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Membrane-initiated non-genomic signaling by estrogens in the hypothalamus: cross-talk with glucocorticoids with implications for behavior

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The estrogen receptor and glucocorticoid receptor are members of the nuclear receptor superfamily that can signal using both non-genomic and genomic transcriptional modes. Though genomic modes of signaling have been well characterized and several behaviors attributed to this signaling mechanism, the physiological significance of non-genomic modes of signaling has not been well understood. This has partly been due to the controversy regarding the identity of the membrane ER (mER) or membrane GR (mGR) that may mediate rapid, non-genomic signaling and the downstream signaling cascades that may result as a consequence of steroid ligands binding the mER or the mGR. Both estrogens and glucocorticoids exert a number of actions on the hypothalamus, including feedback. This review focuses on the various candidates for the mER or mGR in the hypothalamus and the contribution of non-genomic signaling to classical hypothalamically driven behaviors and changes in neuronal morphology. It also attempts to categorize some of the possible functions of non-genomic signaling at both the cellular level and at the organismal level that are relevant for behavior, including some behaviors that are regulated by both estrogens and glucocorticoids in a potentially synergistic manner. Lastly, it attempts to show that steroid signaling via non-genomic modes may provide the organism with rapid behavioral responses to stimuli.

Keywords: hypothalamus, spine density, membrane-initiated signaling, GPCR, estrogen receptor variants, aggression, lordosis, glucocorticoid receptor

GENOMIC AND NON-GENOMIC SIGNALING BY NUCLEAR RECEPTORS

Nuclear receptor ligands such as estrogen and glucocorticoids signal via both non-genomic and genomic pathways within cells. The genomic or transcriptional pathway is the best elucidated primarily due to the well-characterized nature of the estrogen receptor (ER) α and β and the glucocorticoid receptor (GR), all of which are members of the nuclear receptor superfamily. Once bound to their cognate ligands, these receptors act as ligand-activated transcription factors in the nucleus by binding to specific enhancer elements such as the estrogen response element (ERE) (1) and glucocorticoid response element (GRE) (2) in the promoters of genes. Both receptors have a modular structure, with a conserved DNA-binding domain, multiple transactivation domains, and a C-terminal ligand-binding domain (3, 4).

On the other hand, non-genomic signaling, first described by Szego and Davis in 1967, as the rapid increase in cAMP in the uterus occurred within 15 min of 17 β -estradiol (17 β -E) administration to ovariectomized mice (5). In the central nervous system (CNS), 17 β -E was shown to rapidly depolarize pro-opiomelanocortin (POMC) hypothalamic neurons via Akt or protein kinase (PK) B, extracellular regulated kinase (ERK/MAPK), PKA, and PKC pathways (6, 7). In other tissues such as rat hippocampal neurons, phospho-cAMP response element binding

protein (pCREB) increased within 1 h of 17 β -E addition and this increase was blocked by inhibitors to both calmodulin kinase II (CamKII) and ERK pathways (8). In the case of corticosterone-mediated rapid actions, treatment of neurons with dexamethasone, a synthetic glucocorticoid, rapidly induced the nuclear localization of the GR (9, 10), an effect potentiated by the inhibition of p38MAPK (11). Extracts from rat hippocampal synaptoneuroosomes showed a reduction in Akt and ERK phosphorylation within 30 min in response to pharmacological inhibition of the GR by RU-486 (12), suggesting that the classical nuclear receptor was required for non-genomic signaling in the hippocampus. Apart from kinase activation, dexamethasone-mediated negative feedback at the corticotropin releasing hormone (CRH) neuron was also rapid, consisting of suppression of the excitatory drive to the CRH neuron, mediated by endocannabinoids acting as a retrograde messenger to the presynaptic glutamatergic neuron (13), an effect mimicked with a membrane-limited dexamethasone conjugated to bovine serum albumin (Dex-BSA) (13). Hence, non-genomic signaling by steroid hormones is extra-nuclear signaling that is initiated by the endogenous ligand within minutes, in contrast to the hours required to detect transcriptionally regulated proteins.

Central to this concept of non-genomic signaling that is typically demonstrated by the use of membrane-limited conjugates

(14), is the idea of a receptor that initiates such signaling from the plasma membrane. However, with the exception of the membrane progesterone receptors (mPRs) that belong to the progestin and adipoQ receptor (PAQR) family, the identity of the membrane ER (mER) and membrane GR (mGR) has remained elusive (15). This review aims to describe the current candidates for the mER and the mGR that mediate rapid non-genomic signaling from the plasma cell membrane as well as focus on rapid actions that are relevant for hypothalamically driven behaviors that are dependent on estrogens but that have a glucocorticoid-regulated component. We concentrate on the hypothalamus since this is a classically steroid-responsive area of the brain and is critical for several estrogen-dependent behaviors (16). For more general reviews on rapid actions of estrogens and glucocorticoids in the CNS, including tools that are typically used to elucidate membrane-initiated non-genomic effects, the reader is referred to (7, 14, 17–19).

THE MEMBRANE ER AND GR

In the CNS, both ^3H -labeled $17\beta\text{-E}$ (20) and a ^{125}I -labeled membrane-impermeant conjugate where $17\beta\text{-E}$ is attached to bovine serum albumin (E2-BSA) (21) showed relatively high affinity binding to rat plasma membranes, suggesting the presence of a mER. What is a good definition for a mER or mGR? Previously, Micevych et al. (22) have suggested that ICI 182,780 antagonism, stereospecific $17\beta\text{-E}$ binding and sequence homology to the ER α and ER β should be considered pre-requisites for a protein to be termed a mER. We propose, given the off-target effects of ICI 182,780 (23) that the definition of the mER or mGR be modified slightly to consider proteins capable of specific binding to $17\beta\text{-E}$ or to dexamethasone (in the case of the mGR), presence at/near the plasma membrane and signaling from the membrane. It should be noted that this definition would exclude the binding of proteins to $17\alpha\text{-E}$, an estrogen often used in studies of as an inactive isomer of $17\beta\text{-estradiol}$, but that recently has been shown to increase neurogenesis and to mediate neuroprotection (24). Most studies described below show evidence of the candidate mER at the plasma membrane and its ability to rapidly within minutes modulate rapid signaling pathways such as kinase regulation or calcium flux. A few studies also demonstrate the regulation of the candidate ER at the membrane.

ER α AS THE mER

As we will focus on outputs dependent on the hypothalamus in this review, we will initially discuss the nuclear classical ER α as a candidate mER because (a) this is the most abundant ER isoform in the medial preoptic area (mPOA), ventromedial hypothalamus (VMH), and arcuate nucleus (ARH) (25–27) and (b) loss of ER α in the hypothalamus abrogated hypothalamically driven lordosis behavior in females (28) and aggressive behavior in males (29). A number of studies using $17\beta\text{-E}$ binding, electron microscopy, immunocytochemistry, and western blotting have examined the idea that a small percentage (3–5%) of the total pool of the classical ER α is present on the plasma membrane (30). Consistent with this idea, immunocytochemistry using minimal fixation in both the breast cancer MCF-7 cell line (31) and anterior pituitary GH3 cell line (32), revealed ER α at the membrane in caveolae in some, but not all cells. MCF-7 cells with ER α at the plasma membrane

showed increased phospho-ERK (pERK) within 10 min of application of either $17\beta\text{-E}$ or a membrane-limited E2-peroxidase conjugate (33). In the CNS, ER α was localized to the dendrites and axon terminals in the guinea pig hypothalamus (34), while ER α was detected in axon terminals, dendritic spines, as well as in astrocytes in the CA1 using immuno-electron microscopy in the proestrous female rat (35). In the dorsal CA1 from the female rat, ER α was present in synaptic vesicles in the axon of some GABAergic basket cells; $17\beta\text{-E}$ moved these ER α -containing vesicles toward synapses within 24 h (36). In addition, some proportion of this extra-nuclear ER α at synaptosomes and vesicles was phosphorylated though the function of phosphorylation in localization of the ER α remains unknown (37). In hypothalamic neurons and astrocytes obtained from both male and female rats, a full-length 66 kDa form of the ER α and a 52 kDa variant has been detected using surface biotinylation and immunocytochemistry (38, 39). These studies demonstrate that the nuclear ER α is present at the plasma membrane in many cell types, including the hypothalamus.

TARGETING OF ER α TO THE PLASMA MEMBRANE

Though hydropathicity analysis shows that ER α may have a potential trans-membrane domain (38), the idea that ER α requires a membrane protein to tether it to the plasma membrane has been cemented primarily by the discovery of caveolin proteins that can anchor the ER α in lipid rafts and caveolae. Caveolin proteins are highly conserved structural proteins that are necessary and sufficient for the existence of caveolae, a subset of lipid rafts (40) that comprise a restricted compartment for signaling molecules associated with the plasma membrane. Initially, ER α was reported to interact with caveolin 1 (Cav-1) in MCF-7 and vascular smooth muscle cells (VSMC) (41). The caveolins in turn act as adaptor proteins to couple ER α selectively to either other proteins such as the metabotropic glutamate receptors (mGluR) or to processes such as palmitoylation. In the hippocampus of the female rat, Cav-1 interacted with mGluR1a so that $17\beta\text{-E}$ activated downstream G α_q and ERK signaling. This, in turn, increased pCREB activation within 5 min of $17\beta\text{-E}$ application (42). Hence, inhibitors to phospholipase C (PLC) and ERK and mGluR1a as well as a dominant negative Cav-1 mutant decreased pCREB activation (42). On the other hand, Cav-3 tethered ER α in hippocampal neurons to mGluR2/3; this pathway activated G α_i signaling, leading to the inhibition of PKA and the subsequent downregulation of pCREB (42). Hence, the type of caveolin dictated the final divergent pCREB response though the physiological conditions wherein ER α may bind to Cav-1 versus Cav-3 are unknown. In hypothalamic astrocytes, the coupling of ER α to mGluR1a resulted in an increase of calcium within 2 min of $17\beta\text{-E}$ application; this was blocked both by the ER α and ER β antagonist, ICI 182,780 and by the mGluR1a antagonist LY 367385 (43). In striatal neurons, Cav-1 facilitated the tethering of ER α to mGluR5 and subsequent G α_q signaling (44). Hence, caveolins may not only tether ERs into caveolae in the plasma membrane in different cell types but also subserve tissue-specific signaling to specific G α subunits by co-opting different mGluR partners. The coupling of ER α to mGluRs may allow downstream signaling to be potentiated by rendering it sensitive to two ligands, i.e., $17\beta\text{-E}$ for the ER and glutamate for the mGluR. This is supported by the data that in hypothalamic

astrocytes, glutamate, and 17β -E combined elicited greater Ca^{+2} increases than either ligand alone (43).

In addition to the role that mGluRs play in linking $\text{ER}\alpha$ to $\text{G}\alpha$ proteins, $\text{ER}\alpha$ can also directly bind $\text{G}\alpha$ subunits. 17β -E inhibited cAMP production within 5 min of addition to GT1-7 immortalized GnRH neurons via $\text{ER}\alpha$ that is physically tethered to $\text{G}\alpha_{i3}$ (45) protein that can be detected in co-immunoprecipitation experiments. Interestingly, addition of 17β -E decreased the amount of membrane-associated $\text{G}\alpha_{i3}$ and this decrease was blocked by the ER antagonist ICI 182,780 suggesting control of the signaling cascade by the ligand, similar to classical G-protein coupled receptors (GPCRs) (45). Though $\text{ER}\alpha$ has been demonstrated to tether to $\text{G}\alpha_i$ in the caveolae of endothelial cells to increase the activity of nitric oxide synthase (46), this mode of regulation of nitric oxide synthase by 17β -E has not been shown in the CNS.

Apart from serving as adaptors, the binding of caveolins to the ER also increases palmitoylation of the ER, a process by which palmitate, a C16 fatty acid is added to an internal cysteine via a thioester bond. In the human $\text{ER}\alpha$, a canonical palmitoylation site exists at the cysteine amino acid residue at position 447 (C447); the corresponding site in the mouse $\text{ER}\alpha$ is the C451 (31). Mutant $\text{ER}\alpha$ where the C447 site was mutated to alanine (C447A) did not bind Cav-1, was not localized at the membrane in HeLa cells and showed decreased pERK activation in response to a 10-min application of 17β -E (47). Additionally, when a non-classical palmitoylation site, S522, in the $\text{ER}\alpha$ was mutated, localization at the plasma membrane and Cav-1 binding to $\text{ER}\alpha$ was decreased (48) in Chinese hamster ovary (CHO) cells. This mutant also decreased the ability of 17β -E to increase pCREB in hippocampal neurons from the female rat (42). Other binding partners for the $\text{ER}\alpha$ also lead to increases in palmitoylation and subsequent membrane localization. For example, the amino acid residue cysteine at position 451 of the mouse $\text{ER}\alpha$ also binds the heat-shock protein 27 (hsp27),

which increased the rate of palmitoylation at this site. Therefore, siRNA to hsp27 decreased palmitoylation with subsequent decrease of $\text{ER}\alpha$ at the plasma membrane and decreased phospho-Akt (pAkt) normally induced by a 10 min application of 17β -E (49). The C-terminal of mouse $\text{ER}\alpha$ (Leu512–514) is important in $\text{ER}\alpha$ dimerization at the plasma membrane; mutation of these residues decreased pAkt and cAMP generation within 5 min of 17β -E addition and decreased the activation of both $\text{G}\alpha_s$ and $\text{G}\alpha_q$ subunits in the CHO cell line (50) (**Figure 1**). In the CNS, it is unknown if the $\text{ER}\alpha$ at the membrane exists as dimers and if these dimers use caveolin proteins as adaptors. It is also unclear if caveolin binding to the $\text{ER}\alpha$ precedes greater palmitoylation since $\text{ER}\alpha$ mutations that destroy the interaction with caveolin also destroy palmitoylation. Recently, E2-BSA has been shown to transcriptionally upregulate Cav-1 via a PI3K and ERK pathway within 12 h in endothelial cells (51). Hence, 17β -E may increase non-genomic signaling both by increasing the palmitoylation of $\text{ER}\alpha$ within rapid time frames and within longer time frames via the transcriptional upregulation of Cav-1.

