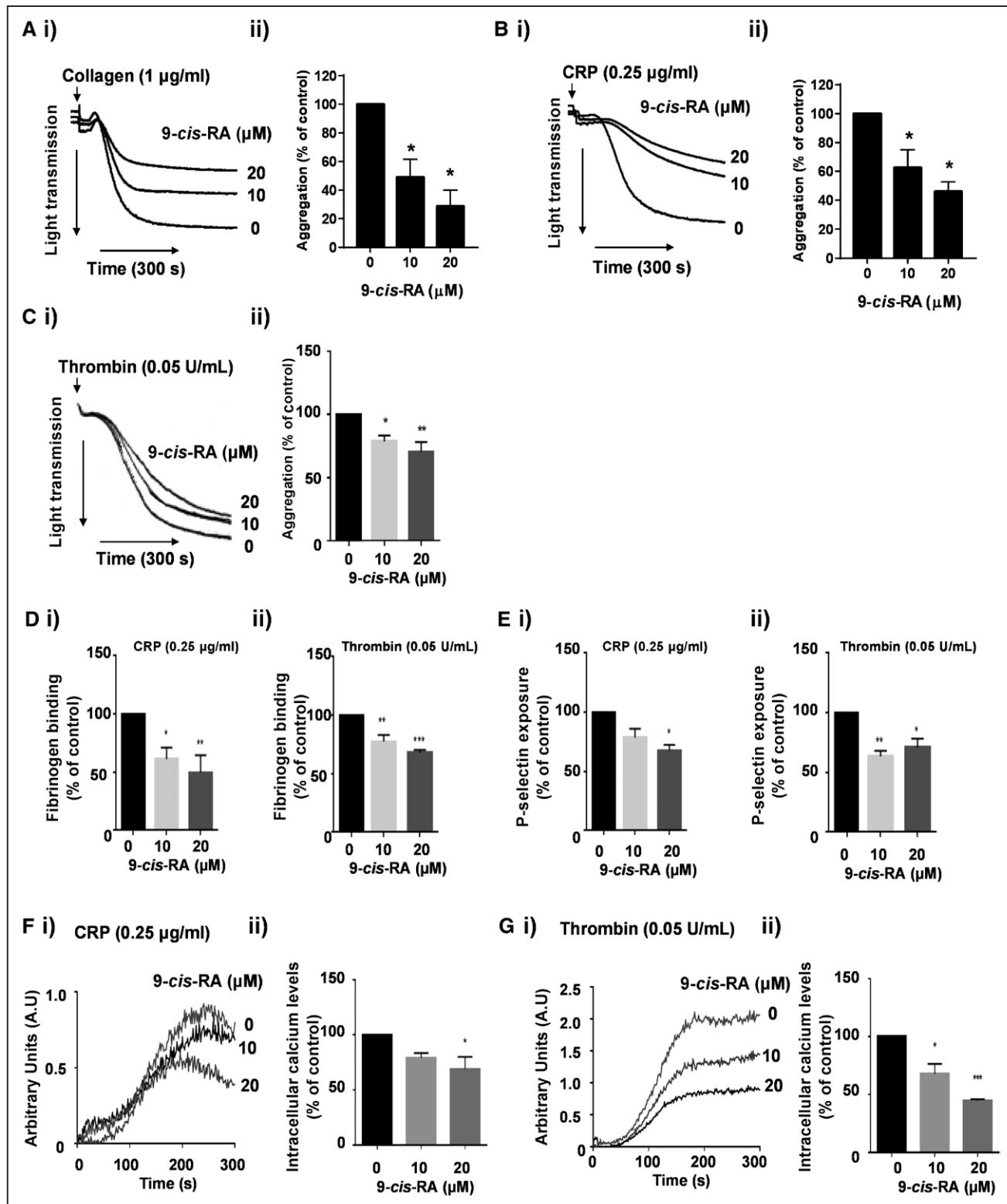
and the membrane marker CD41 confirmed the presence of RXR in both human and mouse platelets with punctate staining throughout the cytosol within resting human platelets, which was not altered on platelet activation. Coimmunoprecipitation experiments were performed to determine whether RXR could form heterodimers with other nuclear receptors in platelets (as in other cell types). RXR was found to coimmunoprecipitate with the peroxisome proliferator-activated receptors (PPARs), PPAR $\alpha$ , PPAR $\beta$ , and PPAR $\gamma$ , and LXR (liver X receptor; Figure 1C), thereby suggesting that RXR can form heterodimers with other nuclear receptors in human platelets.

### RXR Agonists Inhibit Platelet Aggregation to a Range of Agonists

Light transmission aggregometry using human washed platelets was used to analyze the effects of the endogenous RXR ligands 9-cis-RA, docosahexaenoic acid, and synthetic ligand methoprene acid on platelet aggregation. We found that 9-cis-RA, docosahexaenoic acid, and methoprene acid at the concentrations used (10 and 20  $\mu$ mol/L) did not cause platelet aggregation in the absence of platelet agonists (Figure 1A in the [online-only Data Supplement](#)). As shown in Figure 2, pretreatment with 9-cis-RA (20  $\mu$ mol/L) inhibited platelet aggregation to collagen (1  $\mu$ g/mL; Figure 2A), the GPVI collagen receptor-specific agonist, CRP-XL (0.25  $\mu$ g/mL; Figure 2B), and thrombin (0.05 U/mL; Figure 2C) compared with vehicle controls (containing 0.1% dimethyl sulfoxide), with  $\approx$ 60% and 20% inhibition after stimulation by collagen (and CRP-XL) and thrombin, respectively. Treatment with docosahexaenoic acid or methoprene acid (10 or 20  $\mu$ mol/L) also inhibited



**Figure 1.** Expression and localization of retinoid X receptors (RXR) in platelets. Human and mouse washed platelets were (A) lysed in (SDS-PAGE) Laemmli sample buffer, separated by SDS-PAGE and transferred to polyvinylidene fluoride (PVDF) membranes before blotting with an antibody that recognizes both  $\alpha$  and  $\beta$  isoforms of RXR. Representative blots are shown. B, Human platelets (i) resting and (ii) activated and (iii) mouse platelets (resting) were fixed in 4% paraformaldehyde and permeabilized with 0.2% Triton-X-100 and stained for RXR (in red) and CD41 (in green, as a marker for the platelet membrane) with primary antibodies raised against RXR and CD41. Secondary antibodies conjugated to Alexa-647 and Alexa-488 were used to visualize RXR and CD41, respectively. No primary antibody-treated samples were also included as a negative control. Representative images shown. C, Human washed platelets lysed in NP40 buffer before immunoprecipitation of RXR using 1  $\mu$ g/mL of primary antibody overnight at 4°C and protein A/G magnetic beads. Pull down samples lysed in (SDS-PAGE) Laemmli sample buffer, separated by SDS-PAGE, and transferred to PVDF membranes before blotting with antibodies that recognize different nuclear receptors, including PPAR $\alpha$ , PPAR $\gamma$ , and LXR (liver X receptor) and a secondary antibody that does not recognize denatured IgG. RXR IgG loaded as a control for IgG contamination. Representative blot shown.



**Figure 2.** The effect of retinoid X receptors (RXR) ligands on platelet function. Washed human platelets were pretreated for 10 min with increasing concentrations of 9-cis-RA (10 and 20  $\mu$ mol/L) before stimulation with (A) collagen (1  $\mu$ g/mL), (B) CRP-XL (0.25  $\mu$ g/mL), or (C) thrombin (0.05 U/mL) and aggregation monitored using optical light transmission aggregometry, (i) representative traces and (ii) quantified data shown. D, Integrin activation measured as fibrinogen binding in (i) CRP-XL-stimulated and (ii) thrombin-stimulated platelets. E,  $\alpha$ -Granule secretion measured as P-selectin exposure in (i) CRP-XL-stimulated and (ii) thrombin-stimulated platelets. Intracellular calcium levels determined in FURA-2 AM-loaded platelets after stimulation with (F) CRP-XL (0.25  $\mu$ g/mL) and (G) thrombin (0.05 U/mL); (i) representative traces and (ii) quantified data shown. Data expressed as the percentage of untreated control. Results are mean  $\pm$  SEM for  $n \geq 3$ . \* $P \leq 0.05$  in comparison to vehicle controls.

aggregation to collagen and thrombin (Figure 1B through 1E in the [online-only Data Supplement](#)). As collagen-induced platelet aggregation is partially dependent on the release of secondary mediators and RXR agonists have been shown to inhibit platelet responses to ADP and thromboxane A2,<sup>5</sup> we determined whether the inhibitory effects of the RXR agonists were because of their ability to inhibit secondary mediator signaling. The effects of 9-cis-RA on collagen-evoked aggregation were found to be additive to the inhibition caused by blockade of secondary mediator signaling and suggested that 9-cis-RA was able to inhibit collagen-evoked signaling directly (Figure 1F in the [online-only Data Supplement](#)).

Currently available antagonists for RXR, including HX531, are classified by their ability to inhibit the genomic functions of the receptor, and it is unknown how they affect the nongenomic actions of RXR. End point aggregation assays were performed in 96-well microtiter plates in the presence or absence of HX531 (10 and 30  $\mu$ mol/L) to a range of platelet agonist concentrations (Figure 1G in the [online-only Data Supplement](#)). Interestingly, HX531 caused inhibition of platelet aggregation after stimulation by collagen, thrombin, and the thromboxane A2 receptor agonist U46619, which is similar to the inhibition observed previously by 9-cis-RA, docosahexaenoic acid, and methoprene acid. HX531 binds to the same binding pocket in the RXR receptor as the 2 RXR agonists 9-cis-RA and methoprene acid. Although HX531 functions to block the DNA-binding ability of the RXR receptor, it is possible that alteration of the DNA-binding region is not involved in the nongenomic functions of this receptor, whereas other conformational changes that occur after ligand interaction with the binding pocket may be involved. These findings suggest that HX531 acts as an agonist and not an antagonist of the nongenomic functions of RXR.

### RXR Agonists Reduce Integrin Activation and $\alpha$ -Granule Secretion

Platelet inside-out signaling is essential for activation of integrin  $\alpha_{IIb}\beta_3$  which supports aggregation by enabling integrin binding to fibrinogen and von Willebrand factor. Secretion of platelet granule contents amplifies platelet activation through recruitment of surrounding platelets, enhancing thrombus formation. To determine whether RXR ligands alter the activation of integrin  $\alpha_{IIb}\beta_3$  and  $\alpha$ -granule secretion, fibrinogen binding and P-selectin exposure (a marker of  $\alpha$ -granule secretion) on platelets pretreated with RXR ligands were determined in CRP-XL (0.25  $\mu$ g/mL) and thrombin (0.05 U/mL)-stimulated platelets (Figure 2D and 2E). Consistent with the inhibition of platelet aggregation, fibrinogen binding and P-selectin exposure to both agonists was inhibited in 9-cis-RA-treated platelets compared with controls. Maximum inhibition of both fibrinogen binding (50% CRP-XL and 25% thrombin) and P-selectin exposure (20%–25% to either agonist) observed at the highest concentration of 9-cis-RA (20  $\mu$ mol/L; Figure 2D and 2E). Similar results were obtained after treatment with methoprene acid (Figure 2A and 2B in the [online-only Data Supplement](#)).

### 9-cis-RA Inhibits Elevation of Intracellular Calcium

Elevation of cytosolic  $\text{Ca}^{2+}$  levels is a critical event in platelet activation and is stimulated by both collagen and thrombin,

although via distinct mechanisms.<sup>26</sup> As shown in Figure 2F and 2G, treatment with 9-cis-RA resulted in a reduction in peak cytosolic calcium levels of 25% after stimulation with CRP-XL (20  $\mu$ mol/L 9-cis-RA) and  $\approx$ 50% after thrombin stimulation in comparison to vehicle control. Treatment with methoprene acid had similar effects (Figure 2H and 2I in the [online-only Data Supplement](#)). These observations indicate that RXR agonists are able to inhibit  $\text{Ca}^{2+}$  elevation evoked by both of these platelet agonists, despite the distinct signaling pathways that are activated by them.

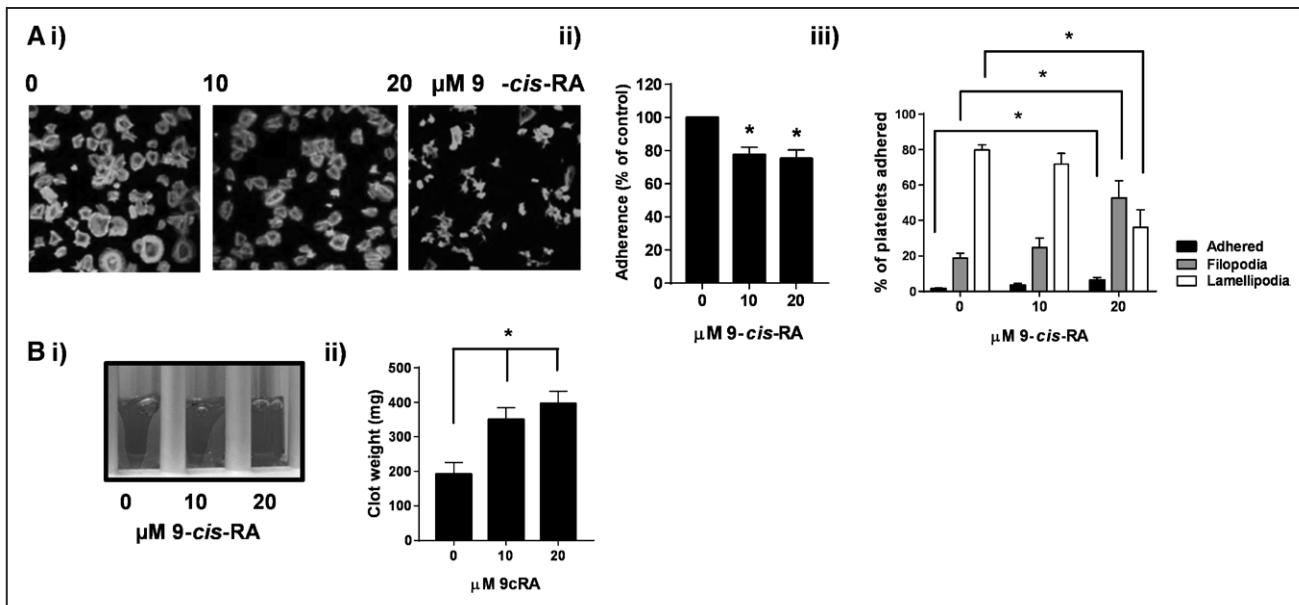
### RXR Ligands Negatively Regulate Outside-In Signaling

Given that RXR agonists have broader inhibitory effects than initially appreciated, the effect of the RXR agonists on adhesion to fibrinogen and outside-in signaling evoked by integrin  $\alpha_{IIb}\beta_3$  was also investigated. After treatment with 9-cis-RA (10 and 20  $\mu$ mol/L), both adhesion and spreading on fibrinogen-(100  $\mu$ g/mL)-coated coverslips were found to be inhibited in 9-cis-RA-treated samples compared with vehicle-treated controls (Figure 3A). Similar effects on adhesion and spreading on fibrinogen were observed after treatment with docosahexaenoic acid (Figure 3B in the [online-only Data Supplement](#)).

Outside-in signaling is also essential for the regulation of clot retraction, which is required for thrombus stabilization. Consistent with the inhibition of adhesion and spreading on fibrinogen, treatment of platelets with 9-cis-RA resulted in an inhibition of clot retraction because clot weight was increased after treatment with 9-cis-RA compared with vehicle-treated controls (Figure 3B). Similar observations for docosahexaenoic acid and methoprene acid support the notion that the effect on clot retraction is mediated by RXR activity (Figure 3C in the [online-only Data Supplement](#)).

### RXR Ligands Inhibit Thrombus Formation on Collagen Under Flow

As we have shown that RXR ligands affect several aspects of platelet activation, the effect of the 9-cis-RA on thrombus formation was assessed in vitro. Human whole blood was perfused over collagen-coated (100  $\mu$ g/mL) Vena8 biochips for 10 minutes at an arterial shear rate of 20 dynes/cm<sup>2</sup>. Consistent with the observed inhibition of platelet activity, a significant reduction in thrombus formation ( $\approx$ 35%) was observed in 9-cis-RA-treated whole blood in comparison to vehicle-treated control samples (Figure 4A). To determine whether this inhibition of thrombus formation was because of a reduction in the ability of platelets to adhere to collagen, platelets were treated with integrilin, an antagonist of integrin  $\alpha_{IIb}\beta_3$ , to prevent platelet-platelet interactions. As shown in Figure 4B, no significant difference in platelet adhesion on collagen was observed in platelets treated with 20  $\mu$ mol/L 9-cis-RA compared with vehicle-treated controls, suggesting that 9-cis-RA does not affect adhesion to collagen under flow but instead inhibits thrombus growth. This was further supported by observations under static conditions, where treatment with 9-cis-RA resulted in an inhibition of the ability of platelets to spread on collagen-coated coverslips, but adhesion to collagen was not significantly altered (Figure 4C).



**Figure 3.** The effect of 9-cis-retinoic acid (9-cis-RA) on integrin  $\alpha$ IIb $\beta$ 3 outside-in signaling. Human washed platelets pretreated for 10 min with increasing concentrations of 9-cis-RA (10 and 20  $\mu$ mol/L) or vehicle control were exposed to fibrinogen (100  $\mu$ g/mL) coated coverslips. **A (i)** representative images of spreading and adhesion after 45 min. Platelets were stained with phalloidin Alexa-488 for visualization. Images were taken under oil immersion with magnification  $\times 100$ . **(ii)** Adhesion: the number of platelets adhered were counted in 5 randomly selected fields of view, and the number of cells adhered were expressed as the percentage of the vehicle-treated control. **(iii)** Spreading platelets were classified into 3 different categories to determine the extent of their spreading (adhered but not spread; filopodia: platelets in the process of extending filopodia and lamellipodia: platelets in the process of extending lamellipodia including those fully spread). Results expressed (as relative frequency) as the percentage of the total number of platelets adhered. **B**, Clot retraction: human washed platelets pretreated for 10 min with increasing concentrations of 9-cis-RA (10 and 20  $\mu$ mol/L) or vehicle control were added to aggregometer tubes in the presence of 2 mg/mL fibrinogen and 2 mmol/L  $\text{CaCl}_2$ . Clot retraction was initiated by addition of thrombin 1 U/mL (final concentration) and left to proceed for 1 h at room temperature. Extent of clot retraction was determined by comparing clot weight. **(i)** Representative images using red blood cell stained platelet-rich plasma, **(ii)** data expressed as clot weight (mg). Unless stated otherwise, results are mean  $\pm$  SEM for  $n \geq 3$ . \* $P \leq 0.05$  in comparison to vehicle controls.

### RXR Ligands Inhibit Hemostasis and Thrombosis in Mice

To determine the impact of RXR ligands on the acute regulation of platelet function *in vivo*, the effect of 9-cis-RA on laser-induced thrombosis in mouse cremaster muscle arterioles was explored. Figure 4C shows that although the initial kinetics of thrombus formation were similar, thrombi formed in 9-cis-RA–treated mice were consistently smaller ( $\approx 35\%$ ) than those formed after treatment with vehicle control (0.1% dimethyl sulfoxide; Figure 4Cii and 4Ciii). These results suggest an antithrombotic effect of RXR ligands *in vivo*. 9-cis-RA was also found to impair hemostasis *in vivo* as a significant increase in time to cessation of bleeding was seen after removal of the tail tip of mice treated with 9-cis-RA compared with vehicle control (Figure 4D).

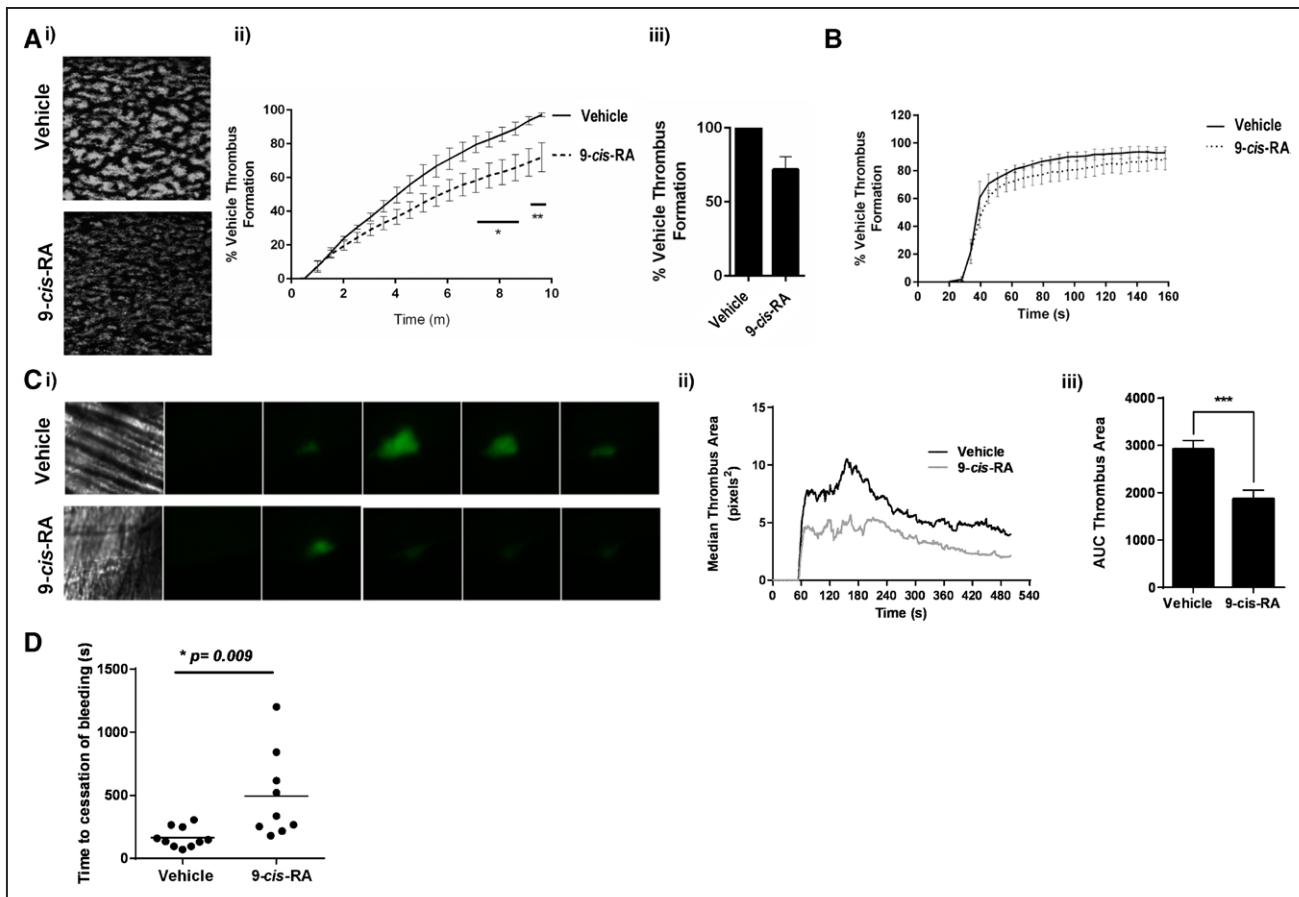
### 9-cis-RA Does Not Modulate Early GPVI Signaling Events

We have shown that platelet activation after stimulation by GPVI agonists and thrombin is reduced after treatment with RXR ligands. To determine whether this was because of alterations in early signaling events, the phosphorylation levels of different signaling proteins involved in the GPVI and thrombin-signaling pathways were analyzed. Washed human platelets were pretreated with 9-cis-RA (10 and 20  $\mu$ mol/L) or vehicle for 10 minutes before stimulation for 5 minutes by CRP-XL (1  $\mu$ g/mL) or thrombin (0.1 U/mL). Higher concentrations

of platelet agonists were used to enable phosphorylation of signaling components to be detected by Western blotting. Interestingly, despite the observed inhibition of platelet functions and thrombus formation, total tyrosine phosphorylation and phosphorylation of Syk and PLC $\gamma$ 2 (phospholipase C) after stimulation by CRP-XL were not altered by treatment with 9-cis-RA (10 and 20  $\mu$ mol/L; Figure IV A and IV B in the [online-only Data Supplement](#)). PKC (protein kinase C) activity was also assessed using an antibody raised against the phosphorylated PKC substrate recognition sequence. PKC activity after stimulation by either CRP-XL or thrombin for 5 minutes was unaffected by treatment with 9-cis-RA (Figure IV C and IV D in the [online-only Data Supplement](#)) but was reduced at earlier time points, with inhibition observed at 90 seconds (CRP-XL) and 30 seconds (thrombin). These observations indicate that the RXR agonists cause a reduction in the rate but not the magnitude of PKC activation.

### 9-cis-RA Increases PKA Activity in Resting and Stimulated Platelets

The data presented here demonstrates a role for RXR in the negative regulation of platelet function, although this is not associated with prominent alterations in the early platelet signaling events associated with GPVI agonists or thrombin. Such broad effects are unlikely to be explained by inhibition of specific elements of activation pathways; therefore, activation of inhibitory pathways was investigated. Treatment



**Figure 4.** The effect of retinoid X receptor (RXR) ligands on thrombus formation in vitro and in vivo. **A** and **B**, DiOC6-loaded human whole blood was pretreated with vehicle (black) or 20  $\mu$ mol/L 9-cis-retinoic acid (9-cis-RA; dotted) in the absence (**A**) or presence (**B**) of 10  $\mu$ mol/L integrilin for 10 min before perfusion through collagen-coated (100  $\mu$ g/mL) Vena8Biochips at an arterial shear rate of 20 dyn/cm<sup>2</sup>. Thrombus formation was determined after 10 min (**A**) or 3 min (**B**) by comparing fluorescence intensity in the vehicle- and 9-cis-RA-treated samples. (ii) Data expressed as percentage of vehicle control, where maximum fluorescence observed in vehicle-treated platelets is considered to be 100% thrombus formation. (iii) Data expressed as % of vehicle control after 10-min thrombus formation. **C**, In vivo thrombosis was assayed by intravital microscopy using the laser-induced injury model. 9-cis-RA (estimated concentration 20  $\mu$ mol/L) or vehicle (0.1% v/v dimethyl sulfoxide) was administered intravenously to mice, and platelets were fluorescently labeled with Alexa-488-conjugated anti-GPIb antibody. After laser injury, platelet accumulation and thrombus formation was assessed. (i) Representative images are shown, and data expressed as (ii) median thrombus area over time, and (iii) thrombus size determined by calculating the area under the curve. Between 8 and 10 thrombi were analyzed from 4 mice treated for each condition. **D**, Tail bleeding as determined as time to cessation of bleeding in mice pretreated with vehicle or 20  $\mu$ mol/L 9-cis-RA for 10 min ( $n=10$  for vehicle-treated and 9 for 9-cis-RA-treated samples). Unless stated otherwise, results are mean $\pm$ SEM for  $n\geq 3$ . \* $P\leq 0.05$  in comparison to vehicle controls.