REGULATION OF $\text{ER}\alpha$ AT THE MEMBRANE BY 17β -E

Similar to the tissue specificity shown by $\text{ER}\alpha$ -mediated rapid, non-genomic signaling, $\text{ER}\alpha$ at the membrane is also regulated by 17β -E in both temporal and tissue-specific manner. In hypothalamic astrocytes (38) and neurons (39) and in a hypothalamic cell line, mHypoE-38 (52), 17β -E increased the amount of both the 52 and 66 kDa mER in the cell membrane within 30 min. This increase of mER at the plasma membrane can be blocked by ICI 182,780 and the mGluR1a antagonist LY 367385 but does not occur with E2-BSA, suggesting that this is an internal, non-membrane-initiated rapid effect that is dependent on the mGluR1a receptor. In contrast, in MCF-7 cells, 17β -E decreased Cav-1 within 8 h, thus decreasing the amount of $\text{ER}\alpha$ at the membrane. Similarly, 17β -E

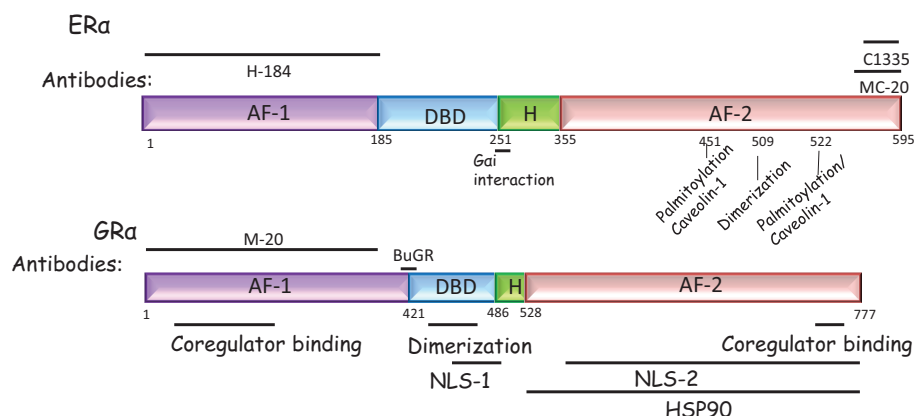


FIGURE 1 | Domain structure of the $\text{ER}\alpha$ and the $\text{GR}\alpha$. $\text{ER}\alpha$ and $\text{GR}\alpha$ are classical intracellular nuclear receptors but may also be present on the plasma membrane. Both receptors have a N-terminal AF-1 domain, a central DNA binding domain, a hinge domain (H), and a C-terminal AF-2 domain, required to bind ligand. $\text{ER}\alpha$: the antibodies most commonly used to detect $\text{ER}\alpha$ are the H184 (Santa Cruz Biotechnology, TX, USA) antibody raised to the N-terminal domain and the MC-20 (Santa Cruz Biotechnology, TX, USA) and C1335 (Upstate Biotechnology, NY, USA) raised to the C-terminal domain. In addition,

the sites that may tether the $\text{ER}\alpha$ to the plasma membrane are marked. These are residues required for caveolin or $\text{G}\alpha$ binding or residues that are palmitoylated or required for dimerization at the membrane. $\text{GR}\alpha$: the antibodies most commonly used to detect $\text{GR}\alpha$ are the BuGR antibody (Abcam Inc., MA, USA) and the M-20 (Santa Cruz Biotechnology, TX, USA) antibody, directed to the N-terminal domain. The NLS refers to the nuclear localization signal since nuclear localization of this receptor can occur rapidly and is a non-genomic effect.

treatment of hippocampal slices from female rats removed ER α from the synaptosomal plasma membrane by depalmitoylation within 20 min (37). In CHO cells, palmitate incorporation into ER α and association with Cav-1 decreased within 60 min of 17 β -E administration, suggesting that ER α at the plasma membrane was lowered, consistent with this idea, ERK activation also decreased (47). Decreasing ER α at the membrane in these cells may allow for a decrease in non-genomic signaling and may mark a shift toward genomic transcription. In contrast, in the VSMC, 17 β -E increased both Cav-1 and Cav-2 and increased ER α localization at the plasma membrane (41). Such increased ER α localization in response to the endogenous ligand may be a mechanism to sustain non-genomic signaling in some cell types.

Similar to other trans-membrane GPCRs, which may undergo desensitization via internalization in a G-protein receptor kinase 2 (GRK2)- β -arrestin-dependent manner (18), 17 β -E application to cortical neuronal cultures rapidly induced GRK2 phosphorylation within 5 min and β -arrestin1 and c-Src recruitment to the ER α within 60 min. Predictably, since β -arrestin1 tagging of trans-membrane GPCRs provides a scaffold for c-Src attachment and subsequent ERK activation (53), siRNA to β -arrestin1 abrogated the 17 β -E induction of pERK in these neurons (54). Though the exact partners of the ER α in this internalized signalosome are unknown, ER α internalization is initiated from the membrane since E2-BSA also promotes internalization (54). Internalization of the ER α has also been tested more directly by using a surface biotinylation technique followed by stripping the surface biotin with glutathione after 17 β -E addition to cells. Hence, the continued presence of biotin in this preparation would argue that the receptor was internalized and protected against glutathione action. In hypothalamic neurons, addition of 17 β -E caused internalization of both the 52 and 66 kDa forms within 30 min, with internalization of the 52 kDa form persisting for 2 h (54), demonstrating that recycling and internalization of the classical ER α as well as the 52 kDa variant occurs in a rapid time frame. An unresolved issue is the rapid decrease in total ER α protein and mRNA in the hypothalamus on 17 β -E application that has been shown by several investigators (55, 56). However, it is difficult to reconcile this with the increase in plasma mER α that is seen in the hypothalamic primary neuronal and astrocytic cultures (38, 39) and in the hypothalamic cell line mHypoE-38 (52) unless ER α from other non-membrane locations decrease to a large extent; this could be a parameter to be investigated in future studies. This could also be due to the recent studies that show an increase ER α at the plasma membrane being carried out in primary cultures versus hypothalamic tissue that retain neuronal connectivity in the older studies.

DEPENDENCE OF RAPID SIGNALING ON ER α AND ER β

In addition to direct demonstration of the ER α either at the membrane or tethering to an intrinsic membrane protein, the ER dependence of kinase activation has also been investigated. Several studies, described in this review, have used the ER α and ER β antagonist ICI 182,780 to antagonize a rapid signaling pathway or a genetic approach that deletes either ER α or ER β specifically in the whole animal. The availability of ER α and ER β knockout animals, i.e., ER α KO and ER β KO, has been useful to understand the

ER dependence of 17 β -E-mediated non-genomic signaling *in vivo*, particularly in light of an absence of a specific antibody to the ER β (57). In wild-type female mice, subcutaneous administration of 17 β -E caused the activation of CREB and ERK within an hour, as measured by immunohistochemistry for the phosphorylated forms of these molecules. When either ER α or ER β or both were deleted, a complex brain nuclei-specific and isoform-specific ER dependence was revealed, which may in part be due to the tissue distribution of the ER isoforms in the brain (Table 1 below) (58). For example, in the ventrolateral VMH (vl-VMH), the loss of ER α results in the reduction of pCREB on 17 β -E administration and this could be because the predominant ER isoform is the ER α and it is possible that the loss of ER α cannot be compensated by smaller amounts of ER β (58). The use of ER α KO, ER β KO, and ER α β KO mice in this study showed that ER α is hence both necessary and sufficient for the induction of pCREB. In the mPOA, 17 β -E could induce pCREB via either ER α or ER β but required both ER α and ER β to act in conjunction in order to induce pERK (58), demonstrating that ER isoforms can regulate rapid signaling differentially in the same tissue. The regulation in male mice is unknown and is under investigation in our laboratory.

ER α VARIANTS

In both hypothalamic neurons and astrocytes (38), a variant 52 kDa ER α (ER α -52), detected with a C-terminal antibody that also detects the full-length 66 kDa, was expressed in much higher amounts than the ER α -66 kDa form. Based on RT-PCR analysis using flanking primers to this exon in the mouse hypothalamic cell line mHypoE-38, this is believed to be an ER α that lacks exon 4 (ER α Δ 4) (52). In astrocytes, only the 66 kDa form is thought to be interact with mGluR1 (43) and the interaction, signaling properties, and function, if unique, of the 52 kDa variant remain unknown. A 36 kDa variant lacking the AF-1 and AF-2 domains with a unique C-terminus was detected in human breast

Table 1 | Comparison of CREB and ERK between wild-type (WT), ER α knockout (ER α KO), ER β knockout (ER β KO), and double ER α ER β knockout (ER α β KO) female mouse brain.

Genotype	vlVMH		mPOA		Medial septum	
	pCREB	pERK	pCREB	pERK	pCREB	pERK
WT	++	NR	++	++	++	NR
ER α KO	–	NR	++	–	++	NR
ER β KO	++	NR	++	–	–	NR
ER α β KO	–	NR	–	–	–	NR

pCREB and pERK in 17 β -E injected mice was quantitated by immunohistochemistry and compared to the levels of pCREB and pERK present in mice injected with the vehicle, in the ventrolateral VMH (vl-VMH), the medial preoptic area (mPOA) and the medial septum in female ovariectomized mice injected with 10 μ g 17 β -E and sacrificed an hour after injection. In WT mice, 17 β -E increased pCREB in the vl-VMH and pERK and pCREB in the mPOA. ER isoform dependence varies according to the output (pCREB or pERK) measured and the nuclei. ++: upregulation compared to vehicle-treated mice. –: no upregulation compared to vehicle-treated mice. NR = data not recorded. Data are from Ref. (58).

cancer cells, coded for by a unique promoter located in intron 1 of the *Esr1* gene (59) and can be detected by a specific antibody. ER α -36 was localized to the plasma membrane, mediated rapid ERK signaling, and is induced by the selective activation of the GPR30 (discussed in Section “GPR30”) by the agonist, G-1 (60). Importantly, this GPR30-selective activator, G-1, has also been demonstrated to bind and activate ER α -36 (60), suggesting that some of the responses attributed to the GPR30 might be via ER α -36. A 46 kDa variant observed in endothelial cells is also localized to the plasma membrane (61) and is composed of a splice variant missing the first 173 amino acids of ER α -66 (62). ER α -46 cannot be detected with an N-terminus antibody raised to ER α -66 such as H-184 (Figure 1), but should be identifiable with a C-terminus ER α antibody. A novel ER is the ER-X, an ER of 63 kDa that is expressed in the neocortex in the caveolae membrane fraction, can be identified with ER α and ER β antibodies and is present in the ER α knockout animal (63). This ER, whose structure remains unidentified, mediated ERK signaling by both 17 α -estradiol and 17 β -estradiol and would not meet the criteria for a mER, based on our previous definition. The function and expression pattern of the ER-X is unknown though it was upregulated after ischemic injury (63) in the adult. If it is not present in the adult hypothalamus, it is unlikely to be a receptor that mediates normal behavior that is dependent on the hypothalamus (63). So far, neither ER α -36 nor ER α -46 has been demonstrated in the CNS, though it is possible that some antibodies directed toward the DNA-binding domain or, in the case of the 46 kDa variant, the C-terminal domain of the full-length 66 kDa ER α such as MC-20 or C1335 (Figure 1) may detect these forms in studies that use solely immunocytochemistry.

G α -COUPLED mERs

The idea that the mER is a G α_q -coupled receptor has been explored in the POMC neuron in the ARH, primarily by the laboratories of Kelly and Ronneklev (7, 64, 65). In POMC neurons, 17 β -E and E2-BSA uncoupled the inhibitory μ -opioid receptor from the G-protein coupled inward rectifying K $^+$ (GIRK) channel, in a PLC-, PKC-, and PKA-dependent manner (66, 67), thus decreasing hyperpolarization of these neurons and increasing neuronal excitability. Pharmacological characterization revealed that a G α_q receptor upregulated PLC and calcium release, which in turn activated PKC and PKA. Subsequent PKA phosphorylation of the GIRK channel uncoupled the μ -opioid receptor (7, 68) from GIRK. Though this response was specific for 17 β -E versus 17 α -E and was blocked by the ER α and ER β antagonist ICI 162,480, it also occurred in ER α KO, ER β KO, and GPR30 KO mice (69), suggesting that a novel mER is coupled to G α_q . This signaling pathway can be activated by a selective ligand, STX, which does not bind ER α , ER β , or GPR30. Both STX and 17 β -E also reduced post-ovariectomy weight gain, suggesting that this novel G α_q -coupled mER is important in energy homeostasis. Leptin receptors on POMC neurons caused depolarization rapidly via the JAK/STAT and PI3K pathways that open calcium-dependent TRPC channels (70); 17 β -E signaling may synergize with leptin action by increasing calcium rapidly in these neurons via this G α_q -mER. Several reviews (7, 68, 69, 71–74) detail the studies on this STX-activated receptor.

GPR30

The GPR30, also known as GPER1, is a former orphan GPCR that was shown to bind 17 β -E and increase cAMP in breast cancer SKBR3 cells via an increase in adenylyl cyclase activity (75). In the rat hypothalamus, GPR30 expression is particularly high in the PVN and SON (76) with low expression in the VMH. Cell fractionation and immunocytochemistry revealed GPR30 to be localized at the plasma membrane in both the SKBR3 (GPR30 $^+$, ER α $^-$, ER β $^-$) and MDA-MB-231 (GPR30 $^-$, ER α $^-$, ER β $^+$; GPR30 overexpression) breast cancer cell lines (77). Though GPR30 was not detected at the plasma membrane using surface biotinylation in hypothalamic astrocytes (38), GPR30 was localized to the post-synaptic density in the hippocampus and associates with PSD-95 through its C-terminal tail (78). This association with PSD-95 can localize GPR30 to the plasma membrane, independent of 17 β -E. Immuno-electron microscopy analysis of rat hippocampi also revealed membrane localization with no intracellular staining (79). However, it can be detected intracellularly in normal mammary gland epithelial cells (80) and colocalized with a marker of the Golgi apparatus in primary cultures of the hippocampus (81). Also, overexpressed GPR30 in COS7 cells exhibits localization at the endoplasmic reticulum and the Golgi (82), suggesting that cell type can determine localization. In addition, HeLa cells transfected with FLAG-GPR30 show mostly staining at the membrane while cells transfected with GFP-GPR30 show staining mainly in the endoplasmic reticulum [(79) #6908], suggesting that inclusion of molecular tags may interfere with proper intracellular trafficking, possibly confounding the interpretation of experiments relying on ectopic expression of GPR30.