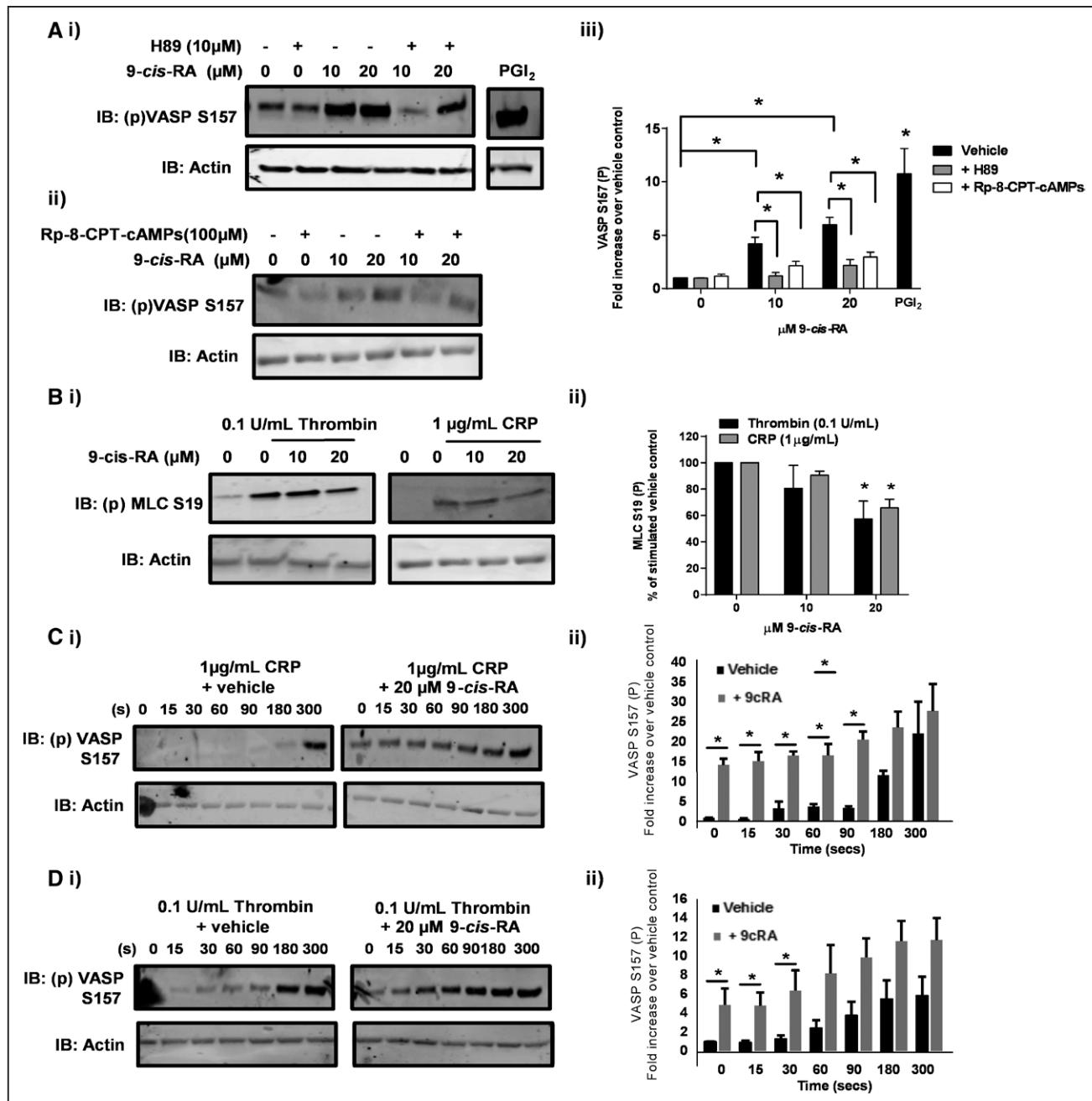
of platelets with ligands for PPARs has previously been described to cause upregulation of PKA activity in some cases via increasing intracellular levels of cAMP.<sup>2,4,12,27</sup> As we have identified that both PPAR $\alpha$  and PPAR $\gamma$  can heterodimerize with RXR in platelets, we determined whether or not RXR agonists were capable of altering PKA activity.

To determine whether RXR agonists altered PKA activity, VASP (vasodilator-stimulated phosphoprotein) S157 phosphorylation (a PKA-specific phosphorylation site and marker of PKA activity) was measured. Unstimulated platelets treated with 9-cis-RA (10 and 20  $\mu$ mol/L; Figure 5A) or methoprene acid (Figure V in the *online-only Data Supplement*; 10 and 20  $\mu$ mol/L) showed increased phosphorylation of VASP S157 in comparison to untreated controls. 9-cis-RA-dependent increase in VASP S157 phosphorylation was prevented after treatment with 2 different PKA inhibitors H89 (10  $\mu$ mol/L) and Rp-8-CPT-cAMPs (100  $\mu$ mol/L; Figure 5A). Similar

results were also observed after treatment with either docosahexaenoic acid or methoprene acid because both alternative RXR ligands also caused a significant increase in VASP S157 phosphorylation that was reversed after treatment with the PKA inhibitor H89 (Figure V in the *online-only Data Supplement*).

It has been shown that VASP can also be phosphorylated and regulated in platelets by the PKC isoforms and PKB/Akt. However, treatment with either a pan-PKC inhibitor GF109203X (10  $\mu$ mol/L) or AKT inhibitor, AKT inhibitor IV (5  $\mu$ mol/L), did not prevent the 9-cis-RA-induced increases in VASP S157 phosphorylation (Figure VI in the *online-only Data Supplement*), providing further support that RXR agonists exert their effect through PKA.

Activation of PKA has been shown to result in the negative regulation of the family of Rho GTPases including RhoA and Rac1 that have been identified as key regulators of platelet



**Figure 5.** Retinoid X receptor (RXR) ligands and protein kinase A (PKA) activity. Resting human washed platelets were treated with 9-cis-retinoic acid (9-cis-RA; 10 and 20  $\mu$ mol/L) in the presence or absence of (A) PKA inhibitors (i) H89 (10  $\mu$ mol/L) or (ii) Rp-8-CPT-cAMPs (100  $\mu$ mol/L) for 10 min and samples tested for VASP S157 phosphorylation, a marker of PKA activity. PGI<sub>2</sub>, which activates platelets through binding to the prostaglandin receptor (IP receptor) leading to activation of adenylyl cyclase, was included as a positive control for PKA activation. i and ii, Representative blots are shown and (iii) levels of phosphorylation were quantified and expressed as fold increase compared with vehicle control. **B**, Thrombin-stimulated (0.1 U/mL) or CRP-stimulated (1  $\mu$ g/mL) platelets (0- and 180-s stimulation) pretreated with 9-cis-RA (10 and 20  $\mu$ mol/L) were analyzed for myosin light chain (MLC) Ser19 phosphorylation. **C**, CRP-stimulated (1  $\mu$ g/mL) or (D) thrombin-stimulated (0.1 U/mL) platelets (0, 15, 30, 60, 90, 180, and 300 s of stimulation) pretreated with 9-cis-RA (20  $\mu$ mol/L) were also analyzed for VASP S157 phosphorylation. Blotting samples were lysed in Laemmli sample buffer before separation by SDS-PAGE and transferred onto polyvinylidene fluoride (PVDF) membranes. Actin was used as a loading control. i, Representative blots are shown and (ii) levels of phosphorylation were quantified and expressed as fold increase compared with vehicle control. Results are mean $\pm$ SEM for  $n\geq 3$ . \* $P\leq 0.05$  in comparison to vehicle controls.

function. Negative regulation of RhoA, in turn, negatively regulates cytoskeleton rearrangements and phosphorylation of the myosin light chain. In further support of the inhibition of platelet function by 9-cis-RA being caused by an increase in PKA

activity, we observed an inhibition of myosin light chain phosphorylation at Ser19 in both thrombin-treated (0.1 U/mL) and CRP-treated (1  $\mu$ g/mL) platelets after treatment with 9-cis-RA (10 and 20  $\mu$ mol/L) compared with vehicle controls (Figure 5B).

To investigate whether PKA activity was also altered in agonist-stimulated platelets, human washed platelets were pretreated with 9-*cis*-RA (20  $\mu$ mol/L) before stimulation with CRP-XL (1  $\mu$ g/mL) or thrombin (0.1 U/mL), and VASP S157 phosphorylation monitored over a period of 5 minutes. A time-dependent increase in VASP S157 phosphorylation was observed in both CRP-XL-stimulated and thrombin-stimulated platelets as has been described previously.<sup>24</sup> As shown in Figure 5C and 5D VASP S157 phosphorylation was significantly increased in 9-*cis*-RA-treated platelets before stimulation with either CRP-XL or thrombin and remained significantly higher than vehicle control at early time points. At later time points, no significant difference in VASP S157 phosphorylation was observed in 9-*cis*-RA-treated platelets compared with controls. This suggests that RXR agonists may exert their inhibitory effects via elevation of PKA activity, whereas CRP and thrombin activate PKA as a form of negative feedback once platelet activation has occurred.

### RXR Ligand-Mediated Increases in PKA Activity Are Dependent on Adenylyl Cyclase Activity but Not Prostaglandin I2 Receptor Signaling

Traditionally, activation of PKA occurs after the production of cAMP downstream of prostaglandin I2 (IP) receptor signaling. Binding of PGI<sub>2</sub> to the IP receptor causes Gs-coupled signaling that activates adenylyl cyclase and stimulates the production of cAMP, which then activates PKA. RXR ligand-mediated increases in PKA activity were found to be independent of IP receptor activation, as the IP receptor antagonist Ro1138452 (10  $\mu$ mol/L)<sup>28,29</sup> was unable to reverse 9-*cis*-RA-mediated increases in VASP S157 phosphorylation (Figure 6A). However, RXR ligand-dependent activation of PKA was attenuated after treatment with SQ22536 (100  $\mu$ mol/L), an adenylyl cyclase inhibitor, because 9-*cis*-RA-mediated (and docosahexaenoic acid-mediated) increases in VASP S157 phosphorylation were reduced after pretreatment with SQ22536 (Figure 6B). Interestingly, however, under the same experimental conditions, treatment with 9-*cis*-RA did not alter cAMP levels in resting platelets (Figure 6Ci). In the presence of the phosphodiesterase inhibitor, IBMX (1 mmol/L), a minor increase in cAMP was observed after treatment with 9-*cis*-RA (Figure 6Cii) but not after treatment with methoprene acid or docosahexaenoic acid (Figure VIIA and VIIIB in the *online-only Data Supplement*). These data suggest that RXR ligand-dependent increases in VASP S157 phosphorylation and PKA activity are dependent on intracellular cAMP but not associated with significant increases in cAMP levels.

### 9-*cis*-RA-Induced Increases in PKA Activity Are Associated With NF $\kappa$ B Activity

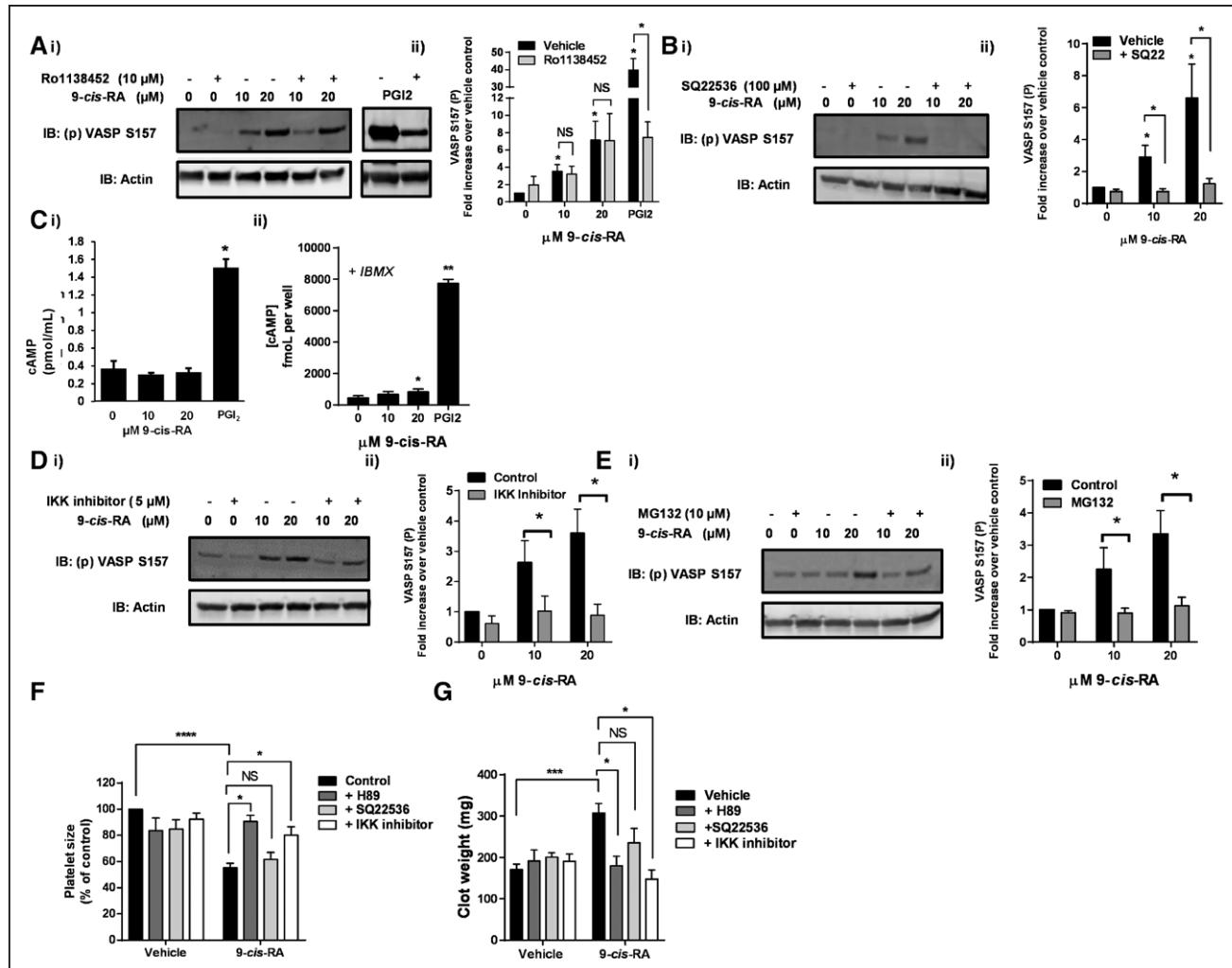
In addition to cAMP-mediated activation of PKA, PKA has also been shown to be activated via a mechanism that is dependent on NF $\kappa$ B, after stimulation of platelets by thrombin or collagen.<sup>24</sup> In this mechanism, a subpopulation of PKA is associated with NF $\kappa$ B–I $\kappa$ B $\alpha$  and after stimulation by collagen or thrombin, NF $\kappa$ B is activated, leading to degradation and release of I $\kappa$ B $\alpha$ .<sup>30</sup> This enables dissociation of the catalytic subunit of PKA from NF $\kappa$ B–I $\kappa$ B $\alpha$ , resulting in the activation

of PKA. To investigate whether RXR agonists activate PKA via NF $\kappa$ B–I $\kappa$ B $\alpha$ , we treated platelets with IKK inhibitor VII (5  $\mu$ mol/L) to determine whether it was able to reverse the observed platelet inhibition. IKK inhibitor VII was used previously to establish the role of NF $\kappa$ B in PKA activation and prevents the degradation and release of I $\kappa$ B $\alpha$ . IKK inhibitor VII prevented 9-*cis*-RA-induced VASP S157 phosphorylation in resting platelets, highlighting a role for the activation of NF $\kappa$ B and dissociation of I $\kappa$ B $\alpha$  in RXR-ligand-induced regulation of PKA activity (Figure 6D). Similar observations were also made after treatment with docosahexaenoic acid and methoprene acid (Figure VIIIC and VIIID in the *online-only Data Supplement*). Furthermore, treatment with a proteasome inhibitor, MG132 (10  $\mu$ mol/L), used to prevent degradation of I $\kappa$ B $\alpha$ , also prevented 9-*cis*-RA-induced phosphorylation of VASP S157 (Figure 6E). Both H89 and the IKK inhibitor, but not SQ22536 the adenylyl cyclase inhibitor, were found to reverse 9-*cis*-RA-mediated inhibition of platelet spreading on fibrinogen and clot retraction, implicating RXR-dependent upregulation of NF $\kappa$ B–I $\kappa$ B $\alpha$  and PKA in the negative regulation of platelet  $\alpha$ IIb $\beta$ <sub>3</sub> outside-in signaling (Figure 6F and 6G).

We have previously described that PPAR $\gamma$  ligands are capable of negatively regulating integrin  $\alpha$ IIb $\beta$ 3 outside-in signaling through upregulation of PKA activity.<sup>12</sup> As we found PPAR $\gamma$  can be coimmunoprecipitated with RXR from platelets, we determined whether it is the activation and modulation of RXR:PPAR $\gamma$  heterodimers that regulates the NF $\kappa$ B-mediated upregulation of PKA activity. However, the IKK inhibitor was unable to reverse 15dPGJ2-mediated (an endogenous ligand for PPAR $\gamma$ ) increases in VASP S157 phosphorylation (Figure VIIIA in the *online-only Data Supplement*), suggesting that PPAR $\gamma$ -dependent increases in PKA activity are not linked to the regulation of NF $\kappa$ B. In further support of this, the IKK inhibitor was also unable to reverse the increase in VASP S157 phosphorylation and PKA activity observed after treatment with the RXR modulator LG101506, which specifically modulates RXR:PPAR heterodimers (Figure VIIIB in the *online-only Data Supplement*). This suggests that ligands of RXR and PPAR $\gamma$  upregulate PKA activity via different mechanisms.

### Discussion

RXRs are intracellular nuclear receptors expressed in human platelets where they function to negatively regulate platelet responses to agonists.<sup>3,4,11,21</sup> In its genomic role, RXR is thought to heterodimerize with several intracellular nuclear receptors including PPARs, LXR, and FXR and is, therefore, also associated with the regulation of glucose, triglyceride, cholesterol, and bile acid homeostasis.<sup>20</sup> Dysregulation of these homeostatic control pathways can result in several metabolic disorders including obesity, type 2 diabetes mellitus, hyperlipidemia, atherosclerosis, and cardiovascular disease. Treatment with RXR ligands shows some efficacy at reducing the progression of atherosclerosis in apolipoprotein E knockout mice.<sup>18</sup> Alitretinoin, the commercial name for RXR ligand 9-*cis*-RA, is used for the treatment of Kaposi sarcoma and eczema, and decreased blood clotting is currently listed as one of its side effects.<sup>18,22,23</sup> In support of the ability of 9-*cis*-RA to



**Figure 6.** 9-cis-retinoic acid (9-cis-RA)-dependent increase in protein kinase A (PKA) activity is dependent on cAMP and nuclear factor κ-light-chain-enhancer of activated B cell (NFκB). Resting human washed platelets were treated with 9-cis-RA (10 and 20 μmol/L), (B) adenyl cyclase inhibitor SQ22536 (100 μmol/L), (D) IKK inhibitor (5 μmol/L), or (E) proteasome inhibitor MG132 (10 μmol/L) for 10 min, and samples tested for VASP S157 phosphorylation, a marker of PKA activity. Blotting samples were lysed in Laemmli sample buffer before separation on SDS-PAGE and transferred onto polyvinylidene fluoride (PVDF) membranes. Actin was used as a loading control. (i) Representative blots are shown and (ii) levels of phosphorylation were quantified and expressed as fold increase compared with vehicle control. C, Levels of cAMP were determined in 9-cis-RA-treated (10 and 20 μmol/L) resting washed platelets using a cAMP ELISA assay kit (as per manufacturer's instructions) in the (i) absence and (ii) presence of IBMX (1 mmol/L). PGI<sub>2</sub> was included as a positive control for PKA activation and intracellular increases in cAMP. F and G, Human washed platelets were treated with 9-cis-RA (20 μmol/L) in the presence or absence of H89 (10 μmol/L), IKK inhibitor (5 μmol/L) or SQ22536 (100 μmol/L) for 10 min (F) before exposure to fibrinogen-coated (100 μg/mL) coverslips and left to spread and adhere for 45 min. Data presented as average platelet size as calculated by looking at individual platelet surface area of platelets adhered in 5 randomly selected fields of view using ImageJ software. The larger the surface area, the more spread the platelet is. G, In the presence of 2 mg/mL fibrinogen and 2 mmol/L CaCl<sub>2</sub>, in aggregometer tubes, clot retraction was initiated by addition of thrombin 1 U/mL (final concentration) and left to proceed for 1 h at room temperature. Extent of clot retraction was determined by comparing clot weight. Results are mean±SEM for n≥3. \*P≤0.05 in comparison to vehicle controls.

reduce blood clotting, we show that treatment with 9-cis-RA increases bleeding time in mice and elicits other antithrombotic effects, which we attribute to an activation of PKA via a mechanism that requires cAMP and involves NFκB. As platelets play an important role in the pathogenesis of cardiovascular disease, our findings that RXR ligands inhibit platelet activation and thrombus formation may help to explain the efficacy of these compounds in vivo.

We observed that treatment of platelets with RXR agonists 9-cis-RA, methoprene acid, and docosahexaenoic acid caused a significant inhibition of α-granule secretion; mobilization

of intracellular calcium; platelet aggregation and integrin α<sub>IIb</sub>β<sub>3</sub> activation to several platelet agonists including collagen, CRP-XL, and thrombin; and an inhibition of integrin α<sub>IIb</sub>β<sub>3</sub> outside-in signaling. This inhibition of platelet activity by 9-cis-RA also correlated with inhibition of thrombus formation in vitro and in vivo, suggesting that the previously described cardioprotective effects of RXR agonists in reducing atherosclerosis<sup>18</sup> could also potentially be attributed to their negative regulation of platelet function.

Treatment of platelets with ligands for PPARs, PPARα and PPARβ/δ or PPARγ common binding partners for RXR

(Figure 1) have been previously described to cause upregulation of cAMP levels or activation of PKA. RXR ligand–stimulated inhibition of platelet function and thrombus formation were also found to be associated with an upregulation of PKA activity, as treatment of platelets with RXR agonists resulted in an increase in VASP phosphorylation at S157 (the PKA phosphorylation site) in both resting and agonist-stimulated platelets, and this increase was reversed after treatment with the PKA inhibitors H89 and Rp-8-CPT-cAMPs or adenylyl cyclase inhibitor SQ22358 but not after treatment with an IP receptor antagonist (Ro1138452). This suggests that RXR agonists activate PKA through a mechanism that is dependent on cAMP, although no major alterations in cAMP levels were observed after treatment with the different RXR agonists. It is possible that treatment with RXR agonists does cause small increases in platelet cAMP levels that are not detected given current limitations in sensitivity of the assays used. It has been shown that even minor increases in cAMP levels can cause significant activation of cellular PKA and large increases in VASP S157 phosphorylation.<sup>31</sup> As such, a role for RXR ligands in the upregulation of adenylyl cyclase activity cannot be ruled out. It is, however, interesting to note that RXR ligand–mediated inhibition of integrin  $\alpha$ IIb $\beta$ 3 outside-in signaling cannot be reversed by inhibition of adenylyl cyclase, suggesting a role for another PKA-linked signaling pathway in this negative regulation of platelet function.

It has been previously shown that activation of NF $\kappa$ B is involved in regulation of PKA activity after platelet activation by collagen or thrombin, where the inactive NF $\kappa$ B–I $\kappa$ B $\alpha$  complex binds to and inactivates the PKA catalytic subunit.<sup>24</sup> In cancer cells, activation of NF $\kappa$ B has also been shown to occur after treatment with RXR ligands, including 9-*cis*-RA, and inhibition or downregulation of NF $\kappa$ B has been shown to reduce RXR-mediated cell differentiation and apoptosis of cancerous cells.<sup>32</sup> We, therefore, hypothesized that treatment with RXR ligands could enable targeted degradation of the inactive NF $\kappa$ B–I $\kappa$ B $\alpha$  complex, releasing the PKA catalytic subunit and relieving the inhibition of PKA, resulting in an increase in PKA activity and subsequent substrate phosphorylation. This was supported by observations that treatment with H89 and the IKK inhibitor prevented 9-*cis*-RA–mediated upregulation of PKA activity and also reversed 9-*cis*-RA–mediated inhibition of platelet spreading on fibrinogen and clot retraction, which is dependent on integrin  $\alpha$ IIb $\beta$ 3 outside-in signaling. It is interesting to note, however, that the upregulation of PKA activity via NF $\kappa$ B is not a mechanism that is shared with the PPARs, typical binding partners of RXR, as the increase in VASP S157 phosphorylation observed after treatment with the RXR:PPAR modulator LG101506 could not be reversed by pretreatment with the IKK inhibitor.

Activation of PKA has been shown to negatively regulate the family of Rho GTPases including RhoA and Rac1.<sup>24,33,34</sup> Rho GTPases including Rac1 are key mediators of platelet function, and mice deficient in Rac1 display major defects in platelet activation downstream of both GPVI and GPCRs and reduced thrombus formation in vivo.<sup>35–37</sup> Previously published work suggests that Rac1 activity is reduced after treatment of platelets with RXR ligands, and this is attributed to an interaction of RXR with Gq.<sup>5</sup> In agreement with alteration of RhoA

activity, a reduction in myosin light chain phosphorylation was observed after treatment with 9-*cis*-RA. The upregulation of PKA activity that we describe here could also provide an additional mechanism that underlies this reduction in Rac1 activity that may explain the broad-spectrum inhibition by RXR ligands of platelet activity to multiple platelet agonists. Because RXR has the potential to form heterodimers with several different nuclear receptors, multiple mechanisms of regulation could exist.

The data presented here significantly build on previously observed inhibitory effects of RXR ligands on platelet activity<sup>5</sup> and highlight a relatively unknown mechanism of PKA activation as a potential target for antiplatelet therapy. The data presented here suggest that RXR ligands could offer extra protective effects in vivo if developed as drug targets for the treatment of other diseases such as diabetes mellitus, although these effects would need to be carefully balanced to ensure there is no increased risk of bleeding.

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A.J. Unsworth designed the research, performed experiments, analyzed results, and wrote the article. G.D. Flora designed the research, performed experiments, and analyzed results. A.P. Bye performed experiments, analyzed results, and wrote the article. P. Sasikumar, N. Kriek, T. Sage, and M. Crescente performed experiments. J.M. Gibbins designed the research and wrote the article.

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## Disclosures

None.