Studies describing the regulation of internalization of GPR30 have also yielded conflicting results. When FLAG-GPR30 was ectopically expressed in HeLa cells, receptor endocytosis was ligand dependent and internalized GPR30 occupied a diffuse cytoplasmic localization consistent with classical receptor recycling or proteasomal degradation pathways (79). In human embryonic kidney (HEK)-293 cells, GPR30 internalization was not only ligand-dependent but also occurs constitutively (83). While most GPCRs are either recycled to the plasma membrane via endosomes or degraded in lysosomes to limit excessive signaling, GPR30 rapidly accumulates in the perinuclear compartment via clathrin-coated vesicles (83). It is thought that this Rab 11-dependent accumulation is in the trans-Golgi network and degradation occurs via an ubiquitin-proteasome-mediated pathway, unlike the endosomal degradation or recycling to the surface that is typical for other GPCRs (83). Since this pathway for internalization and degradation is rapid and constitutive, Cheng et al. propose this as the reason for the difficulty in detecting GPR30 at the plasma membrane in some cell lines (83).

GPR30 can signal via both the G α_s and the G $\beta\gamma$ subunits that are associated with the receptor upon activation. The G $\beta\gamma$ subunit transactivated the EGFR receptor leading to downstream activation ERK protein, while simultaneous activation of the G α_s subunit by 17 β -E inactivated ERK signaling through activation of adenylyl cyclase and PKA, thus limiting cAMP signaling to a short time frame (23). Furthermore, although GPR30 protein was readily detectable in both the microsomal and plasma membrane subcellular fractions of breast cancer cell lines, only GPR30

in the plasma membrane fraction bound ligand and activated G-protein signaling, suggesting that only membrane-associated GPR30 protein is functional (77). However, GPR30 activation in COS7 cells initiated intracellular calcium mobilization and nuclear accumulation of PIP3 via EGFR transactivation (82) via a non-membrane-initiated signaling mechanism since calcium flux was not replicated by membrane restricted estradiol derivatives (84). These discrepant results have yet to be reconciled. A recent study reported that GPR30 could decrease cAMP that was elevated by heterologous ligands, such as forskolin, via the C-terminal PDZ domain (85). This domain could bind membrane-activated guanylate cyclases (MAGUKs), which act as adaptors for AKAP5 (PKA anchoring protein) that in turn decreased adenylate cyclase activity. This would decrease the cAMP that is elevated by 17 β -E binding of the GPR30 and may serve to limit the time frame of GPR30 signaling (85). Hence, both ERK and cAMP signaling downstream of the GPR30 may be restricted to very short time frames.

THE iGR AS THE mGR

Similar to the ER α being considered a possible mER, the intracellular GR (iGR) that exists as a complex with heat-shock protein 90 (HSP90), p23, and a tetratricopeptide protein (86) in the cytoplasm has been proposed as a candidate mGR (87). Stable reduction of mGR levels in CD14⁺ monocytes using stably transfected siRNA to iGR α suggested that both mGR and iGR were derived from the same transcript (88). GR exists in two isoforms, GR α and GR β . GR β , generated by alternative splicing, lacks the last 50 amino acids of the GR α carboxy terminus, and instead possesses a unique 15 amino acid sequence at its C-terminus (89). GR β neither binds glucocorticoids nor has intrinsic transcriptional activity, but has been implicated as a dominant negative inhibitor of GR α activity through formation of a non-functional heterodimer (90). Higher concentrations of GR β relative to GR α , therefore, result in decreased glucocorticoid sensitivity. However, levels of GR β protein in the brain are extremely low (89) and this is not thought to be a regulator of GR α in the brain. The presence of mGR α was detected at very low levels in human lymphocytes and leukocytes using membrane-impermeable fluorescent liposomes and GR-specific antibodies (91) such as M-20 (**Figure 1**). Similar to the tethering of ER α to the plasma membrane, mGR α has been associated with Cav-1 in MCF-7 cells (92). However, while plasma membrane association of other steroid hormone receptors, including ER, is dependent on palmitoylation, mutation of the homologous sequence in GR did not affect membrane localization (93), suggesting that other mechanisms must tether mGR to the plasma membrane. In the brain, most of the rapid effects of glucocorticoids have been confined to the hippocampus and to the hypothalamus. Membrane glucocorticoid receptors were first observed in the synaptic plasma membrane fractions (SPM) of rat brain via [3H]-corticosterone binding assays (20). Hypothalamic SPM had a higher binding capacity than hippocampal or cortical SPM, which is in contrast to corticosterone binding of the iGR, which is lower in the hypothalamus and much higher in cerebral cortex and hippocampus (94). Synaptosomal fractions from rat hippocampus contained plasma membrane-associated GR (95) and GR immunoreactivity was observed at the plasma

membranes and vesicle membranes of the hypothalamus (96) while in the CA1, GR was shown in the spine (97). Functionally, JNK and p38 MAPK were activated within 10 min of glucocorticoid administration in primary hippocampal neurons (98). The hypothalamo–pituitary–adrenal (HPA) axis is also subject to rapid negative feedback at the level of the hypothalamus (99). Bath application of glucocorticoids to rat hypothalamic slices caused a rapid suppression of glutamate-mediated excitatory synaptic currents onto CRH neurons that was decreased by a CB1 receptor antagonist (13). The release of endocannabinoids that bind to the CB1 receptor is dependent on corticosterone rapidly acting on the CRH neuron and is dependent on G α_s -driven PKA activation in the CRH neuron (100). However, the release of nitric oxide (NO) that increased GABAergic inhibition (101) onto the CRH neuron was dependent on G $\beta\gamma$ signaling (100). Hence, in the PVN, rapid negative feedback by glucocorticoids on post-synaptic CRH neurons is exerted by a combination of suppression of presynaptic glutamatergic neurons and excitation of presynaptic GABAergic neurons. The identity of the mGR that can signal via both G α_s and G $\beta\gamma$ subunits in the CRH neuron remains unknown.

UNRESOLVED QUESTIONS: mER AND mGR

As is evident in the preceding sections, several questions about the identity of the mER and mGR remain. Though the strongest evidence exists for a post-translationally modified classical ER, there are other viable candidates for the ER (102) based both on evidence of proteins that interact with antibodies raised to the classical ER or to the continued existence of non-genomic effects in ER α KO and ER β KO mice (63). Different ER proteins might be mERs in different tissues (14) or different proteins might be mERs in the same tissue at different times to generate divergent rapid signaling outcomes within the same cell or different tissues that is congruent with incoming stimuli. It is also possible that the time frame of non-genomic signaling might be different when there are different mERs present – a GPCR such as GPR30 may activate the ERK and PKA pathways very transiently whereas ER α or ER β at the membrane may be capable of more sustained activation. Again, this would change the response of the cell to stimuli, depending on the mER present. Second, if the mER is the ER α , the mechanism by which only a small proportion of the total ER α is targeted to the membrane is not known and strengthens the idea that different pools of ER α may exist within cells (30) with different functions. The rationale for the variable amount of ER α at the membrane in some cells versus others in cell lines is also unclear but could allow for 17 β -E to employ non-genomic versus genomic signaling to different extents in different cells, which maybe at different physiological states. Third, apart from the difficulty in studying ER β localization or interaction with other proteins due to the unavailability of a reliable antibody (57), the localization of the ER α variants is also particularly understudied, but the predominance of the 52 kDa form as opposed to the full-length 66 kDa form in the CNS (39, 52) argues for an important role of these variants in estrogen signaling in the brain, as opposed to other classical estrogen-responsive tissues such as the uterus. The rapid actions of these different ER α variants may also oppose each other in some cases (39). It is, therefore, likely that the ratio of different ERs, including mERs

expressed in each cell that determines the cellular response to 17β -E exposure. The significance of these spatially separated and functionally opposing ER populations may be to modulate the hypothalamic and hippocampal neuron response to estrogen and prevent runaway signaling. Fourth, though the GPR30 has a role to play in estrogen-mediated physiology, the acceptance of GPR30 as a mER has not been universal (103) with some investigators preferring to term it as a “collaborator” to the ER α , which is deemed to be the primary mER that signals from the membrane (30). If GPR30 or G α_q -coupled novel proteins are the mERs in some tissues, it is also possible that they crosstalk with classical ER α or ER β , accounting in some studies for the ICI-mediated antagonism of the effect. Though this cooperation between GPR30 and ER α has been shown to have a proliferative effect on ovarian cancer cells (104), a functional effect has not been demonstrated in the hypothalamus. In the ventral hippocampus in male mice, Hart et al. showed an injection of G-1 could increase the phosphorylation of an ERK-sensitive serine site on the ER α at position 118 within 30 min, suggesting that these two receptors could interact with each in the CNS (105). Future studies on the colocalization and the functional interaction between ER α and GPR30 in the hypothalamus will prove useful in this regard. There are far fewer studies on the mGR, when compared to the mER, and most studies investigate the possibility of the mGR being equivalent to the iGR. Surprisingly, the idea that different G-protein subunits, particularly that the G $\beta\gamma$ subunit can support signaling from the unidentified mGR in the CRH neuron and from the G α_q -coupled, STX-activated mER in the POMC neuron is also seen with a traditional GPCR such as GPR30. Whether signaling from both G-protein subunits is concomitant or if one signaling pathway predominates, under certain physiological conditions, is not known.

Lastly, the model generated by the existing literature is that the mER or mGR is at the inner leaflet of the membrane, associated with intrinsic membrane proteins or scaffolds such as caveolin. However, surface biotinylation experiments imply that some part of the mER or mGR is exposed to the extracellular milieu (18). Though an abundance of hydrophobic amino acids in the ligand-binding domain of the mER predicts that this domain might insert into the plasma membrane (106), no study has demonstrated this in the CNS. While recycling of this receptor from the membrane is to be expected, the pathways (Rab-mediated or β -arrestin mediated) are possibly cell-specific and are worthy of more attention since non-genomic signaling is possibly terminated during internalization.

MORPHOLOGICAL AND BEHAVIORAL OUTPUTS IN THE CNS DEPENDENT ON NON-GENOMIC SIGNALING BY ESTROGENS AND GLUCOCORTICOIDS

Though kinase regulation and calcium flux has been shown to occur rapidly when cells are exposed to the hormone in a number of tissues and the idea of non-genomic signaling more accepted than ever before, the relevance of non-genomic signaling for behavior has remained murky. Here, we shall confine ourselves to primarily hypothalamically driven behaviors or possible neural correlates that are either rapidly induced by 17β -E or E2-BSA or that may have a non-genomic component. Finally, we will

discuss the intersection of glucocorticoid rapid signaling in a 17β -E-mediated behavior.

SPINOGENESIS

Estrogen is critical in the display of female sex behavior in rodents by acting on the hypothalamus; retrograde tract tracing using the pseudorabies virus (PRV) injected into the lumbar muscles labeled the lordosis circuit, in particular, the plexus of oxytocin fibers in the vl-VMH (107). In the hypothalamus, estradiol benzoate (EB) injections to ovariectomized rats increased spine density by 48% in the vl-VMH as compared to oil injections (108). Surprisingly, only 3% of PRV-labeled neurons were ER α^+ and these were not the neurons that showed an increase in spine density (109) upon EB administration. These data argue that the increase in spine density in the VMH may be only indirectly dependent on ER α signaling or could be due to another ER while the longer time frames used in the study do not allow us to conclude if there is a non-genomic component. A candidate could be the GPR30 receptor, which is expressed in the vl-VMH; the role of this receptor in spinogenesis in the hypothalamus is not known. Rapid effects on spinogenesis and spine morphology following GR activation have been observed in the hippocampus, but not in the hypothalamus. CA1 neurons from male rat hippocampal slices treated with dexamethasone for 1 h demonstrated a translation-independent increase in spine density, which was lost with co-application of dexamethasone with either the GR antagonist RU-486 or the NMDA receptor blocker MK-801 (95). The proportion of mushroom-shaped and thin-type spines was also increased following GR activation (95).

Does non-genomic signaling play a role in the increase in spine density by estrogens or by glucocorticoids? The mGluR1a antagonist, LY367385 in the ARH decreased phosphorylated cofilin levels that are induced within an hour of EB injection and that are required for actin reorganization and spinogenesis, suggesting that at least some of the initial aspects of spinogenesis are mediated via non-genomic signaling from the ER α -mGluR1a complex at the membrane (110). Though EB can increase filopodial spines in the ARH within 4 h of treatment, mushroom shaped, more stable spines require time frames in excess of 20 h and parallels the time frames required for the full display of lordosis behavior, suggesting that rapid non-genomic signaling is insufficient for formation of stable spines (110). Stabilization of the PSD-95 protein, a scaffolding protein enriched in mushroom-shaped dendritic spines, was dependent on pAkt and also required 48 h of 17β -E treatment in differentiated NG-108 cells (111). Another model for the increase in 17β -E-mediated increase in spine density in the hypothalamus is that 17β -E stimulates PI3K activation pre-synaptically, inducing glutamate release, followed by NMDA receptor activation and ERK signaling post-synaptically, with a subsequent increase in spinophilin protein that was correlated with an increase in stable dendritic spines (112) on the post-synaptic neuron. Both ERK and PI3K signaling are implicated in the increase in spine density, though the formation of mature mushroom spines also appears to require transcription (113). Though there are no reports of glucocorticoid-mediated regulation of spine density in the hypothalamus, suppression of PKA, PKC, MAPK, or PI3K signaling completely blocked GR-mediated spinogenesis in CA1 neurons, suggesting that GR signals through convergent kinase pathways

to increase actin polymerization, which would allow for spine changes (114). In CA1 neurons, treatment with the synthetic glucocorticoid dexamethasone increased p-cofilin levels within 30 min, similar to the rapid induction seen in the ARH by EB (97).