## References

1. Bishop-Bailey D. The platelet as a model system for the acute actions of nuclear receptors. *Steroids*. 2010;75:570–575. doi: 10.1016/j.steroids.2009.09.005.
2. Ali FY, Davidson SJ, Moraes LA, Traves SL, Paul-Clark M, Bishop-Bailey D, Warner TD, Mitchell JA. Role of nuclear receptor signaling in platelets: antithrombotic effects of PPARbeta. *FASEB J*. 2006;20:326–328. doi: 10.1096/fj.05-4395fje.
3. Moraes LA, Spyridon M, Kaiser WJ, Jones CI, Sage T, Atherton RE, Gibbins JM. Non-genomic effects of PPARgamma ligands: inhibition of GPVI-stimulated platelet activation. *J Thromb Haemost*. 2010;8:577–587. doi: 10.1111/j.1538-7836.2009.03732.x.
4. Ali FY, Armstrong PC, Dhanji AR, Tucker AT, Paul-Clark MJ, Mitchell JA, Warner TD. Antiplatelet actions of statins and fibrates are mediated by PPARs. *Arterioscler Thromb Vasc Biol*. 2009;29:706–711. doi: 10.1161/ATVBAHA.108.183160.
5. Moraes LA, Swales KE, Wray JA, Damazo A, Gibbins JM, Warner TD, Bishop-Bailey D. Nongenomic signaling of the retinoid X receptor through binding and inhibiting Gq in human platelets. *Blood*. 2007;109:3741–3744. doi: 10.1182/blood-2006-05-022566.
6. Ali FY, Hall MG, Desvergne B, Warner TD, Mitchell JA. PPARbeta/delta agonists modulate platelet function via a mechanism involving PPAR receptors and specific association/repression of PKCalpha—brief report. *Arterioscler Thromb Vasc Biol*. 2009;29:1871–1873. doi: 10.1161/ATVBAHA.109.193367.
7. Akbiyik F, Ray DM, Gettings KF, Blumberg N, Francis CW, Phipps RP. Human bone marrow megakaryocytes and platelets express PPARgamma, and PPARgamma agonists blunt platelet release of CD40 ligand and thromboxanes. *Blood*. 2004;104:1361–1368. doi: 10.1182/blood-2004-03-0926.
8. Li D, Chen K, Sinha N, Zhang X, Wang Y, Sinha AK, Romeo F, Mehta JL. The effects of PPAR-gamma ligand pioglitazone on platelet aggregation

- and arterial thrombus formation. *Cardiovasc Res.* 2005;65:907–912. doi: 10.1016/j.cardiores.2004.11.027.
- Moraes LA, Paul-Clark MJ, Rickman A, Flower RJ, Goulding NJ, Perretti M. Ligand-specific glucocorticoid receptor activation in human platelets. *Blood.* 2005;106:4167–4175. doi: 10.1182/blood-2005-04-1723.
  - Ray DM, Spinelli SL, Pollock SJ, Murant TI, O'Brien JJ, Blumberg N, Francis CW, Taubman MB, Phipps RP. Peroxisome proliferator-activated receptor gamma and retinoid X receptor transcription factors are released from activated human platelets and shed in microparticles. *Thromb Haemost.* 2008;99:86–95. doi: 10.1160/TH07-05-0328.
  - Spiridon M, Moraes LA, Jones CI, Sage T, Sasikumar P, Bucci G, Gibbins JM. LXR as a novel antithrombotic target. *Blood.* 2011;117:5751–5761. doi: 10.1182/blood-2010-09-306142.
  - Unsworth AJ, Kriek N, Bye AP, Naran K, Sage T, Flora GD, Gibbins JM. PPAR $\gamma$  agonists negatively regulate  $\alpha IIb\beta 3$  integrin outside-in signaling and platelet function through up-regulation of protein kinase A activity. *J Thromb Haemost.* 2017;15:356–369. doi: 10.1111/jth.13578.
  - Mangelsdorf DJ, Borgmeyer U, Heyman RA, Zhou JY, Ong ES, Oro AE, Kakizuka A, Evans RM. Characterization of three RXR genes that mediate the action of 9-cis retinoic acid. *Genes Dev.* 1992;6:329–344.
  - Kliewer SA, Umesono K, Mangelsdorf DJ, Evans RM. Retinoid X receptor interacts with nuclear receptors in retinoic acid, thyroid hormone and vitamin D3 signalling. *Nature.* 1992;355:446–449. doi: 10.1038/355446a0.
  - Mangelsdorf DJ, Evans RM. The RXR heterodimers and orphan receptors. *Cell.* 1995;83:841–850.
  - Nagy L, Thomazy VA, Heyman RA, Davies PJ. Retinoid-induced apoptosis in normal and neoplastic tissues. *Cell Death Differ.* 1998;5:11–19.
  - Tyagi S, Gupta P, Saini AS, Kaushal C, Sharma S. The peroxisome proliferator-activated receptor: a family of nuclear receptors role in various diseases. *J Adv Pharm Technol Res.* 2011;2:236–240. doi: 10.4103/2231-4040.90879.
  - Claudel T, Leibowitz MD, Fiévet C, Tailleux A, Wagner B, Repa JJ, Torpier G, Lobaccaro J-M, Paterniti JR, Mangelsdorf DJ. Reduction of atherosclerosis in apolipoprotein E knockout mice by activation of the retinoid X receptor. *Proc Natl Acad Sci.* 2001;98:2610–2615.
  - Leibowitz MD, Ardecky RJ, Boehm MF, et al. Biological characterization of a heterodimer-selective retinoid X receptor modulator: potential benefits for the treatment of type 2 diabetes. *Endocrinology.* 2006;147:1044–1053. doi: 10.1210/en.2005-0690.
  - Moraes LA, Piqueras L, Bishop-Bailey D. Peroxisome proliferator-activated receptors and inflammation. *Pharmacol Ther.* 2006;110:371–385. doi: 10.1016/j.pharmthera.2005.08.007.
  - Ray DM, Spinelli SL, O'Brien JJ, Blumberg N, Phipps RP. Platelets as a novel target for PPARgamma ligands: implications for inflammation, diabetes, and cardiovascular disease. *BioDrugs.* 2006;20:231–241.
  - Walmsley S, Northfelt DW, Melosky B, Conant M, Friedman-Kien AE, Wagner B. Treatment of AIDS-related cutaneous Kaposi's sarcoma with topical altretinoin (9-cis-retinoic acid) gel. Panretin Gel North American Study Group. *J Acquir Immune Defic Syndr.* 1999;22:235–246.
  - Ghasri P, Scheinfeld N. Update on the use of altretinoin in treating chronic hand eczema. *Clin Cosmet Investig Dermatol.* 2010;3:59–65.
  - Gambaryan S, Kobsar A, Rukoyatkina N, Herterich S, Geiger J, Smolenski A, Lohmann SM, Walter U. Thrombin and collagen induce a feedback inhibitory signaling pathway in platelets involving dissociation of the catalytic subunit of protein kinase A from an NF $\kappa$ B-I $\kappa$ B complex. *J Biol Chem.* 2010;285:18352–18363. doi: 10.1074/jbc.M109.077602.
  - Rowley JW, Oler AJ, Tolley ND, Hunter BN, Low EN, Nix DA, Yost CC, Zimmerman GA, Weyrich AS. Genome-wide RNA-seq analysis of human and mouse platelet transcriptomes. *Blood.* 2011;118:e101–e111. doi: 10.1182/blood-2011-03-339705.
  - Bye AP, Unsworth AJ, Gibbins JM. Platelet signaling: a complex interplay between inhibitory and activatory networks. *J Thromb Haemost.* 2016;14:918–930. doi: 10.1111/jth.13302.
  - Du H, Hu H, Zheng H, Hao J, Yang J, Cui W. Effects of peroxisome proliferator-activated receptor  $\gamma$  in simvastatin antiplatelet activity: influences on cAMP and mitogen-activated protein kinases. *Thromb Res.* 2014;134:111–120. doi: 10.1016/j.thromres.2014.05.005.
  - Bley KR, Bhattacharya A, Daniels DV, Gever J, Jahangir A, O'Yang C, Smith S, Srinivasan D, Ford AP, Jett MF. RO1138452 and RO3244794: characterization of structurally distinct, potent and selective IP (prostacyclin) receptor antagonists. *Br J Pharmacol.* 2006;147:335–345. doi: 10.1038/sj.bjp.0706554.
  - Jones RL, Wise H, Clark R, Whiting RL, Bley KR. Investigation of the prostacyclin (IP) receptor antagonist RO1138452 on isolated blood vessel and platelet preparations. *Br J Pharmacol.* 2006;149:110–120. doi: 10.1038/sj.bjp.0706841.
  - Zhong H, SuYang H, Erdjument-Bromage H, Tempst P, Ghosh S. The transcriptional activity of NF- $\kappa$ B is regulated by the I $\kappa$ B-associated PKA $\alpha$  subunit through a cyclic AMP-independent mechanism. *Cell.* 1997;89:413–424.
  - Eigenthaler M, Nolte C, Halbrügge M, Walter U. Concentration and regulation of cyclic nucleotides, cyclic-nucleotide-dependent protein kinases and one of their major substrates in human platelets. Estimating the rate of cAMP-regulated and cGMP-regulated protein phosphorylation in intact cells. *Eur J Biochem.* 1992;205:471–481.
  - Jiménez-Lara AM, Aranda A, Gronemeyer H. Retinoic acid protects human breast cancer cells against etoposide-induced apoptosis by NF- $\kappa$ B-dependent but cIAP2-independent mechanisms. *Mol Cancer.* 2010;9:15. doi: 10.1186/1476-4598-9-15.
  - Nagy Z, Wynne K, von Kriegsheim A, Gambaryan S, Smolenski A. Cyclic nucleotide-dependent protein kinases target ARHGAP17 and ARHGEF6 complexes in platelets. *J Biol Chem.* 2015;290:29974–29983. doi: 10.1074/jbc.M115.678003.
  - Aburima A, Wraith KS, Raslan Z, Law R, Magwenzsi S, Naseem KM. cAMP signaling regulates platelet myosin light chain (MLC) phosphorylation and shape change through targeting the RhoA-Rho kinase-MLC phosphatase signaling pathway. *Blood.* 2013;122:3533–3545. doi: 10.1182/blood-2013-03-487850.
  - McCarty OJ, Larson MK, Auger JM, Kalia N, Atkinson BT, Pearce AC, Ruf S, Henderson RB, Tybulewicz VL, Machesky LM, Watson SP. Rac1 is essential for platelet lamellipodia formation and aggregate stability under flow. *J Biol Chem.* 2005;280:39474–39484. doi: 10.1074/jbc.M504672200.
  - Delaney MK, Liu J, Zheng Y, Berndt MC, Du X. The role of Rac1 in glycoprotein Ib-IX-mediated signal transduction and integrin activation. *Arterioscler Thromb Vasc Biol.* 2012;32:2761–2768. doi: 10.1161/ATVBAHA.112.254920.
  - Pleines I, Elvers M, Strehl A, Pozgajova M, Varga-Szabo D, May F, Chrostek-Grashoff A, Brakebusch C, Nieswandt B. Rac1 is essential for phospholipase C-gamma2 activation in platelets. *Pflugers Arch.* 2009;457:1173–1185. doi: 10.1007/s00424-008-0573-7.

## Highlights

- Platelets express the retinoid X receptors (RXR $\alpha$  and RXR $\beta$ ). RXR ligands cause a widespread inhibition of platelet function to multiple platelet agonists.
- RXR ligands inhibit thrombus formation and impair hemostasis.
- This nongenomic regulation by RXR ligands is mediated by activation of PKA that involves cAMP and nuclear factor  $\kappa$ -light-chain-enhancer of activated B cell.

# Arteriosclerosis, Thrombosis, and Vascular Biology

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## RXR Ligands Negatively Regulate Thrombosis and Hemostasis

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## MATERIALS AND METHODS

### **RXR ligands negatively regulate thrombosis and haemostasis.**

*A.J. Unsworth, G. Flora, P. Sasikumar, A.P. Bye, T. Sage, N. Kriek, M.Crescente, J.M. Gibbins.*

## MATERIALS AND METHODS

### Reagents

9-cis-retinoic acid, docosahexaenoic acid, methoprene acid, bovine thrombin, H89, SQ22536 and MG132 were purchased from Sigma Aldrich (Poole, UK). Rp-8-CPTs-cAMP and Ro1138452 were purchased from Tocris. Horm collagen was purchased from Nycomed, Austria, CRP-XL from Prof. R Farndale (University of Cambridge, UK). Primary anti- RXR, Syk (N-19), PLC $\gamma$ 2 (Q20) and actin (C11) antibodies were purchased from Santa Cruz Biotechnology (Calne, UK). Anti-Phospho-PKC substrate antibody, phospho-myosin light chain S19 and phospho-Ser157 VASP antibodies were purchased from New England BioLabs, USA (Cell Signalling Hitchin, UK), anti-phospho-Tyr 4G10 antibody and IKK inhibitor VII were purchased from Millipore (Watford, UK). Fluorophore conjugated secondary antibodies, Fura-2AM calcium indicator dye and Alexa-488 conjugated phalloidin were purchased from Life Technologies (Paisley, UK). All other reagents were from previously described sources <sup>[1, 2]</sup>.

### Platelet preparation

Human blood was obtained from consenting aspirin-free, healthy volunteers following procedures approved by the University of Reading Research Ethics Committee. Blood was collected into 3.8% (w/v) sodium citrate before mixing with acid citrate dextrose (29.9 mM Na<sub>3</sub>C<sub>6</sub>H<sub>5</sub>O<sub>7</sub>, 113.8 mM glucose, 72.6 mM NaCl, and 2.9 mM citric acid [pH 6.4]). Human washed platelets were prepared by centrifugation as described previously <sup>[3]</sup>. Platelets were resuspended in modified Tyrode's-HEPES buffer, (134mM NaCl, 0.34mM Na<sub>2</sub>HPO<sub>4</sub>, 2.9mM KCl, 12mM NaHCO<sub>3</sub>, 20mM N-2-hydroxyethylpiperazine-N-2-ethanesulfonic acid, 5mM glucose and 1mM MgCl<sub>2</sub>, pH 7.3) and rested for 30 minutes at 30°C before use.

### Immunofluorescence microscopy

Human and mouse platelets stimulated with or without U46619 (3 $\mu$ M) were left to settle on poly-L-lysine coverslips for 1 hour at 37°C before permeabilisation and blocking (0.2% Triton-X-100, 1% BSA, 2% donkey serum). Coverslips were then incubated with primary antibodies for RXR and CD41 (a marker of the platelet membrane) overnight at 4°C and washed in PBS before staining with Alexa-fluorophore conjugated secondary antibodies (488 nm and 647 nm) for 1 hour at room temperature in the dark. Coverslips were washed and mounted onto slides. Platelets were imaged with a 100 x magnification oil immersion lens on a Nikon A1-R confocal microscope.

### Platelet aggregation

Aggregation of human washed platelets was measured by optical aggregometry (Chrono-log Corp., Havertown, PA, USA) as described previously <sup>[4]</sup>.

### Fibrinogen binding and alpha granule secretion

Activation of the integrin  $\alpha_{IIb}\beta_3$  and alpha granule secretion were measured by detecting levels of fibrinogen binding and P-selectin exposure at the platelet surface by flow cytometry using fluorescein isothiocyanate-labelled (FITC) anti-fibrinogen antibody and PE/Cy5 anti-human CD62P respectively. Using a BD Accuri C6 flow cytometer, 5,000 events were analysed using the CFlow Sampler software as described previously <sup>[5]</sup>.

### Intracellular Calcium Levels

PRP was loaded with Fura-2 AM (2  $\mu$ M) for 1h at 30°C and then washed by centrifugation at 350 xg for 20 mins and resuspended in Tyrode's-HEPES buffer containing 0.4 U/ml apyrase. Fura-2AM loaded platelets were incubated with inhibitors or vehicle at 37°C for 10 minutes prior to addition of agonists. Fluorescence measurements with excitation at 340 and 380 nm

and emission at 510 nm were recorded over a period of 5 mins using a NOVOstar plate reader (BMG Labtech).  $[Ca^{2+}]_i$  was estimated using the ratio of the 340 and 380 nm excited signals, using the method of Gryniewicz *et al*<sup>[6]</sup> and  $[Ca^{2+}]_i$  concentrations were calculated as described previously<sup>[7, 8]</sup>.

### **Adhesion and spreading on fibrinogen**

Washed platelets ( $2 \times 10^7$  cells/mL), treated with or without RXR ligands were exposed to fibrinogen (100  $\mu$ g/ml) coated coverslips and incubated for 45 minutes at 37°C. Non adherent platelets were removed before fixing using 0.2% paraformaldehyde solution. Adhered platelets were permeabilised with 0.1% Triton-X-100 prior to staining with Alexa 488 conjugated-phalloidin for 1 hr at room temperature. Adherent platelets were then imaged with a 100 x magnification oil immersion lens on a Nikon A1-R confocal microscope. Adhesion and spreading data in each experiment were obtained by counting, for each sample, the number of platelets in 5 randomly chosen fields of view. The number of platelets generating filopodia or lamellipodia were also counted, and platelets scored as being adhered but not spread, extending filopodia and fully spread (formation of lamellipodia) and the relative frequency determined.

### **Clot retraction assay**

Human washed platelets at  $5 \times 10^8$ /mL were added to aggregometer tubes in the presence of 2 mg/mL fibrinogen and 2 mM CaCl<sub>2</sub>. Clot formation was initiated by adding an equal volume of 2 U/mL thrombin and allowed to progress for 1 hour at room temperature. Weight of the clot and volume of extruded serum were measured.

### **Thrombus formation on collagen**

Thrombus formation in the presence or absence of integrillin (10  $\mu$ M) was studied *in vitro* using microfluidic flow cells (Vena8, CellixLtd, Dublin, Ireland) coated with collagen (100  $\mu$ g/mL). Blood was passed through the flow cells at an arterial shear rate of 20 dyn/cm<sup>2</sup> as described previously<sup>[5]</sup>.

### **Tail bleeding assay**

Tail bleeding experiments were performed on 20–35 g male mice, anesthetized with ketamine (100 mg/kg) and xylazine (10 mg/kg) injected intraperitoneally. RXR ligand 9-cis-RA (20  $\mu$ M) or vehicle control (DMSO 0.1% v/v), calculated by taking into consideration mouse weight and blood volume, was injected into the femoral vein 10 minutes prior to removal of the tip of the tail using a sharp razor blade. The tail tip was then placed in sterile saline (37 °C) and time to cessation of bleeding (secs) measured.

### **Laser injury induced thrombus formation**

*In vivo* thrombosis was assayed using a laser injury model by intravital microscopy as described previously<sup>[9]</sup>. In brief, treatment with RXR ligand 9-cis-RA (20  $\mu$ M) or vehicle control (DMSO 0.1% v/v), as calculated by taking into consideration mouse weight and blood volume, was administered intravenously to mice and platelets fluorescently labelled by injection of Alexa 488-conjugated anti GPIb antibody for 10 minutes prior to laser injury. After laser induced injury of the inner wall of the cremaster muscle arterioles, accumulation of platelets was assessed. Fluorescence and brightfield images were recorded using an Olympus BX61W microscope with a 60 x/1.0 NA water immersion objective and a high speed camera, and data analyzed using Image J software.

### **Immunoblotting and immunoprecipitation**

Washed platelets ( $4 \times 10^8$  cells/mL) were lysed in an equal volume of NP40 buffer (300 mM NaCl, 20 mM Tris base, 2 mM EGTA, 2 mM EDTA, 1 mM PMSF, 10  $\mu$ g/ml aprotinin, 10

µg/ml leupeptin, 0.7µg/ml pepstatin A, 2mM sodium orthovanadate, 2% NP-40, pH 7.3), and proteins of interest were isolated using 1 µg/mL of appropriate antibodies as described previously [4]. Immunoblotting was performed using standard techniques as described previously [3].

Levels of phosphorylated proteins were detected using fluorophore conjugated secondary antibodies and visualised using a Typhoon Trio Fluorimager and Image Quant software (GE Healthcare). Band intensities were quantified and levels of the immunoprecipitated protein were used to control for protein loading using Image Quant software.

### **Detection of cAMP levels.**

Human washed platelets ( $2 \times 10^8$  cells/mL) were treated with RXR ligands in the presence and absence of phosphodiesterase inhibitor IBMX (1mM) for 10 minutes, lysed in lysis buffer provided and cAMP levels measured using a cAMP ELISA kit (ENZO Life sciences) and (GE Healthcare) respectively as per manufacturers instructions, and as described previously [10] [11].

### **Statistical analysis**

Statistical analyses were performed using GraphPad prism software. Data were analysed using student T-test and if more than two means were present, significance was determined by one way ANOVA. Values obtained in several experiments were converted into percentages for comparison of controls with treated samples or expressed as fold change compared to control. Where data was normalised, statistical analysis was performed prior to normalisation and also using the non-parametric Wilcoxon signed-rank test.  $P \leq 0.05$  was considered statistically significant. Unless stated otherwise, values are expressed as mean  $\pm$ SEM, n values are  $\geq 3$ .

1. Bye AP, Unsworth AJ, Vaiyapuri S, et al., Ibrutinib Inhibits Platelet Integrin alphaiIbbeta3 Outside-In Signaling and Thrombus Stability But Not Adhesion to Collagen. *Arterioscler Thromb Vasc Biol*, 2015.
2. Jones CI, Moraes LA, and Gibbins JM, Regulation of platelet biology by platelet endothelial cell adhesion molecule-1. *Platelets*, 2012; 23(5):331-5.
3. Kaiser WJ, Holbrook LM, Tucker KL, Stanley RG, and Gibbins JM, A functional proteomic method for the enrichment of peripheral membrane proteins reveals the collagen binding protein Hsp47 is exposed on the surface of activated human platelets. *J Proteome Res*, 2009; 8(6):2903-14.
4. Moraes LA, Spyridon M, Kaiser WJ, et al., Non-genomic effects of PPARgamma ligands: inhibition of GPVI-stimulated platelet activation. *J Thromb Haemost*, 2010; 8(3):577-87.
5. Vaiyapuri S, Jones CI, Sasikumar P, et al., Gap junctions and connexin hemichannels underpin hemostasis and thrombosis. *Circulation*, 2012; 125(20):2479-91.
6. Grynkiewicz G, Poenie M, and Tsien RY, A new generation of Ca<sup>2+</sup> indicators with greatly improved fluorescence properties. *J Biol Chem*, 1985; 260(6):3440-50.
7. Poenie M, Alteration of intracellular Fura-2 fluorescence by viscosity: a simple correction. *Cell Calcium*, 1990; 11(2-3):85-91.
8. Bye AP, Unsworth AJ, Vaiyapuri S, et al., Ibrutinib Inhibits Platelet Integrin alphaiIbbeta3 Outside-In Signaling and Thrombus Stability But Not Adhesion to Collagen. *Arterioscler Thromb Vasc Biol*, 2015; 35(11):2326-35.
9. Spyridon M, Moraes LA, Jones CI, et al., LXR as a novel antithrombotic target. *Blood*, 2011; 117(21):5751-61.

10. Moraes LA, Unsworth AJ, Vaiyapuri S, et al., Farnesoid X Receptor and Its Ligands Inhibit the Function of Platelets. *Arterioscler Thromb Vasc Biol*, 2016.
11. Aburima A, Wraith KS, Raslan Z, et al., cAMP signaling regulates platelet myosin light chain (MLC) phosphorylation and shape change through targeting the RhoA-Rho kinase-MLC phosphatase signaling pathway. *Blood*, 2013; 122(20):3533-45.