Consistent with the idea that 17 β -E-mediated spinogenesis in the VMH has a functional consequence, cytochalasin D, an actin polymerization inhibitor, blocked the formation of spines in the ARH and reduced lordosis (110) though the relevance of such spinogenesis to sex behavior in rodents (see Sex Behavior or Lordosis in Females) is unknown. **The consequences of spine disruption for other behaviors such as aggressive behavior or male sex behavior (see Male Sex Behavior) are also unknown.** Though most of the studies have shown effects on spine density or morphology in longer time frames that do not allow one to parse non-genomic actions from transcription, spine density on the apical dendrite in the stratum radiatum and in the stratum lacunosum moleculare of the CA1 neuron was increased within 40 min by the ER α selective agonist, propyl pyrazole triol (PPT); however, these were at high doses that were not correlated with doses of PPT that led to an improvement in social memory within 40 min of administration (115). In mature rat cortical neurons, 17 β -E rapidly increased ERK and p21-activated kinase to increase dendritic spine density (116) within 30 min. Though this suggests that non-genomic signaling may be sufficient for an increase in spine density at least in the hippocampus and cortex, this has to be confirmed with experiments that utilize a transcription inhibitor. A similar rapid effect of estrogen on spine density in the hypothalamus has not been demonstrated.

SEX BEHAVIOR OR LORDOSIS IN FEMALES

Lordosis in rodents is a 17 β -E-driven behavior where integration of sensory information within the limbic-hypothalamic circuit culminates in VMH projections onto neurons of the periaqueductal gray (PAG) and spinal cord motor neurons, resulting in the classical lordosis posture (16). A series of elegant experiments from the laboratory of Micevych delineated a microcircuit consisting of neuronal afferents from the ARH to the mPOA (113). In this circuit, the initial membrane-initiated signaling by 17 β -E in the ARH neurons increased PKC θ (117) and released neuropeptide Y that activates β -endorphin expressing ARH neurons that project into the mPOA (118). In the mPOA, β -endorphin released from the ARH caused the internalization of μ -opioid receptors (MOR) within 30 min of E2-BSA (119) administration into the ARH, suggesting that this is a membrane-initiated non-transcriptional event. This transient inhibition of lordosis initiated by MOR internalization is removed 30 h later by progesterone (120). A number of different receptors, including ER α and mGluR1a, play a role in this membrane-initiated signaling to influence lordosis behavior. Not only does ER α and mGluR1a colocalize in about 23% of ER α -expressing neurons but deletion of ER α also blocked MOR internalization in the mPOA (121) and abolishes lordosis (122). Blocking of MOR internalization with application of an mGluR1 antagonist or Cav-1 siRNA into the ARH also, in turn, reduced lordosis (119, 123). Though this suggests that ER α and mGluR1 signal together as in the hippocampus (124), the selective mER agonist, STX in the ARH also induced MOR internalization and lordosis (125), an effect blocked by the

mGluR1a antagonist. This suggests that in the ARH, both ER α and the G α_q -mER may couple to mGluR1a and facilitate lordosis, though the mechanisms by which this occurs are unclear. Apart from these receptors, the localization of the GPR30 in the hypothalamus (126) and the ability of G-1, an agonist at the GPR30 to facilitate lordosis in female ovariectomized mice (127) argues that GPR30 signaling is also important, at least in mice. It is unknown if the ER α , the G α_q -mER, and the GPR30 interact or signal to each other or represent independent parallel signaling pathways that drive lordosis in estrogen primed ovariectomized female rodents, though any model must take into account the necessity of ER α in the VMH for lordosis.

Since Kow et al. demonstrated that E2-BSA alone in the VMH does not induce lordosis (128), we can presume that non-genomic signaling is insufficient but may potentiate or prime lordosis that is in itself dependent on nuclear transcription (129) via a coupled signaling pathway (130). Consistent with this idea, a mouse that possesses an ER α that cannot bind an ERE (ER $\alpha^{-/AA}$) (131) did not display lordosis behavior (132), demonstrating the importance of 17 β -E-bound ER α transcriptional action at an ERE for lordosis. Also, combined E2-BSA and 17 β -E administrations into the VMH increase progesterone receptor (PR) protein levels higher than either hormone alone (133), demonstrating potentiation of transcription by a membrane-limited estrogen conjugate. The reduction of ER α in the VMH using adenovirus-associated shRNA viral vectors that abolished lordosis behavior (29) shows the absolute requirement for the ER α , which could be due to its participation both as a mER and as a nuclear transcription factor in the VMH. An additional role could be the signaling initiated by ER α acting as a mER in the ARH but as a transcription factor that upregulates the PR in the VMH (134), both processes that are required for lordosis. In this scenario, membrane-initiated signaling in the ARH results in transient inhibition of lordosis so that 17 β -E can transcriptionally activate genes such as the PR in the VMH that are important for the full display of lordosis that occurs 30 h later (113).

MALE SEX BEHAVIOR

In males, 17 β -E given intraperitoneally at high concentrations to castrated adult rats increased some aspects of male sex behavior, i.e., genital sniffs and mounting within 15 min, a time frame commensurate with non-genomic actions (135). However, since protein synthesis inhibitors (136) and the presence of the ERE-binding mutant, the ER $\alpha^{-/AA}$ mouse abolished male sex behavior (131), it is reasonable to conclude that male sex behavior, similar to female sex behavior, has a non-genomic component that is not sufficient for the full display of the behavior. Though most of the studies investigate the role of testosterone in males (137, 138), testosterone can be converted to 17 β -E by aromatase (139). In the castrated and testosterone supplemented quail that was treated with the aromatase inhibitor, vorozole, administration of 17 β -E or E2-BSA intracerebroventricularly (icv) increased appetitive sexual behavior but not the final consummatory aspects of sexual behavior within 15 min (140). This appetitive sexual behavior was also blocked by vorozole, administered icv 15 min before behavioral testing (140). This implies that neuroestrogens that are rapidly generated in response to stimuli such as the sight of a female signal

rapidly to initiate appetitive sexual behavior. Though ICI 162,780 antagonized the increase in appetitive sexual behavior seen rapidly with 17β -E administration (140), the exact mER that mediates the rapid regulation of sex behavior or aromatase in the quail or the mouse is unknown. In the male mouse, consistent with the interpretation that arises from the data in the quail, 17β -E administered to aromatase knockout male mice reversed the lack of sex behavior while vorozole decreased sex behavior in C57BL/6 mice within 15 min of administration (141). Stimuli-dependent rapid regulation of aromatase could represent a mechanism to locally elevate neuroestrogens, which then increase sex behavior in male quail or in the male rodent. However, contrary to this expectation, aromatase activity decreased rapidly when male quail were exposed to female stimuli but neuroestrogen concentrations itself increased in the mPOA (142), a nuclei important for male sex behavior. However, it is important to note that the acute action of 17β -E on appetitive sexual behavior in the male quail was always preceded by initial copulation with a receptive female to establish this learned response, wherein the sight of the same female would initiate appetitive sexual behavior (143). In this case, it is possible that prior sexual experience may initiate transcriptional signaling by 17β -E, which in turn primes rapid non-genomic signaling to achieve greater efficiency in subsequent sexual interactions and optimize reproduction; this remains to be tested. Similar to female sex behavior, the non-genomic component in male sex behavior is important for the initial approach and mounting aspects of male sex behavior.

AGGRESSION IN MALES

Castration in male rodents removes both testosterone and estrogens and results in loss of aggression in response to territorial intrusion (144, 145). The importance of estrogen in male aggressive behavior is shown by the deletion of aromatase, which abolished aggression by male mice in a resident-intruder paradigm, though this is possibly also due to organizational defects since 17β -E reinstated aggression only when administered before post-natal day 7 (146). In CD-1 outbred male mice, ER α concentrations in several areas of the circuit involved in aggression such as the bed nucleus of the stria terminalis (BNST), the lateral septum (LS), and the anterior hypothalamus (AH) was higher in more aggressive mice (147). Reduction of ER α in the VMH by adenovirus mediated transfer of siRNA to ER α abolished aggressive behavior in male mice, demonstrating that ER α expression in the VMH is necessary for aggression in adult male mice (29), just as it was important for sex behavior in male mice. In California, beach mice that were castrated and supplemented with testosterone and an aromatase inhibitor so that they possessed testosterone but not estrogen, aggression increased rapidly within 15 min of injection with cyclodextrin-conjugated 17β -E (148). This suggests that testosterone alone was not sufficient to elicit aggressive behavior in a resident-intruder paradigm and that 17β -E is necessary to activate aggression. The time frame and the inability of cycloheximide (149) to decrease this aggression in the beach mouse argue that 17β -E acts non-genomically, though most studies on aggression, with the exception of those cited in this paragraph, in the rat and mouse have used time frames that do not allow us to parse a non-genomic effect from a genomic/coupled signaling effect.

What is the source of 17β -E? In rodents, aromatase is expressed in axon terminals (150–152) particularly in the hypothalamus (153) and brain 17β -E concentrations in the male rat are equal to or higher (~ 8 nM) than in the plasma (154). Hence, 17β -E generated at the synapse has been envisioned to function as a neuromodulator in short time frames (155), in line with the data in male birds where neuroestrogens generated at axon terminals are important in learning in songbirds (156) and in sex behavior in the quail (157, 158). In the castrated male rat, EB synergized with dihydrotestosterone (DHT) to increase aromatase activity (159). In addition, 17β -E can transcriptionally induce the aromatase gene via ER α -c-jun complex binding to AP-1 elements in the brain-specific promoter of aromatase (160). Therefore, similar to spinogenesis, the regulation of aromatase by 17β -E itself may represent another correlate where both rapid, non-genomic and slower, transcriptional mechanisms converge. The areas of the brain wherein this may occur in the rodent are not clear and are being investigated in our laboratory.

Though 17β -E can rapidly increase aggression by modulating neurotransmitter release [(145) and references therein], it may also do via regulating the HPA axis. In the white-crowned sparrow, neither brain 17β -E concentrations nor aromatase activity in nuclei involved in aggression were correlated with aggressive behavior (161) though corticosterone did increase rapidly. What is the link between glucocorticoids and aggression? Though aggression itself is stressful and leads to secretion of glucocorticoids, dominance in an aggressive encounter typically leads to lowering of the corticosterone level (162). However, pre-fight corticosterone levels have been shown to be associated with aggressiveness in fish and acute treatment with adrenocorticotrophic hormone (ACTH) increased fighting behavior in male mice (163) while acute treatment with corticosterone 2 min before an encounter increased aggressiveness in male rats (164) and risk assessment in the open field and elevated plus maze tests (165). Supporting the idea that corticosterone is important in regulation the onset of aggressive behavior, acute treatment with GR antagonists (166, 167) or prevention of glucocorticoid synthesis (164) prevented aggression when presented with social challenge. Since acute central application of corticosterone increased aggression within 7 min and protein synthesis inhibitors did not decrease it, non-genomic signaling by glucocorticoids in the CNS is most likely required to increase aggression (164). Consistent with this idea, acute corticosterone injection also decreased the magnitude of electrical stimulation to the hypothalamus required to elicit attack (168). In addition, corticosterone administration to rats in established colonies did not change aggression, demonstrating that glucocorticoids promote aggression only in ethologically relevant situations such as territorial intrusion (169). However, chronic elevated levels of glucocorticoids, such as seen in stressed or socially defeated animals decrease aggression and increased submissiveness in hamsters (170, 171), mice (172), and rats (173, 174). This is thought to be due to genomic signaling by glucocorticoids though the molecular mechanisms by which glucocorticoids facilitate or inhibit aggression are unclear (175).