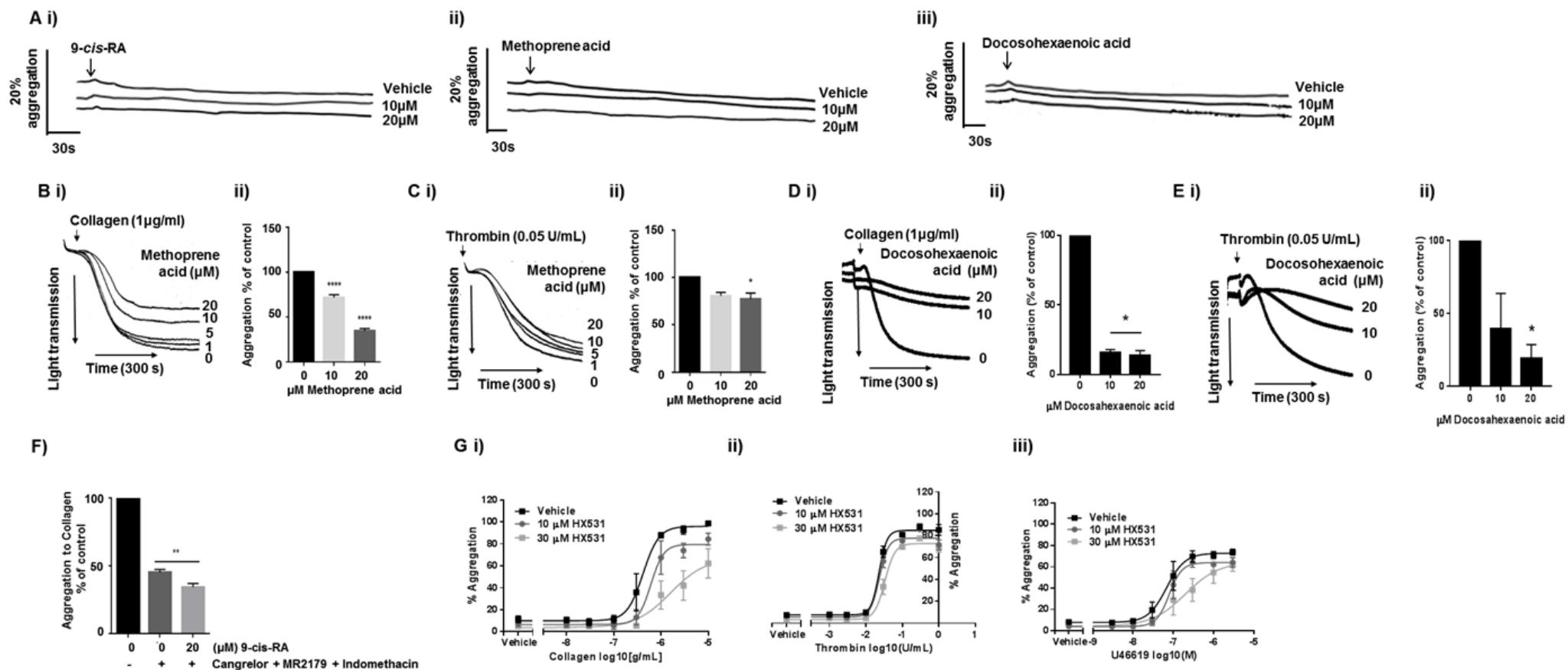
## SUPPLEMENTAL MATERIAL

### RXR ligands negatively regulate thrombosis and haemostasis.

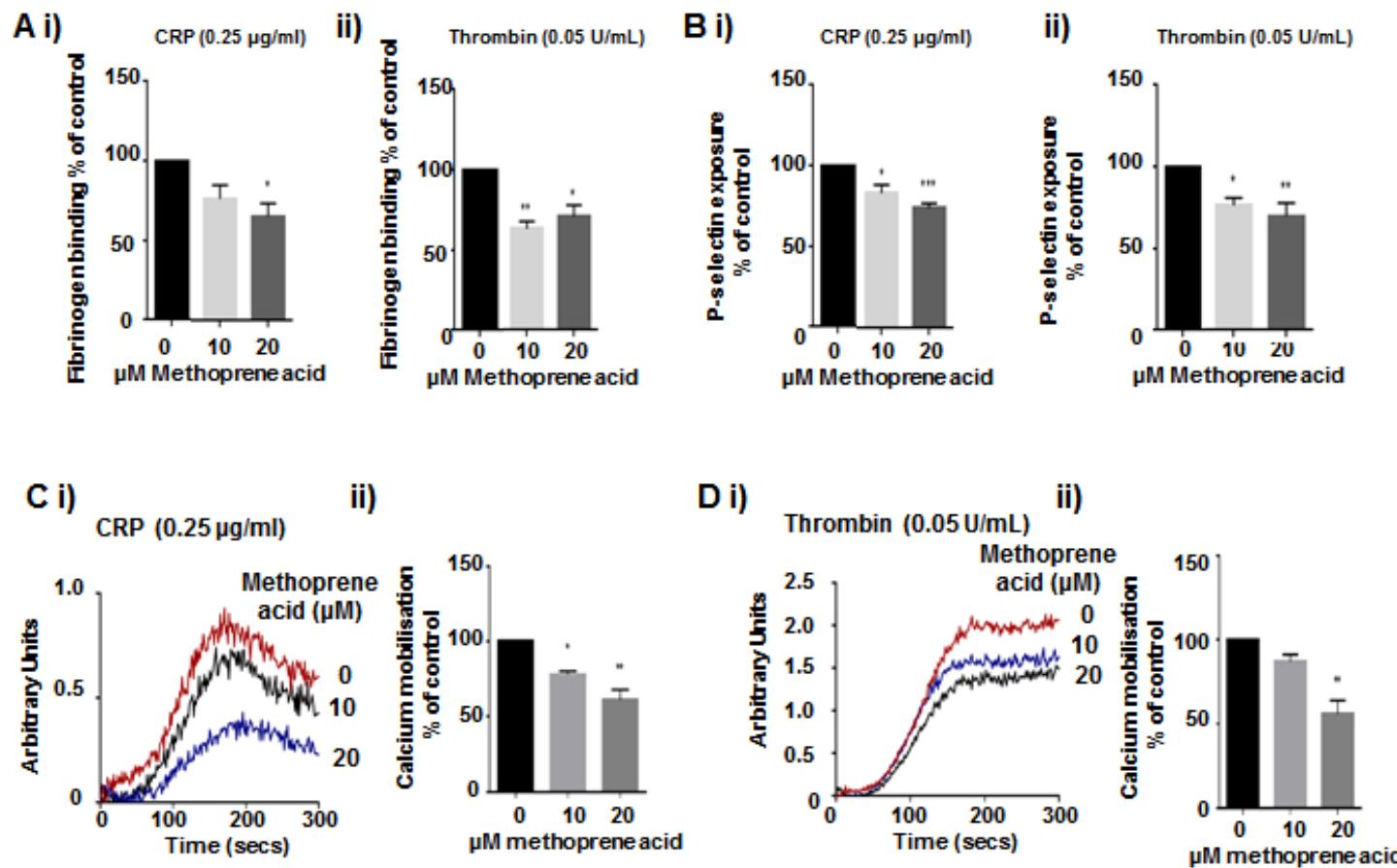
*A.J. Unsworth, G. Flora, P. Sasikumar, A.P. Bye, T. Sage, N. Kriek, M.Crescente, J.M. Gibbins.*

## SUPPLEMENTAL MATERIAL

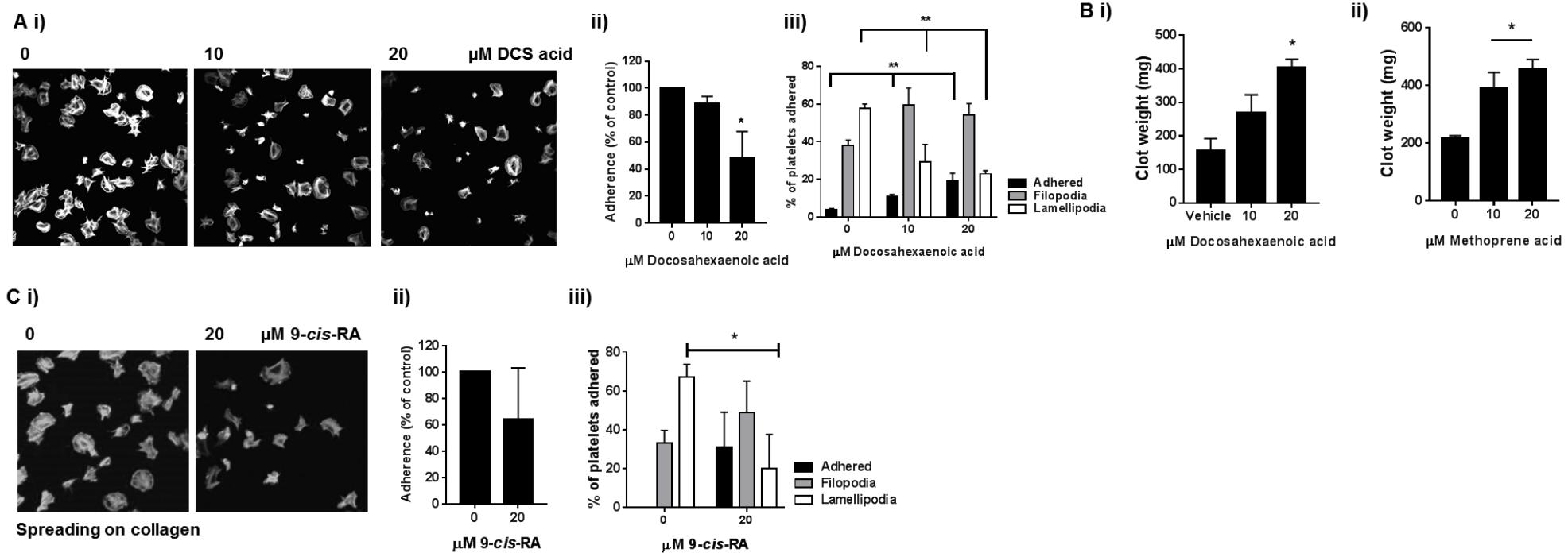
**Supplemental Figure I.** Washed human platelets were A) treated with i) 9-cis-RA, ii) synthetic RXR agonist methoprene and iii) endogenous RXR agonist docosahexaenoic acid and their ability to stimulate platelet aggregation in the absence of platelet agonist monitored using optical light transmission aggregometry, representative traces shown. Human washed platelets were B,C) pre-treated with methoprene acid (10, 20  $\mu$ M) or D,E) pre-treated with docosahexaenoic acid (10, 20  $\mu$ M) for 10 minutes prior to stimulation with either B,D) collagen (1  $\mu$ g/mL) or C,E) thrombin (0.05 U/mL) and aggregation monitored using optical light transmission aggregometry i) Representative traces and ii) quantified data shown. F) pre-treated with 9-cis-RA (20  $\mu$ M) for 10 minutes or vehicle control, in the presence of P $\gamma$ 12 inhibitor, cangrelor (1  $\mu$ M), P2Y1 inhibitor, MRS2179 (100  $\mu$ M) and cyclooxygenase inhibitor, indomethacin (20  $\mu$ M) quantified data shown. G) pre-treated with RXR antagonist HX531 (10, 30  $\mu$ M) for 10 minutes and aggregation to i) collagen (0-10  $\mu$ g/mL), ii) thrombin (0-1 U/mL), iii) U46619 (0-3  $\mu$ M) was monitored using an optical light transmission plate based aggregometry assay, quantified data shown. Data expressed as a percentage of vehicle treated control, results are mean  $\pm$  S.E.M. for  $n \geq 3$ , \* indicates  $p \leq 0.05$  in comparison to vehicle controls.



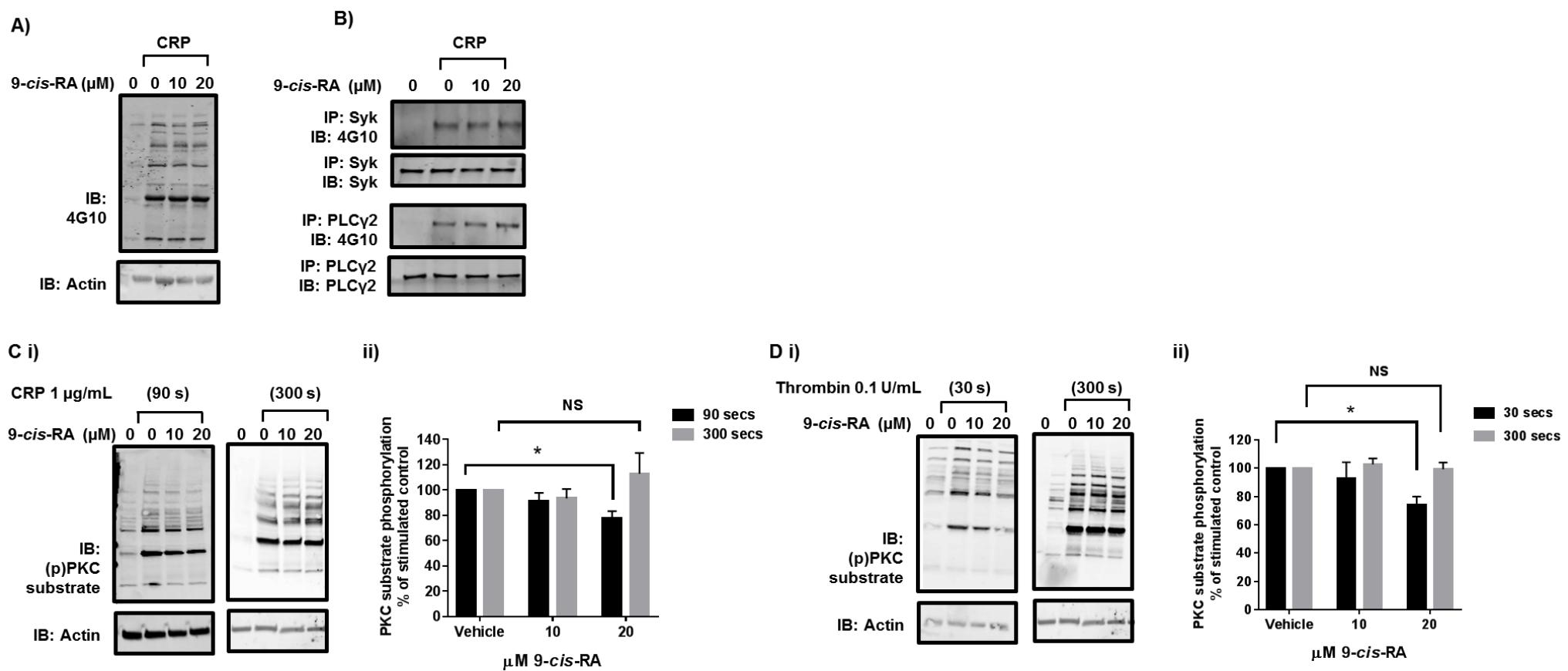
**Supplemental Figure II. The effect of methoprene acid on platelet function.** Human washed platelets were pre-treated for 10 minutes with increasing concentrations of methoprene acid (10, 20  $\mu$ M) before stimulation by CRP-XL (0.25  $\mu$ g/mL) or thrombin (0.05 U/mL). A) Platelet activation measured as fibrinogen binding in i) CRP-XL stimulated and ii) thrombin stimulated platelets. B) Alpha granule secretion as P-selectin exposure in i) CRP-XL and ii) thrombin stimulated platelets. Mobilisation of intracellular calcium determined in Fura-2 AM loaded platelets following stimulation with C) CRP-XL, D) thrombin. i) Representative traces and ii) quantified data shown. Data expressed as a percentage of untreated control. Results are mean + S.E.M. for  $n \geq 3$ , \* indicates  $p \leq 0.05$  in comparison to vehicle controls.



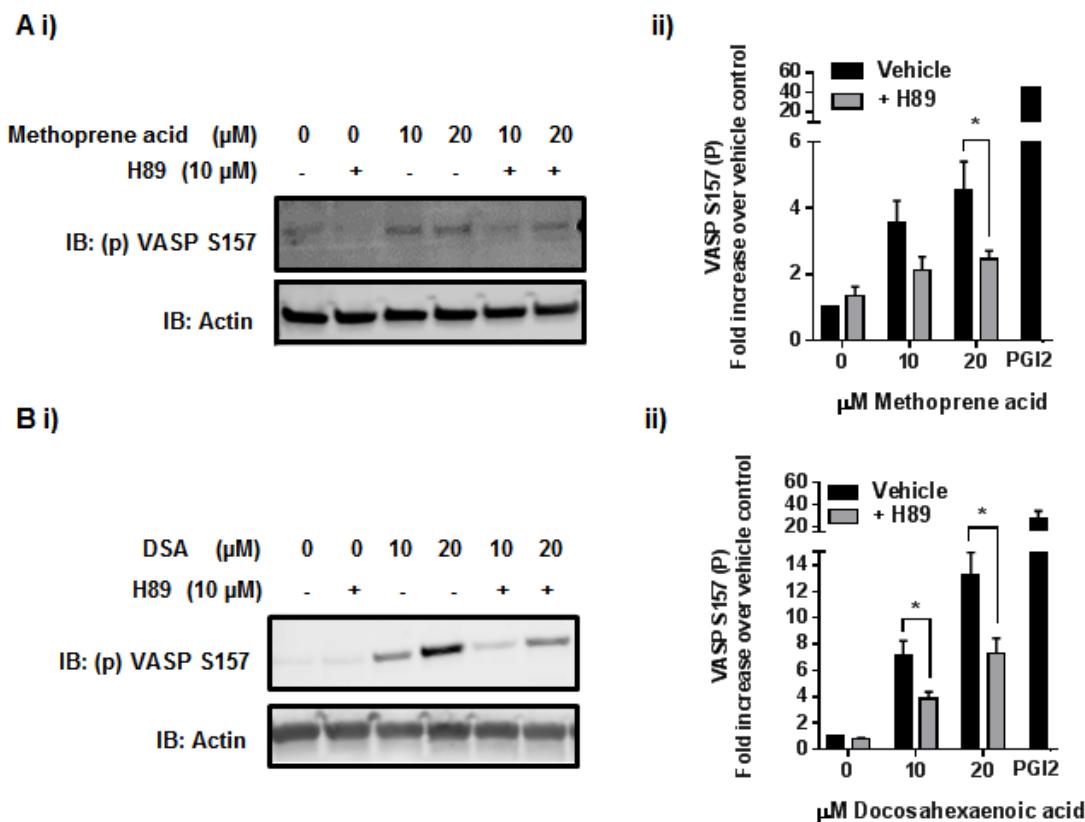
**Supplemental Figure III. The effect of RXR ligands on adhesion and spreading** Human washed platelets pre-treated for 10 minutes with docosahexaenoic acid (20  $\mu$ M) or vehicle control were exposed to fibrinogen (100  $\mu$ g/mL) coated coverslips. B) Clot retraction, human washed platelets pre-treated for 10 minutes with increasing concentrations of i) docosahexaenoic acid (10, 20  $\mu$ M), ii) methoprene acid (10, 20  $\mu$ M) or vehicle control were added to aggregometer tubes in the presence of 2 mg/mL fibrinogen and 2 mM CaCl<sub>2</sub>. Clot retraction was initiated by addition of thrombin 1 U/mL (final concentration) and left to proceed for 1 hour at room temperature. Extent of clot retraction was determined by comparing clot weight. i) Representative images using red blood cell stained PRP, ii) data expressed as clot weight (mg). C) Human washed platelets pre-treated for 10 minutes with 9-cis-RA (20  $\mu$ M) or vehicle control were exposed to collagen (100  $\mu$ g/mL) coated coverslips. A,C i) Representative images of spreading and adhesion after 45 min. Platelets were stained with phalloidin Alexa-488 for visualisation. Images were taken under oil immersion lens with magnification  $\times 100$ . ii) Adhesion, number of platelets adhered were counted in 5 randomly selected fields of view and the number of cells adhered expressed as a percentage of the vehicle treated control. iii) Spreading, platelets were classified into 3 different categories to determine the extent of their spreading (Adhered but not spread, Filopodia: platelets in the process of extending filopodia and Lamellipodia: platelets in the process of extending lamellipodia including those fully spread). Results expressed, as a percentage of the total number of platelets adhered. Unless stated otherwise results are mean  $\pm$  S.E.M. for  $n \geq 3$ , \* indicates  $p \leq 0.05$  in comparison to vehicle controls.