Gonadal status can modify the actions of glucocorticoids on the hypothalamus or can regulate corticosterone levels itself. For example, gonadal steroids can modify glucocorticoid-mediated

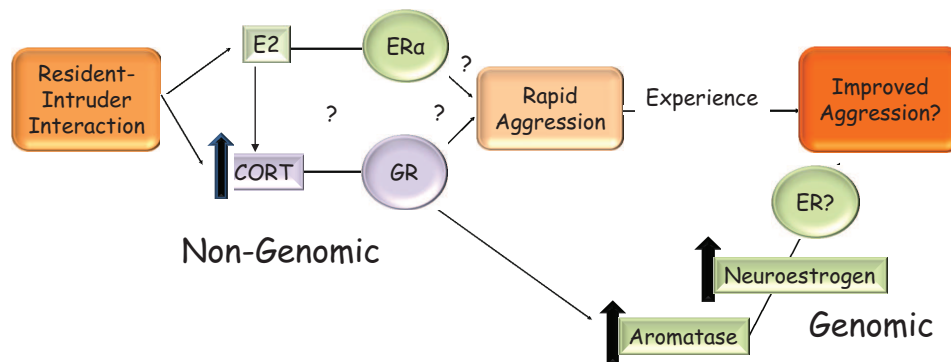


FIGURE 2 | Hypothetical model in the rodent for the rapid and sustained increase in aggression that is seen when an intruder enters a resident's home-cage. On visual and/or olfactory cues, 17 β -E in the brain may increase corticosterone via an unknown mechanism. This increase in corticosterone may rapidly and non-genomically increase aggressive behavior. The mechanism by which corticosterone does so is not known but may involve the regulation of aromatase and could be GR mediated. It is not known if 17 β -E's regulation of aggressive behavior is

solely via corticosterone increase or these are two parallel pathways, which require ER α , as shown in the figure. Elevated corticosterone levels can increase transcription from the brain-specific aromatase promoter, which in turn increases neuroestrogens. This may contribute to shorter latencies of attack in successive bouts. The interaction of estrogen and corticosterone ensures that the resident rodent is aggressive only in the presence of a relevant external cue, i.e., intruder and a relevant internal cue, i.e., sufficient 17 β -E.

negative feedback on CRH. 17 β -E chronically administered to gonadectomized and adrenalectomized female rats treated with high doses of corticosterone increased CRH mRNA levels, thus counteracting negative feedback, while DHT treatment to male rats had an opposing effect and caused a further decrease in CRH levels (176). In female ovariectomized rats, icv, but not systemic, 17 β -E injection increased corticosterone rapidly within 30 min, an effect mimicked with the ER α selective agonist, PPT, injected into the PVN (177). In gonadectomized male rats, CRH mRNA was higher when they were treated with EB but lower when treated with DHT, implying that estrogens can regulate the HPA axis in males also (178). A number of mechanisms may be involved in the rapid increase of aggression by glucocorticoids in response to social challenge, including the regulation of dopamine (179, 180) and serotonin (181, 182) neurotransmission. Here, we will focus on one such mechanism that is responsive to both estrogens and glucocorticoids in the hypothalamus, namely, the regulation of aromatase. Acute stressors, which increase glucocorticoids, increased neuroestrogen concentrations in the PVN and aromatase mRNA in the PVN within an hour in female rats, suggesting that corticosteroid release may also increase neuroestrogen in the hypothalamus (183). Similarly, in male quail, rapid increases in aromatase activity also occurred on restraint stress in the mPOA (184), demonstrating that corticosterone can affect aromatase using a non-genomic signaling mechanism that is yet unknown, in both birds and rodents. Furthermore, aromatase protein levels are increased in response to glucocorticoid treatment in a hypothalamic cell line via a transcriptional mechanism (185), leading to a possible increase in the amount of aromatase that may increase the efficiency of winning future bouts. These data demonstrate that 17 β -E in the brain can increase plasma corticosterone, which in turn can increase aromatase activity and/or transcription in the hypothalamus leading to a positive feedback circuit that may be active under physiologically relevant stressful contexts such territorial intrusion. The relevance of this pathway

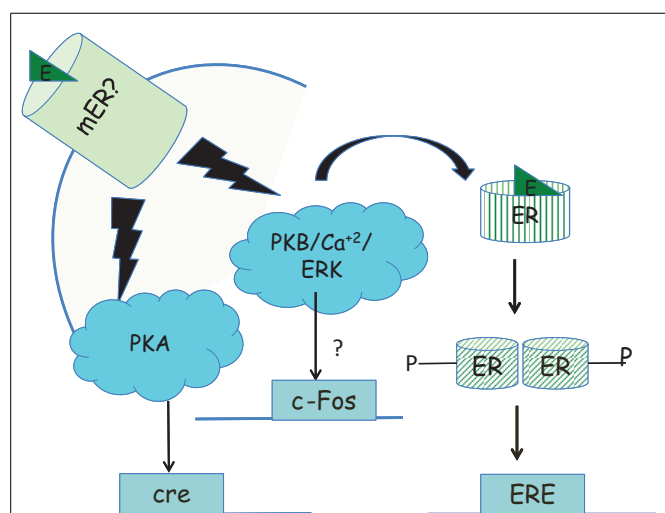


FIGURE 3 | Priming of transcription within neurons by non-genomic signaling. Non-genomic signaling, initiated at the plasma membrane by an unknown mER can activate ERK signaling, which in turn may cause phosphorylation of the ER α and increased transcription from promoters that contain EREs (130). Alternatively, ERK signaling may also lead via unknown mechanisms to increased transcription from the c-Fos promoter (190) while PKA signaling generated by the mER may lead to increased transcription from promoters that have a cAMP response element (CRE) enhancer (189).

(Figure 2) to aggression is under active investigation in our laboratory, with the idea that glucocorticoid control of aggressive behavior is dependent on neuroestrogen concentrations in the brain.

FUNCTIONS OF NON-GENOMIC SIGNALING

One function of non-genomic signaling within a cell could be to provide cells with another pathway by which they can respond

differently to the same endogenous ligand. For example, acute signaling by glucocorticoids increases aggression in an ethologically relevant context but chronic signaling by glucocorticoids increases submission (186). A parallel is seen with the regulation of the CRH neuron by acute or chronic glucocorticoid exposure via the regulation of synaptic inputs in the opposite direction. In the acute phase, rapid signaling by glucocorticoids increases negative feedback on the CRH neuron decreasing the firing of the neuron (187), while chronic exposure to glucocorticoids decreases negative feedback on the CRH and increases CRH mRNA, increasing HPA axis reactivity (188). A second function of membrane-initiated signaling might be to prime nuclear transcription via the phosphorylation of ER α , seen in a neuroblastoma cell line (130) (**Figure 3**), leading to greater transcription from genes with EREs in their promoters. In some cases, rapid signaling could also activate genes that do not have classical EREs in their promoters. For example, PKA signaling induced by 17 β -E in the SK-N-SH neuronal cell line was required to induce the neurotensin/neuromedin gene (189), while E2-BSA in this cell line activated a reporter gene driven by a c-Fos promoter in an ERK-dependent manner (190). The idea that non-genomic signaling might prime later outputs such as behavior is also evident from the studies on sex behavior in female and male rodents. Apart from the potentiation of transcription by rapid signaling,

non-genomic signaling could also decrease transcription if the decrease in transcription is ultimately important in leading to an optimal cellular response. For example, the increase in PI3K and MAPK activation that occurs within 30 min via ER α in the mHypoE-38 hypothalamic cell line is required for the long-term repression of neuropeptide Y by 17 β -E (191). Third, non-genomic signaling via one pathway could also synergize with rapid signaling pathways initiated by other ligands. For instance, 17 β -E-induced increases in calcium in POMC neurons augmented the activation of the calcium-sensitive TRPC channels (192) already upregulated by leptin (65). Additionally, 17 β -E also transcriptionally induced PI3K p85 subunit in the hypothalamus (193, 194) and this combined with the non-genomic signaling-mediated increase in calcium potentiates the PI3K and calcium-mediated TRPC channel opening activated by leptin (70). In this case, 17 β -E utilized both non-genomic, i.e., calcium increase and genomic signaling, i.e., PI3K transcriptional upregulation to synergize with the rapid signaling initiated by the leptin receptor to converge onto neuronal depolarization. Fourth, non-genomic signaling pathways may also synergize with a parallel, independent transcriptional pathway, in response to the same ligand. The rapid action of glucocorticoids in decreasing glutamatergic inputs to the CRH neuron combined with slower transcriptional downregulation of the CRH and vasopressin gene are dual pathways that achieve negative feedback

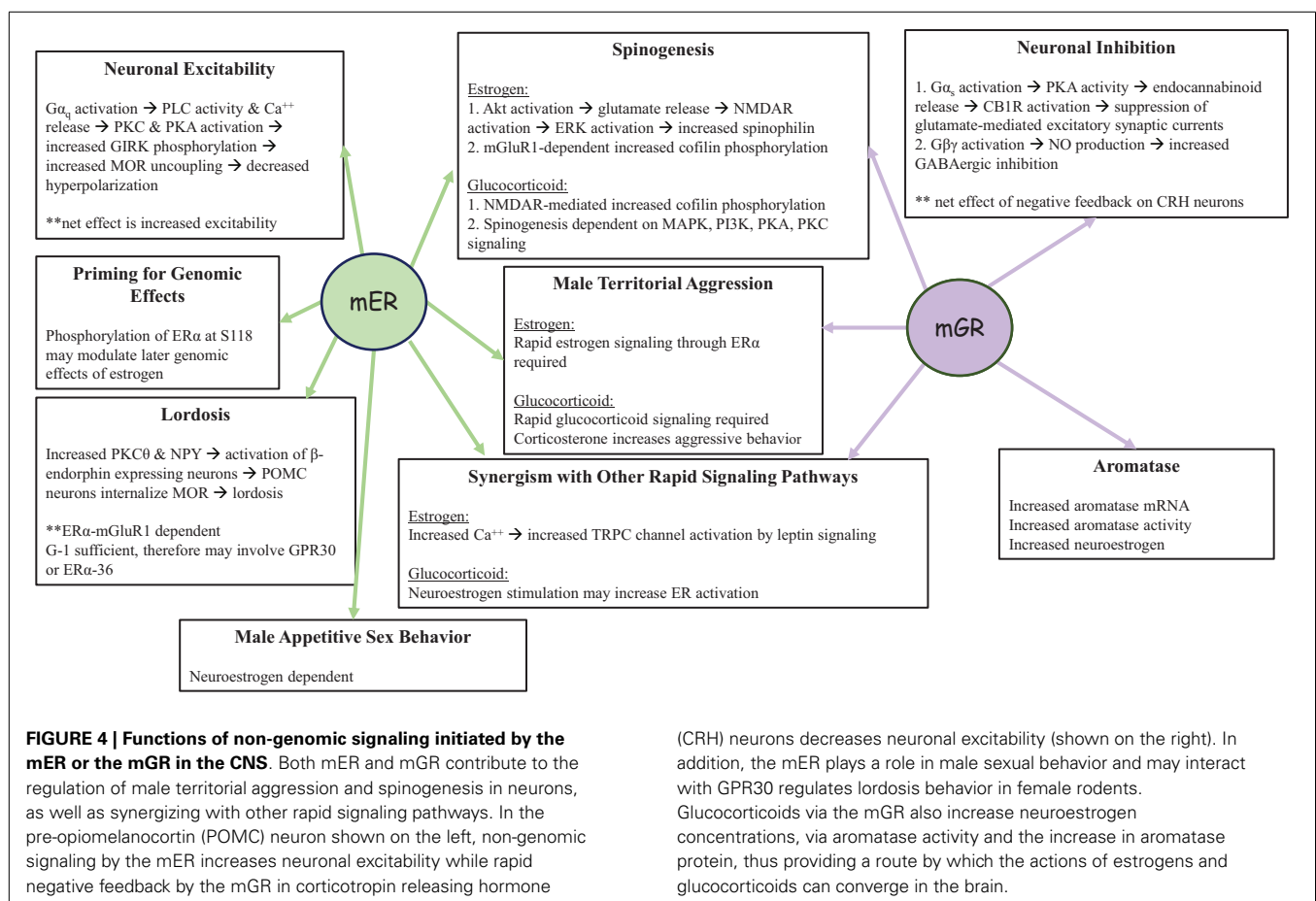


Table 2 | Receptors that are activated or inhibited by the natural estrogen 17 β -estradiol, the natural glucocorticoid, corticosterone, a membrane-limited estrogen conjugate (E2-BSA) or glucocorticoid-conjugate (Dex-BSA), or selective agonists or antagonists.

Hormone/drug	Receptor activation	Receptor antagonism	Reference
17 β -estradiol	All ERs	None	
E2-BSA	All mERs	None	
Propyl pyrazole triol (PPT)	ER α ; 410-fold selectivity over ER β . Also binds GPR30	None reported	(199)
R,R(THC)	ER α	ER β	(200)
Diarylpropionitrile (DPN)	ER β ; 70-fold selectivity over ER α	None reported	(201)
ICI 182,780	GPR30	ER α /ER β	(202)
G-1	GPR30 and ER α -36	None reported	(60)
G-15	ER α or ER β ; activates ERE	GPR30	(203)
G-36	None reported	GPR30	(203)
Corticosterone	All GRs Mineralocorticoid receptor	None reported	(204)
Dexamethasone	All GRs MR (30% of affinity to GR)	None reported	(204)
Dex-BSA	All mGRs	None reported	
RU-486 (mifepristone)	None reported	GR, progesterone receptor	(205)

As can be seen, several selective agonists or antagonists show cross-reactivity to other receptors for estrogens and to other receptors for mineralocorticoids and progestins, in the case of the glucocorticoids. Hence, results obtained using these compounds must be carefully interpreted.

MR, mineralocorticoid receptor.

regulation in the PVN (19). Glucocorticoid-mediated rapid regulation of aromatase combined with the slower transcriptional increase or aromatase mRNA and protein is yet another example where both modes would finally increase neuroestrogens in the hypothalamus. Lastly, non-genomic signaling in one cell type may also synergize with genomic signaling in another cell type to converge onto behavioral outputs. This is demonstrated by the ability of 17 β -E to elicit a rapid calcium increase in hypothalamic astrocytes that is required for the increased synthesis of neuroprogesterone (195). This release of progesterone that binds to the PR that is transcriptionally induced by the liganded ER α in hypothalamic neurons is required for lordosis behavior in rodents (195). Some of these functions of non-genomic signaling are shown in **Figure 4**.

FUTURE DIRECTIONS

Understanding the many facets of non-genomic signaling would also spotlight the role and source of the ligand. The idea that local estrogen synthesis in the brain, i.e., neuroestrogens is important in transducing environmental stimuli to behavior is fairly recent and has mostly been explored in birds (155, 156, 196). In addition, this would bring into focus the role of estrogens in males; traditionally, classical female-typical behaviors such as lordosis have received far more attention in the estrogen field. Second, many of the studies described above were done in rats whereas there are hints that there may be subtle differences in the mouse. For example, though there is very little GPR30 protein reported in the rat amygdala (76), there is high expression of GPR30 in the mouse basolateral amygdala and intra-amygdalar injection of G-1 attenuates behaviors that denote anxiety in the mouse (197). Similarly, though G-1 failed to regulate sex behavior in the female rat (198), it could do so in the female mouse (127). Though studies on rapid non-genomic signaling have become more mainstream, a number of mechanistic aspects of non-genomic signaling and coupled signaling remain unknown including the temporal nature of the shift from non-genomic signaling to genomic signaling. The role of the ER α variants and GR variants in any of these hypothalamically driven behaviors or in spinogenesis is unclear, with almost no studies on the subject. In this regard, it is worth nothing that studies using agonists or antagonists should be careful to verify if the effects are specific – for example, some of the G-1 mediated effects could be via ER α -36 and some of the ICI-mediated effects could be via GPR30 (**Table 2**).

Complementary studies using deletions of the genes in animals or preferably using site-specific or temporally specific deletion would prove useful to establish specificity. The detailed molecular pathways by which 17 β -E regulates spinogenesis or behaviors in the VMH are also not well worked out, though there has been considerable progress in the last decade on the receptors that might be involved. Lastly, understanding non-genomic signaling is possibly a mechanism to convey information about external physiologically relevant stimuli rapidly to the animal to generate both acute and chronic outcomes.

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REFERENCES

1. Nilsson S, Makela S, Treuter E, Tujague M, Thomsen J, Andersson G, et al. Mechanisms of estrogen action. *Physiol Rev* (2001) **81**:1535–65.
2. Hamilton KJ, Arao Y, Korach KS. Estrogen hormone physiology: reproductive findings from estrogen receptor mutant mice. *Reprod Biol* (2014) **14**:3–8. doi:10.1016/j.repbio.2013.12.002
3. Aranda A, Pascual A. Nuclear hormone receptors and gene expression. *Physiol Rev* (2001) **81**:1269–304.
4. Huang P, Chandra V, Rastinejad F. Structural overview of the nuclear receptor superfamily: insights into physiology and therapeutics. *Annu Rev Physiol* (2010) **72**:247–72. doi:10.1146/annurev-physiol-021909-135917

5. Szego CM, Davis JS. Adenosine 3',5'-monophosphate in rat uterus: acute elevation by estrogen. *Proc Natl Acad Sci U S A* (1967) **58**:1711–8. doi:10.1073/pnas.58.4.1711
6. Malyala A, Kelly MJ, Ronnekleiv OK. Estrogen modulation of hypothalamic neurons: activation of multiple signaling pathways and gene expression changes. *Steroids* (2005) **70**:397–406. doi:10.1016/j.steroids.2005.03.004
7. Kelly MJ, Ronnekleiv OK. Membrane-initiated estrogen signaling in hypothalamic neurons. *Mol Cell Endocrinol* (2008) **290**:14–23. doi:10.1016/j.mce.2008.04.014
8. Segal M, Murphy DD. CREB activation mediates plasticity in cultured hippocampal neurons. *Neural Plast* (1998) **6**:1–7. doi:10.1155/NP.1998.1
9. Htun H, Barsony J, Renyi I, Gould DL, Hager GL. Visualization of glucocorticoid receptor translocation and intranuclear organization in living cells with a green fluorescent protein chimera. *Proc Natl Acad Sci U S A* (1996) **93**:4845–50. doi:10.1073/pnas.93.10.4845
10. Hache RJ, Tse R, Reich T, Savory JG, Lefebvre YA. Nucleocytoplasmic trafficking of steroid-free glucocorticoid receptor. *J Biol Chem* (1999) **274**:1432–9. doi:10.1074/jbc.274.3.1432
11. Bouazza B, Debba-Pavard M, Amrani Y, Isaacs L, O'connell D, Ahamed S, et al. Basal p38 mitogen-activated protein kinase regulates unliganded glucocorticoid receptor function in airway smooth muscle cells. *Am J Respir Cell Mol Biol* (2014) **50**:301–15. doi:10.1165/rncmb.2012-0522OC
12. Chen DY, Bambah-Mukku D, Pollonini G, Alberini CM. Glucocorticoid receptors recruit the CaMKIIalpha-BDNF-CREB pathways to mediate memory consolidation. *Nat Neurosci* (2012) **15**:1707–14. doi:10.1038/nn.3266
13. Di S, Malcher-Lopes R, Halmos KC, Tasker JG. Nongenomic glucocorticoid inhibition via endocannabinoid release in the hypothalamus: a fast feedback mechanism. *J Neurosci* (2003) **23**:4850–7.
14. Vasudevan N, Pfaff DW. Membrane-initiated actions of estrogens in neuroendocrinology: emerging principles. *Endocr Rev* (2007) **28**:1–19. doi:10.1210/er.2005-0021
15. Warner M, Gustafsson JA. Nongenomic effects of estrogen: why all the uncertainty? *Steroids* (2006) **71**:91–5. doi:10.1016/j.steroids.2005.09.001
16. Pfaff DW, McCarthy M, Schwartz-Giblin S, Kow LM. Female reproductive behavior. In: Knobil E, Neill J, editors. *The Physiology of Reproduction*. New York, NY: Raven (1994). p. 107–220.
17. Dallman MF. Fast glucocorticoid actions on brain: back to the future. *Front Neuroendocrinol* (2005) **26**:103–8. doi:10.1016/j.yfrne.2005.08.001
18. Micevych P, Dominguez R. Membrane estradiol signaling in the brain. *Front Neuroendocrinol* (2009) **30**:315–27. doi:10.1016/j.yfrne.2009.04.011
19. Hill MN, Tasker JG. Endocannabinoid signaling, glucocorticoid-mediated negative feedback, and regulation of the hypothalamic-pituitary-adrenal axis. *Neuroscience* (2012) **204**:5–16. doi:10.1016/j.neuroscience.2011.12.030
20. Towle AC, Sze PY. Steroid binding to synaptic plasma membrane: differential binding of glucocorticoids and gonadal steroids. *J Steroid Biochem* (1983) **18**:135–43. doi:10.1016/0022-4731(83)90079-1
21. Zheng J, Ali A, Ramirez VD. Steroids conjugated to bovine serum albumin as tools to demonstrate specific steroid neuronal membrane binding sites. *J Psychiatry Neurosci* (1996) **21**:187–97.
22. Micevych P, Kuo J, Christensen A. Physiology of membrane oestrogen receptor signalling in reproduction. *J Neuroendocrinol* (2009) **21**:249–56. doi:10.1111/j.1365-2826.2009.01833.x
23. Filardo EJ. Epidermal growth factor receptor (EGFR) transactivation by estrogen via the G-protein-coupled receptor, GPR30: a novel signaling pathway with potential significance for breast cancer. *J Steroid Biochem Mol Biol* (2002) **80**:231–8. doi:10.1016/S0960-0760(01)00190-X
24. Pietranera L, Brocca ME, Roig P, Lima A, Garcia-Segura LM, De Nicola AF. 17alpha-Oestradiol-induced neuroprotection in the brain of spontaneously hypertensive rats. *J Neuroendocrinol* (2014) **26**:310–20. doi:10.1111/jne.12151
25. Shughrue PJ, Lane MV, Merchenthaler I. Comparative distribution of estrogen receptor-alpha and -beta mRNA in the rat central nervous system. *J Comp Neurol* (1997) **388**:507–25. doi:10.1002/(SICI)1096-9861(19971201)388:4<507::AID-CNE1>3.0.CO;2-6
26. Laflamme N, Nappi RE, Drolet G, Labrie C, Rivest S. Expression and neuro-peptidergic characterization of estrogen receptors (ERalpha and ERbeta) throughout the rat brain: anatomical evidence of distinct roles of each subtype. *J Neurobiol* (1998) **36**:357–78. doi:10.1002/(SICI)1097-4695(19980905)36:3<357::AID-NEU5>3.0.CO;2-V
27. Mitra SW, Hoskin E, Yudkovitz J, Pear L, Wilkinson HA, Hayashi S, et al. Immunolocalization of estrogen receptor beta in the mouse brain: comparison with estrogen receptor alpha. *Endocrinology* (2003) **144**:2055–67. doi:10.1210/en.2002-221069
28. Musatov S, Chen W, Pfaff DW, Kaplitt MG, Ogawa S. RNAi-mediated silencing of estrogen receptor [alpha] in the ventromedial nucleus of hypothalamus abolishes female sexual behaviors. *Proc Natl Acad Sci U S A* (2006) **103**:10456–60. doi:10.1073/pnas.0603045103
29. Sano K, Tsuda MC, Musatov S, Sakamoto T, Ogawa S. Differential effects of site-specific knockdown of estrogen receptor alpha in the medial amygdala, medial pre-optic area, and ventromedial nucleus of the hypothalamus on sexual and aggressive behavior of male mice. *Eur J Neurosci* (2013) **37**:1308–19. doi:10.1111/ejn.12131
30. Levin ER. Membrane oestrogen receptor alpha signalling to cell functions. *J Physiol* (2009) **587**:5019–23. doi:10.1113/jphysiol.2009.177097
31. Zivadinovic D, Gametchu B, Watson CS. Membrane estrogen receptor-alpha levels in MCF-7 breast cancer cells predict cAMP and proliferation responses. *Breast Cancer Res* (2005) **7**:R101–12. doi:10.1186/bcr959
32. Bulayeva NN, Wozniak AL, Lash LL, Watson CS. Mechanisms of membrane estrogen receptor-alpha-mediated rapid stimulation of Ca2+ levels and prolactin release in a pituitary cell line. *Am J Physiol Endocrinol Metab* (2005) **288**:E388–97. doi:10.1152/ajpendo.00349.2004
33. Zivadinovic D, Watson CS. Membrane estrogen receptor-alpha levels predict estrogen-induced ERK1/2 activation in MCF-7 cells. *Breast Cancer Res* (2005) **7**:R130–44. doi:10.1186/bcr959
34. Blaustein JD, Lehman MN, Turcotte JC, Greene G. Estrogen receptors in dendrites and axon terminals in the guinea pig hypothalamus. *Endocrinology* (1992) **131**:281–90. doi:10.1210/endo.131.1.1612006
35. Milner TA, McEwen BS, Hayashi S, Li CJ, Reagan LP, Alves SE. Ultrastructural evidence that hippocampal alpha estrogen receptors are located at extranuclear sites. *J Comp Neurol* (2001) **429**:355–71. doi:10.1002/1096-9861(20010115)429:3<355::AID-CNE1>3.0.CO;2-#
36. Hart SA, Snyder MA, Smejkalova T, Woolley CS. Estrogen mobilizes a subset of estrogen receptor-alpha-immunoreactive vesicles in inhibitory presynaptic boutons in hippocampal CA1. *J Neurosci* (2007) **27**:2102–11. doi:10.1523/JNEUROSCI.5436-06.2007
37. Tabatadze N, Smejkalova T, Woolley CS. Distribution and posttranslational modification of synaptic ERalpha in the adult female rat hippocampus. *Endocrinology* (2013) **154**:819–30. doi:10.1210/en.2012-1870
38. Bondar G, Kuo J, Hamid N, Micevych P. Estradiol-induced estrogen receptor-alpha trafficking. *J Neurosci* (2009) **29**:15323–30. doi:10.1523/JNEUROSCI.2107-09.2009
39. Dominguez R, Micevych P. Estradiol rapidly regulates membrane estrogen receptor alpha levels in hypothalamic neurons. *J Neurosci* (2010) **30**:12589–96. doi:10.1523/JNEUROSCI.1038-10.2010
40. Patel HH, Insel PA. Lipid rafts and caveolae and their role in compartmentation of redox signaling. *Antioxid Redox Signal* (2009) **11**:1357–72. doi:10.1089/ARS.2008.2365
41. Razandi M, Oh P, Pedram A, Schnitzer J, Levin ER. ERs associate with and regulate the production of caveolin: implications for signaling and cellular actions. *Mol Endocrinol* (2002) **16**:100–15. doi:10.1210/mend.16.1.0757
42. Boulware MI, Weick JP, Becklund BR, Kuo SP, Groth RD, Mermelstein PG. Estradiol activates group I and II metabotropic glutamate receptor signaling, leading to opposing influences on cAMP response element-binding protein. *J Neurosci* (2005) **25**:5066–78. doi:10.1523/JNEUROSCI.1427-05.2005
43. Kuo J, Hariri OR, Bondar G, Ogi J, Micevych P. Membrane estrogen receptor-alpha interacts with metabotropic glutamate receptor type 1a to mobilize intracellular calcium in hypothalamic astrocytes. *Endocrinology* (2009) **150**:1369–76. doi:10.1210/en.2008-0994
44. Meitzen J, Mermelstein PG. Estrogen receptors stimulate brain region specific metabotropic glutamate receptors to rapidly initiate signal transduction pathways. *J Chem Neuroanat* (2011) **42**:236–41. doi:10.1016/j.jchemneu.2011.02.002
45. Navarro CE, Abdul Saeed S, Murdock C, Martinez-Fuentes AJ, Arora KK, Krsmanovic LZ, et al. Regulation of cyclic adenosine 3',5'-monophosphate signaling and pulsatile neurosecretion by Gi-coupled plasma membrane estrogen receptors in immortalized gonadotropin-releasing hormone neurons. *Mol Endocrinol* (2003) **17**:1792–804. doi:10.1210/me.2003-0040