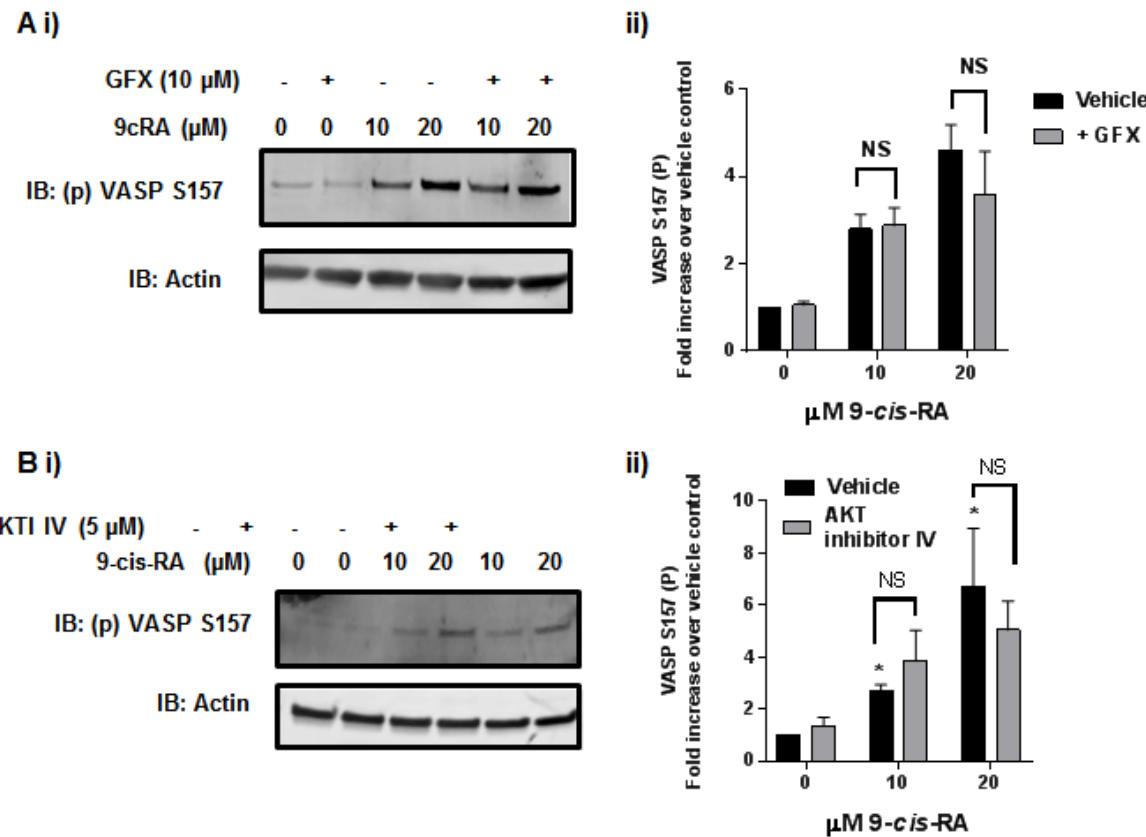
**Supplemental Figure IV. Signalling in 9-cis-RA treated platelets.** 9-cis-RA, (0, 10, 20  $\mu$ M) pre-treated human washed platelet lysates were tested for A) global tyrosine phosphorylation and B) Syk and PLC $\gamma$ 2 phosphorylation in CRP-XL (1  $\mu$ g/mL) stimulated platelets. PKC substrate phosphorylation C) after 90 and 300 secs of CRP-XL (1  $\mu$ g/mL) stimulation and D) 30 and 300 secs of thrombin stimulation (0.1 U/mL). Samples were lysed in Laemmli sample buffer, separated by SDS PAGE and transferred to PVDF membranes before blotting with 4G10 antibody to measure global tyrosine phosphorylation or a phosphosite specific antibody against the PKC substrate recognition sequence. Syk and PLC $\gamma$ 2 were immunoprecipitated from lysates prior to addition of Laemmli sample buffer. Blots were reprobed for total Syk, PLC $\gamma$ 2 or actin to confirm equal loading. A-D) representative blots shown, C-D) i) representative blots shown, ii) Levels of total phosphorylation were quantified and expressed as a percentage of vehicle treated controls. Results are mean  $\pm$  S.E.M. for  $n \geq 3$ , \* indicates  $p \leq 0.05$  in comparison to vehicle controls.



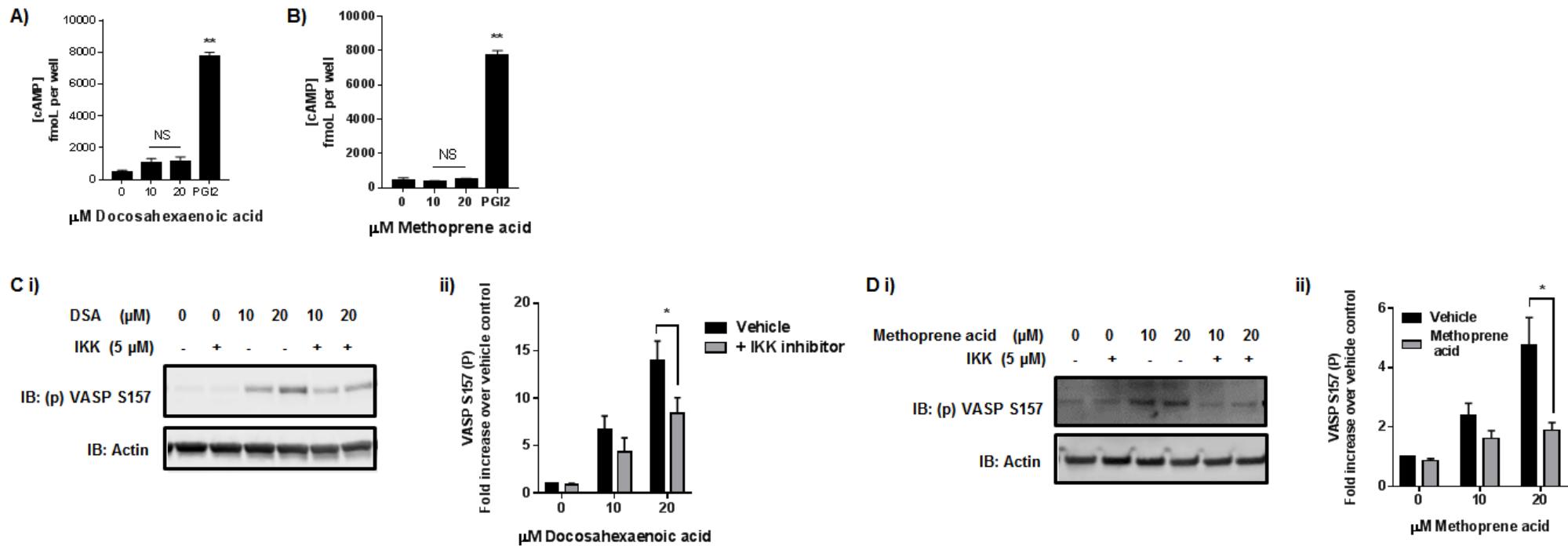
**Supplemental Figure V. RXR ligands and PKA activity.** Resting human washed platelets were treated with A) methoprene acid (10, 20  $\mu$ M) or B) Docosahexaenoic acid (DSA) in the presence and absence of PKA inhibitor H89 (10  $\mu$ M) for 10 minutes and samples tested for VASP S157 phosphorylation, a marker of PKA activity. PGI2 an activator of PKA activity was included as a positive control. Blotting samples were lysed in Laemmli sample buffer before separation by SDS PAGE gels and transfer onto PVDF membranes. Actin was used as a loading control. i) Representative blots are shown and ii) levels of phosphorylation were quantified and expressed as fold increase compared to vehicle control. Results are mean + S.E.M. for  $n \geq 3$ , \* indicates  $p \leq 0.05$  in comparison to vehicle controls.



**Supplemental Figure VI. 9-cis-RA dependent VASP S157 phosphorylation is not dependent on PKC or AKT activity.** Resting human washed platelets were treated with 9-cis-RA (10, 20  $\mu$ M) in the presence and absence of A) a PKC inhibitor GF109203X (GFX) (10  $\mu$ M) B) an AKT inhibitor AKT inhibitor IV (AKTI IV) (5  $\mu$ M) for 10 minutes and samples tested for VASP S157 phosphorylation, a marker of PKA activity. Blotting samples were lysed in Laemmli sample buffer before separation by SDS PAGE gels and transfer onto PVDF membranes. Actin was used as a loading control. i) Representative blots are shown and ii) levels of phosphorylation were quantified and expressed as fold increase compared to vehicle control. Results are mean + S.E.M. for  $n \geq 3$ , \* indicates  $p \leq 0.05$  in comparison to vehicle controls.

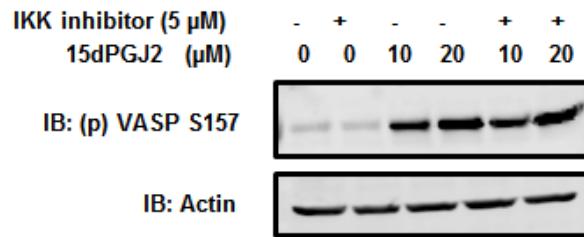


**Supplemental Figure VII. RXR ligand dependent activation of PKA.** Resting human washed platelets were treated with A,C) docosahexaenoic acid (10, 20  $\mu$ M) or B,D) methoprene acid (10, 20  $\mu$ M) for 10 minutes and in the presence and absence of A,B) phosphodiesterase inhibitor IBMX (1 mM) and cAMP levels measured using a cAMP ELISA kit (GE Healthcare as per manufacturers instructions). And C,D) an NF $\kappa$ B inhibitor, IKK inhibitor VII (5  $\mu$ M) and samples tested for VASP S157 phosphorylation, a marker of PKA activity. Blotting samples were lysed in Laemmli sample buffer before separation by SDS PAGE gels and transfer onto PVDF membranes. Actin was used as a loading control. i) Representative blots are shown and ii) levels of phosphorylation were quantified and expressed as fold increase compared to vehicle control. Results are mean  $\pm$  S.E.M. for  $n \geq 3$ , \* indicates  $p \leq 0.05$  in comparison to vehicle controls.

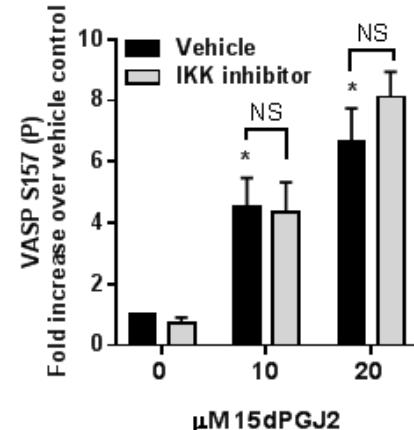


**Supplemental Figure VIII. RXR ligand dependent activation of PKA via NF $\kappa$ B is not dependent on PPARs.** Resting human washed platelets were treated with A) 15dPGJ2 (10, 20  $\mu$ M) a PPAR $\gamma$  ligand or B) LG101506 (10, 20  $\mu$ M) a RXR:PPAR heterodimer modulator, for 10 minutes in the presence and absence of an NF $\kappa$ B inhibitor, IKK inhibitor VII (5  $\mu$ M) and samples tested for VASP S157 phosphorylation, a marker of PKA activity. Blotting samples were lysed in Laemmli sample buffer before separation by SDS PAGE gels and transfer onto PVDF membranes. Actin was used as a loading control. i) Representative blots are shown and ii) levels of phosphorylation were quantified and expressed as fold increase compared to vehicle control. Results are mean + S.E.M. for  $n \geq 3$ , \* indicates  $p \leq 0.05$  in comparison to vehicle controls.

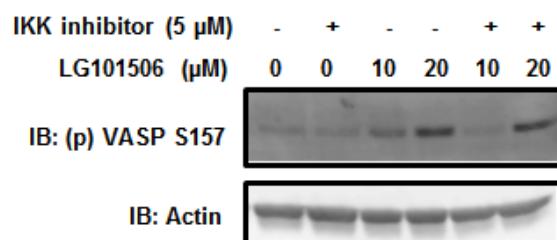
A i)



ii)



B i)



ii)