46. Chambliss KL, Shaul PW. Estrogen modulation of endothelial nitric oxide synthase. *Endocr Rev* (2002) **23**:665–86. doi:10.1210/er.2001-0045
47. Acconcia F, Ascenzi P, Bocedi A, Spisni E, Tomasi V, Trentalancia A, et al. Palmitoylation-dependent estrogen receptor alpha membrane localization: regulation by 17beta-estradiol. *Mol Biol Cell* (2005) **16**:231–7. doi:10.1091/mbc.E04-07-0547
48. Razandi M, Alton G, Pedram A, Ghonshani S, Webb P, Levin ER. Identification of a structural determinant necessary for the localization and function of estrogen receptor alpha at the plasma membrane. *Mol Cell Biol* (2003) **23**:1633–46. doi:10.1128/MCB.23.5.1633-1646.2003
49. Razandi M, Pedram A, Levin ER. Heat shock protein 27 is required for sex steroid receptor trafficking to and functioning at the plasma membrane. *Mol Cell Biol* (2010) **30**:3249–61. doi:10.1128/MCB.01354-09
50. Razandi M, Pedram A, Merchenthaler I, Greene GL, Levin ER. Plasma membrane estrogen receptors exist and functions as dimers. *Mol Endocrinol* (2004) **18**:2854–65. doi:10.1210/me.2004-0115
51. Tan Z, Zhou LJ, Li Y, Cui YH, Xiang QL, Lin GP, et al. E(2)-BSA activates caveolin-1 via PI(3)K/ERK1/2 and lysosomal degradation pathway and contributes to EPC proliferation. *Int J Cardiol* (2012) **158**:46–53. doi:10.1016/j.ijcard.2010.12.106
52. Dominguez R, Dewing P, Kuo J, Micevych P. Membrane-initiated estradiol signaling in immortalized hypothalamic N-38 neurons. *Steroids* (2013) **78**:607–13. doi:10.1016/j.steroids.2012.12.008
53. Dewire SM, Ahn S, Lefkowitz RJ, Shenoy SK. Beta-arrestins and cell signaling. *Annu Rev Physiol* (2007) **69**:483–510. doi:10.1146/annurev.ph.69.013107.100021
54. Dominguez R, Hu E, Zhou M, Baudry M. 17beta-estradiol-mediated neuroprotection and ERK activation require a pertussis toxin-sensitive mechanism involving GRK2 and beta-arrestin-1. *J Neurosci* (2009) **29**:4228–38. doi:10.1523/JNEUROSCI.0550-09.2009
55. Blaustein JD. Cytoplasmic estrogen receptors in rat brain: immunocytochemical evidence using three antibodies with distinct epitopes. *Endocrinology* (1992) **131**:1336–42. doi:10.1210/endo.131.3.1380440
56. Meredith JM, Auger CJ, Blaustein JD. Down-regulation of estrogen receptor immunoreactivity by 17 beta-estradiol in the guinea pig forebrain. *J Neuroendocrinol* (1994) **6**:639–48. doi:10.1111/j.1365-2826.1994.tb00630.x
57. Snyder MA, Smejkalova T, Forlano PM, Woolley CS. Multiple ERbeta antisera label in ERbeta knockout and null mouse tissues. *J Neurosci Methods* (2010) **188**:226–34. doi:10.1016/j.jneumeth.2010.02.012
58. Abraham IM, Todman MG, Korach KS, Herbison AE. Critical in vivo roles for classical estrogen receptors in rapid estrogen actions on intracellular signaling in mouse brain. *Endocrinology* (2004) **145**:3055–61. doi:10.1210/en.2003-1676
59. Wang Z, Zhang X, Shen P, Loggie BW, Chang Y, Deuel TF. A variant of estrogen receptor- α , hER- α 36: transduction of estrogen- and antiestrogen-dependent membrane-initiated mitogenic signaling. *Proc Natl Acad Sci U S A* (2006) **103**:9063–8. doi:10.1073/pnas.0603339103
60. Kang L, Zhang X, Xie Y, Tu Y, Wang D, Liu Z, et al. Involvement of estrogen receptor variant ER- α 36, not GPR30, in nongenomic estrogen signaling. *Mol Endocrinol* (2010) **24**:709–21. doi:10.1210/me.2009-0317
61. Li L, Haynes MP, Bender JR. Plasma membrane localization and function of the estrogen receptor alpha variant (ER46) in human endothelial cells. *Proc Natl Acad Sci U S A* (2003) **100**:4807–12. doi:10.1073/pnas.0831079100
62. Flouriot G, Brand H, Denger S, Metivier R, Kos M, Reid G, et al. Identification of a new isoform of the human estrogen receptor- α (hER- α) that is encoded by distinct transcripts and that is able to repress hER- α activation function 1. *EMBO J* (2000) **19**:4688–700. doi:10.1093/emboj/19.17.4688
63. Toran-Allerand CD, Guan X, Macluskay NJ, Horvath TL, Diano S, Singh M, et al. ER-X: a novel, plasma membrane-associated, putative estrogen receptor that is regulated during development and after ischemic brain injury. *J Neurosci* (2002) **22**:8391–401.
64. Kelly MJ, Qiu J, Ronnekleiv OK. Estrogen signaling in the hypothalamus. *Vitam Horm* (2005) **71**:123–45. doi:10.1016/S0083-6729(05)71005-0
65. Kelly MJ, Ronnekleiv OK. Membrane-initiated actions of estradiol that regulate reproduction, energy balance and body temperature. *Front Neuroendocrinol* (2012) **33**:376–87. doi:10.1016/j.yfrne.2012.07.002
66. Kelly MJ, Lagrange AH, Wagner EJ, Ronnekleiv OK. Rapid effects of estrogen to modulate G protein-coupled receptors via activation of protein kinase A and protein kinase C pathways. *Steroids* (1999) **64**:64–75. doi:10.1016/S0039-128X(98)00095-6
67. Kelly MJ, Ronnekleiv OK, Ibrahim N, Lagrange AH, Wagner EJ. Estrogen modulation of K(+) channel activity in hypothalamic neurons involved in the control of the reproductive axis. *Steroids* (2002) **67**:447–56. doi:10.1016/S0039-128X(01)00181-7
68. Roepke TA, Qiu J, Bosch MA, Ronnekleiv OK, Kelly MJ. Cross-talk between membrane-initiated and nuclear-initiated oestrogen signalling in the hypothalamus. *J Neuroendocrinol* (2009) **21**:263–70. doi:10.1111/j.1365-2826.2009.01846.x
69. Qiu J, Ronnekleiv OK, Kelly MJ. Modulation of hypothalamic neuronal activity through a novel G-protein-coupled estrogen membrane receptor. *Steroids* (2008) **73**:985–91. doi:10.1016/j.steroids.2007.11.008
70. Qiu J, Fang Y, Ronnekleiv OK, Kelly MJ. Leptin excites proopiomelanocortin neurons via activation of TRPC channels. *J Neurosci* (2010) **30**:1560–5. doi:10.1523/JNEUROSCI.4816-09.2010
71. Kelly MJ, Ronnekleiv OK. A selective membrane estrogen receptor agonist maintains autonomic functions in hypoestrogenic states. *Brain Res* (2013) **1514**:75–82. doi:10.1016/j.brainres.2013.03.020
72. Smith AW, Bosch MA, Wagner EJ, Ronnekleiv OK, Kelly MJ. The membrane estrogen receptor ligand STX rapidly enhances GABAergic signaling in NPY/AgRP neurons: role in mediating the anorexigenic effects of 17beta-estradiol. *Am J Physiol Endocrinol Metab* (2013) **305**:E632–40. doi:10.1152/ajpendo.00281.2013
73. Nestor CC, Kelly MJ, Ronnekleiv OK. Cross-talk between reproduction and energy homeostasis: central impact of estrogens, leptin and kisspeptin signaling. *Horm Mol Biol Clin Investig* (2014) **17**:109–28. doi:10.1515/hmbci-2013-0050
74. Smith AW, Ronnekleiv OK, Kelly MJ. Gq-mER signaling has opposite effects on hypothalamic orexigenic and anorexigenic neurons. *Steroids* (2014) **81**:31–5. doi:10.1016/j.steroids.2013.11.007
75. Thomas P, Pang Y, Filardo EJ, Dong J. Identity of an estrogen membrane receptor coupled to a G protein in human breast cancer cells. *Endocrinology* (2005) **146**:624–32. doi:10.1210/en.2004-1064
76. Brailoiu E, Dun SL, Brailoiu GC, Mizuo K, Sklar LA, Oprea TI, et al. Distribution and characterization of estrogen receptor G protein-coupled receptor 30 in the rat central nervous system. *J Endocrinol* (2007) **193**:311–21. doi:10.1677/JOE-07-0017
77. Filardo E, Quinn J, Pang Y, Graeber C, Shaw S, Dong J, et al. Activation of the novel estrogen receptor G protein-coupled receptor 30 (GPR30) at the plasma membrane. *Endocrinology* (2007) **148**:3236–45. doi:10.1210/en.2006-1605
78. Akama KT, Thompson LI, Milner TA, McEwen BS. Post-synaptic density-95 (PSD-95) binding capacity of G-protein-coupled receptor 30 (GPR30), an estrogen receptor that can be identified in hippocampal dendritic spines. *J Biol Chem* (2013) **288**:6438–50. doi:10.1074/jbc.M112.412478
79. Funakoshi T, Yanai A, Shinoda K, Kawano MM, Mizukami Y. G protein-coupled receptor 30 is an estrogen receptor in the plasma membrane. *Biochem Biophys Res Commun* (2006) **346**:904–10. doi:10.1016/j.bbrc.2006.05.191
80. Cheng SB, Graeber CT, Quinn JA, Filardo EJ. Retrograde transport of the transmembrane estrogen receptor, G-protein-coupled-receptor-30 (GPR30/GPER) from the plasma membrane towards the nucleus. *Steroids* (2011) **76**:892–6. doi:10.1016/j.steroids.2011.02.018
81. Matsuda K, Sakamoto H, Mori H, Hosokawa K, Kawamura A, Itose M, et al. Expression and intracellular distribution of the G protein-coupled receptor 30 in rat hippocampal formation. *Neurosci Lett* (2008) **441**:94–9. doi:10.1016/j.neulet.2008.05.108
82. Revankar CM, Cimino DF, Sklar LA, Arterburn JB, Prossnitz ER. A transmembrane intracellular estrogen receptor mediates rapid cell signaling. *Science* (2005) **307**:1625–30. doi:10.1126/science.1106943
83. Cheng SB, Quinn JA, Graeber CT, Filardo EJ. Down-modulation of the G-protein-coupled estrogen receptor, GPER, from the cell surface occurs via a trans-Golgi-proteasome pathway. *J Biol Chem* (2011) **286**:22441–55. doi:10.1074/jbc.M111.224071
84. Revankar CM, Mitchell HD, Field AS, Burai R, Corona C, Ramesh C, et al. Synthetic estrogen derivatives demonstrate the functionality of intracellular GPR30. *ACS Chem Biol* (2007) **2**:536–44. doi:10.1021/cb700072n

85. Broselid S, Berg KA, Chavera TA, Kahn R, Clarke WP, Olde B, et al. G protein-coupled receptor 30 (GPR30) forms a plasma membrane complex with membrane-associated guanylate kinases (MAGUKs) and protein kinase A-anchoring protein 5 (AKAP5) that constitutively inhibits cAMP production. *J Biol Chem* (2014) **289**:22117–27. doi:10.1074/jbc.M114.566893
86. Heitzer MD, Wolf IM, Sanchez ER, Witchel SF, Defranco DB. Glucocorticoid receptor physiology. *Rev Endocr Metab Disord* (2007) **8**:321–30. doi:10.1007/s11154-007-9059-8
87. Stahn C, Buttgerit F. Genomic and nongenomic effects of glucocorticoids. *Nat Clin Pract Rheumatol* (2008) **4**:525–33. doi:10.1038/ncprheum0898
88. Strehl C, Gaber T, Lowenberg M, Hommes DW, Verhaar AP, Schellmann S, et al. Origin and functional activity of the membrane-bound glucocorticoid receptor. *Arthritis Rheum* (2011) **63**:3779–88. doi:10.1002/art.30637
89. Pujols L, Mullol J, Roca-Ferrer J, Torrego A, Xaubet A, Cidlowski JA, et al. Expression of glucocorticoid receptor alpha- and beta-isoforms in human cells and tissues. *Am J Physiol Cell Physiol* (2002) **283**:C1324–31. doi:10.1152/ajpcell.00363.2001
90. Oakley RH, Cidlowski JA. The biology of the glucocorticoid receptor: new signaling mechanisms in health and disease. *J Allergy Clin Immunol* (2013) **132**:1033–44. doi:10.1016/j.jaci.2013.09.007
91. Bartholome B, Spies CM, Gaber T, Schuchmann S, Berki T, Kunkel D, et al. Membrane glucocorticoid receptors (mGCR) are expressed in normal human peripheral blood mononuclear cells and up-regulated after in vitro stimulation and in patients with rheumatoid arthritis. *FASEB J* (2004) **18**:70–80. doi:10.1096/fj.03-0328com
92. Vernocchi S, Battello N, Schmitz S, Revets D, Billing AM, Turner JD, et al. Membrane glucocorticoid receptor activation induces proteomic changes aligning with classical glucocorticoid effects. *Mol Cell Proteomics* (2013) **12**:1764–79. doi:10.1074/mcp.M112.022947
93. Samarasinghe RA, Di Maio R, Volonte D, Galbiati F, Lewis M, Romero G, et al. Nongenomic glucocorticoid receptor action regulates gap junction intercellular communication and neural progenitor cell proliferation. *Proc Natl Acad Sci U S A* (2011) **108**:16657–62. doi:10.1073/pnas.1102821108
94. McEwen BS, De Kloet R, Wallach G. Interactions in vivo and in vitro of corticoids and progesterone with cell nuclei and soluble macromolecules from rat brain regions and pituitary. *Brain Res* (1976) **105**:129–36. doi:10.1016/0006-8993(76)90928-8
95. Komatsuzaki Y, Murakami G, Tsurugizawa T, Mukai H, Tanabe N, Mitsuhashi K, et al. Rapid spinogenesis of pyramidal neurons induced by activation of glucocorticoid receptors in adult male rat hippocampus. *Biochem Biophys Res Commun* (2005) **335**:1002–7. doi:10.1016/j.bbrc.2005.07.173
96. Liposits Z, Bohn MC. Association of glucocorticoid receptor immunoreactivity with cell membrane and transport vesicles in hippocampal and hypothalamic neurons of the rat. *J Neurosci Res* (1993) **35**:14–9. doi:10.1002/jnr.490350103
97. Jafari M, Seese RR, Babayan AH, Gall CM, Lauterborn JC. Glucocorticoid receptors are localized to dendritic spines and influence local actin signaling. *Mol Neurobiol* (2012) **46**:304–15. doi:10.1007/s12035-012-8288-3
98. Qi AQ, Qiu J, Xiao L, Chen YZ. Rapid activation of JNK and p38 by glucocorticoids in primary cultured hippocampal cells. *J Neurosci Res* (2005) **80**:510–7. doi:10.1002/jnr.20491
99. Evanson NK, Tasker JG, Hill MN, Hillard CJ, Herman JP. Fast feedback inhibition of the HPA axis by glucocorticoids is mediated by endocannabinoid signaling. *Endocrinology* (2010) **151**:4811–9. doi:10.1210/en.2010-0285
100. Di S, Maxson MM, Franco A, Tasker JG. Glucocorticoids regulate glutamate and GABA synapse-specific retrograde transmission via divergent nongenomic signaling pathways. *J Neurosci* (2009) **29**:393–401. doi:10.1523/JNEUROSCI.4546-08.2009
101. Di S, Malcher-Lopes R, Marcheselli VL, Bazan NG, Tasker JG. Rapid glucocorticoid-mediated endocannabinoid release and opposing regulation of glutamate and gamma-aminobutyric acid inputs to hypothalamic magnocellular neurons. *Endocrinology* (2005) **146**:4292–301. doi:10.1210/en.2005-0610
102. Toran-Allerand CD. Minireview: a plethora of estrogen receptors in the brain: where will it end? *Endocrinology* (2004) **145**:1069–74. doi:10.1210/en.2003-1462
103. Otto C, Rohde-Schulz B, Schwarz G, Fuchs I, Klewer M, Brittain D, et al. G protein-coupled receptor 30 localizes to the endoplasmic reticulum and is not activated by estradiol. *Endocrinology* (2008) **149**:4846–56. doi:10.1210/en.2008-0269
104. Albanito L, Lappano R, Madeo A, Chimento A, Prossnitz ER, Cappello AR, et al. G-protein-coupled receptor 30 and estrogen receptor-alpha are involved in the proliferative effects induced by atrazine in ovarian cancer cells. *Environ Health Perspect* (2008) **116**:1648–55. doi:10.1289/ehp.11297
105. Hart D, Nilges M, Pollard K, Lynn T, Patsos O, Shiel C, et al. Activation of the G-protein coupled receptor 30 (GPR30) has different effects on anxiety in male and female mice. *Steroids* (2014) **81**:49–56. doi:10.1016/j.steroids.2013.11.004
106. Kumar V, Green S, Staub A, Chambon P. Localisation of the oestradiol-binding and putative DNA-binding domains of the human oestrogen receptor. *EMBO J* (1986) **5**:2231–6.
107. Daniels D, Miselis RR, Flanagan-Cato LM. Central neuronal circuit innervating the lordosis-producing muscles defined by transneuronal transport of pseudorabies virus. *J Neurosci* (1999) **19**:2823–33.
108. Calizo LH, Flanagan-Cato LM. Estrogen selectively regulates spine density within the dendritic arbor of rat ventromedial hypothalamic neurons. *J Neurosci* (2000) **20**:1589–96.
109. Flanagan-Cato LM, Calizo LH, Daniels D. The synaptic organization of VMH neurons that mediate the effects of estrogen on sexual behavior. *Horm Behav* (2001) **40**:178–82. doi:10.1006/hbeh.2001.1679
110. Christensen A, Dewing P, Micevych P. Membrane-initiated estradiol signaling induces spinogenesis required for female sexual receptivity. *J Neurosci* (2011) **31**:17583–9. doi:10.1523/JNEUROSCI.3030-11.2011
111. Akama KT, McEwen BS. Estrogen stimulates postsynaptic density-95 rapid protein synthesis via the Akt/protein kinase B pathway. *J Neurosci* (2003) **23**:2333–9.
112. Schwarz JM, Liang SL, Thompson SM, McCarthy MM. Estradiol induces hypothalamic dendritic spines by enhancing glutamate release: a mechanism for organizational sex differences. *Neuron* (2008) **58**:584–98. doi:10.1016/j.neuron.2008.03.008
113. Micevych P, Christensen A. Membrane-initiated estradiol actions mediate structural plasticity and reproduction. *Front Neuroendocrinol* (2012) **33**:331–41. doi:10.1016/j.yfrne.2012.07.003
114. Komatsuzaki Y, Hatanaka Y, Murakami G, Mukai H, Hojo Y, Saito M, et al. Corticosterone induces rapid spinogenesis via synaptic glucocorticoid receptors and kinase networks in hippocampus. *PLoS One* (2012) **7**:e34124. doi:10.1371/journal.pone.0034124
115. Phan A, Lancaster KE, Armstrong JN, Macluskus NJ, Choleris E. Rapid effects of estrogen receptor alpha and beta selective agonists on learning and dendritic spines in female mice. *Endocrinology* (2011) **152**:1492–502. doi:10.1210/en.2010-1273
116. Srivastava DP, Woolfrey KM, Liu F, Brandon NJ, Penzes P. Estrogen receptor beta activity modulates synaptic signaling and structure. *J Neurosci* (2010) **30**:13454–60. doi:10.1523/JNEUROSCI.3264-10.2010
117. Dewing P, Christensen A, Bondar G, Micevych P. Protein kinase C signaling in the hypothalamic arcuate nucleus regulates sexual receptivity in female rats. *Endocrinology* (2008) **149**:5934–42. doi:10.1210/en.2008-0847
118. Sinchak K, Micevych P. Visualizing activation of opioid circuits by internalization of G protein-coupled receptors. *Mol Neurobiol* (2003) **27**:197–222. doi:10.1385/MN:27:2:197
119. Dewing P, Boulware MI, Sinchak K, Christensen A, Mermelstein PG, Micevych P. Membrane estrogen receptor-alpha interactions with metabotropic glutamate receptor 1a modulate female sexual receptivity in rats. *J Neurosci* (2007) **27**:9294–300. doi:10.1523/JNEUROSCI.0592-07.2007
120. Sinchak K, Micevych PE. Progesterone blockade of estrogen activation of mu-opioid receptors regulates reproductive behavior. *J Neurosci* (2001) **21**:5723–9.
121. Micevych PE, Rissman EF, Gustafsson JA, Sinchak K. Estrogen receptor-alpha is required for estrogen-induced mu-opioid receptor internalization. *J Neurosci Res* (2003) **71**:802–10. doi:10.1002/jnr.10526
122. Ogawa S, Eng V, Taylor J, Lubahn DB, Korach KS, Pfaff DW. Roles of estrogen receptor-alpha gene expression in reproduction-related behaviors in female mice. *Endocrinology* (1998) **139**:5070–81. doi:10.1210/en.139.12.5070
123. Christensen A, Micevych P. CAV1 siRNA reduces membrane estrogen receptor-alpha levels and attenuates sexual receptivity. *Endocrinology* (2012) **153**:3872–7. doi:10.1210/en.2012-1312
124. Mermelstein PG, Micevych PE. Nervous system physiology regulated by membrane estrogen receptors. *Rev Neurosci* (2008) **19**:413–24. doi:10.1515/REVNEURO.2008.19.6.413

125. Christensen A, Micevych P. A novel membrane estrogen receptor activated by STX induces female sexual receptivity through an interaction with mGluR1a. *Neuroendocrinology* (2013) **97**:363–8. doi:10.1159/000351077
126. Hazell GG, Yao ST, Roper JA, Prossnitz ER, O'carroll AM, Lolait SJ. Localisation of GPR30, a novel G protein-coupled oestrogen receptor, suggests multiple functions in rodent brain and peripheral tissues. *J Endocrinol* (2009) **202**:223–36. doi:10.1677/JOE-09-0066
127. Anchan D, Clark S, Pollard K, Vasudevan N. GPR30 activation decreases anxiety in the open field test but not in the elevated plus maze test in female mice. *Brain Behav* (2014) **4**:51–9. doi:10.1002/brb3.197
128. Kow LM, Pfaff DW. The membrane actions of estrogens can potentiate their lordosis behavior-facilitating genomic actions. *Proc Natl Acad Sci U S A* (2004) **101**:12354–7. doi:10.1073/pnas.0404889101
129. Pfaff DW, Ogawa S, Kia HK, Vasudevan N, Krebs C, Frohlich J, et al. Genetic mechanisms in neural and hormonal controls over female reproductive behaviors. In: Pfaff DW, Arnold A, Etgen A, Fahrbach S, Rubin R, editors. *Hormones, Brain and Behavior*. San Diego, CA: Academic Press (2002). p. 441–60.
130. Clark S, Rainville J, Zhao X, Katzenellenbogen BS, Pfaff D, Vasudevan N. Estrogen receptor-mediated transcription involves the activation of multiple kinase pathways in neuroblastoma cells. *J Steroid Biochem Mol Biol* (2014) **139**:45–53. doi:10.1016/j.jsbmb.2013.09.010
131. McDevitt MA, Glidewell-Kenney C, Weiss J, Chambon P, Jameson JL, Levine JE. Estrogen response element-independent estrogen receptor (ER)-alpha signaling does not rescue sexual behavior but restores normal testosterone secretion in male ERalpha knockout mice. *Endocrinology* (2007) **148**:5288–94. doi:10.1210/en.2007-0673
132. McDevitt MA, Glidewell-Kenney C, Jimenez MA, Ahearn PC, Weiss J, Jameson JL, et al. New insights into the classical and non-classical actions of estrogen: evidence from estrogen receptor knock-out and knock-in mice. *Mol Cell Endocrinol* (2008) **290**:24–30. doi:10.1016/j.mce.2008.04.003
133. Sa SI, Pereira PA, Malikov V, Ferreira IM, Madeira MD. The role of plasma membrane estrogen receptors in mediating the estrogen induction of progesterone receptors in hypothalamic ventromedial neurons. *J Comp Neurol* (2014) **522**:298–307. doi:10.1002/cne.23396
134. Parsons B, McEwen B, Pfaff D. A discontinuous schedule of estradiol treatment is sufficient to activate progesterone-facilitated feminine sexual behavior and to increase cytosol receptors for progestins in the hypothalamus of the rat. *Endocrinology* (1982) **110**:613–9. doi:10.1210/endo-110-2-613
135. Cross E, Roselli CE. 17beta-estradiol rapidly facilitates chemoinvestigation and mounting in castrated male rats. *Am J Physiol* (1999) **276**:R1346–50.
136. McGinnis MY, Kahn DF. Inhibition of male sexual behavior by intracranial implants of the protein synthesis inhibitor anisomycin into the medial preoptic area of the rat. *Horm Behav* (1997) **31**:15–23. doi:10.1006/hbeh.1997.1367
137. Soma KK. Testosterone and aggression: Berthold, birds and beyond. *J Neuroendocrinol* (2006) **18**:543–51. doi:10.1111/j.1365-2826.2006.01440.x
138. Gleason ED, Fuxjager MJ, Oyegbile TO, Marler CA. Testosterone release and social context: when it occurs and why. *Front Neuroendocrinol* (2009) **30**:460–9. doi:10.1016/j.yfrne.2009.04.009
139. Naftolin F, MacLusky NJ. Aromatase in the central nervous system. *Cancer Res* (1982) **42**:3274s–6s.
140. Seredynski AL, Balthazart J, Christophe VJ, Ball GF, Cornil CA. Neuroestrogens rapidly regulate sexual motivation but not performance. *J Neurosci* (2013) **33**:164–74. doi:10.1523/JNEUROSCI.2557-12.2013
141. Taziaux M, Keller M, Bakker J, Balthazart J. Sexual behavior activity tracks rapid changes in brain estrogen concentrations. *J Neurosci* (2007) **27**:6563–72. doi:10.1523/JNEUROSCI.1797-07.2007
142. Dickens MJ, De Bournonville C, Balthazart J, Cornil CA. Relationships between rapid changes in local aromatase activity and estradiol concentrations in male and female quail brain. *Horm Behav* (2014) **65**:154–64. doi:10.1016/j.yhbeh.2013.12.011
143. Cornil CA, Ball GF, Balthazart J. Rapid control of male typical behaviors by brain-derived estrogens. *Front Neuroendocrinol* (2012) **33**:425–46. doi:10.1016/j.yfrne.2012.08.003
144. Floody O, Pfaff D. Steroid hormones and aggressive behavior: approaches to the study of hormone-sensitive brain mechanisms for behavior. *Res Publ Assoc Res Nerv Ment Dis* (1974) **52**:149–85.
145. Nelson RJ, Trainor BC. Neural mechanisms of aggression. *Nat Rev Neurosci* (2007) **8**:536–46. doi:10.1038/nrn2174
146. Toda K, Saibara T, Okada T, Onishi S, Shizuta Y. A loss of aggressive behaviour and its reinstatement by oestrogen in mice lacking the aromatase gene (Cyp19). *J Endocrinol* (2001) **168**:217–20. doi:10.1677/joe.0.1680217
147. Trainor BC, Greiwe KM, Nelson RJ. Individual differences in estrogen receptor alpha in select brain nuclei are associated with individual differences in aggression. *Horm Behav* (2006) **50**:338–45. doi:10.1016/j.yhbeh.2006.04.002
148. Trainor BC, Lin S, Finy MS, Rowland MR, Nelson RJ. Photoperiod reverses the effects of estrogens on male aggression via genomic and nongenomic pathways. *Proc Natl Acad Sci U S A* (2007) **104**:9840–5. doi:10.1073/pnas.0701819104
149. Laredo SA, Villalon Landeros R, Dooley JC, Steinman MQ, Orr V, Silva AL, et al. Nongenomic effects of estradiol on aggression under short day photoperiods. *Horm Behav* (2013) **64**:557–65. doi:10.1016/j.yhbeh.2013.06.002
150. Naftolin F, Horvath TL, Jakab RL, Leranthy C, Harada N, Balthazart J. Aromatase immunoreactivity in axon terminals of the vertebrate brain. An immunocytochemical study on quail, rat, monkey and human tissues. *Neuroendocrinology* (1996) **63**:149–55. doi:10.1159/000126951
151. Horvath TL, Roa-Pena L, Jakab RL, Simpson ER, Naftolin F. Aromatase in axonal processes of early postnatal hypothalamic and limbic areas including the cingulate cortex. *J Steroid Biochem Mol Biol* (1997) **61**:349–57. doi:10.1016/S0960-0760(97)80032-5
152. Naftolin F, Horvath TL, Balthazart J. Estrogen synthetase (aromatase) immunohistochemistry reveals concordance between avian and rodent limbic systems and hypothalamus. *Exp Biol Med (Maywood)* (2001) **226**:717–25.
153. Wu MV, Manoli DS, Fraser EJ, Coats JK, Tollkuhn J, Honda S, et al. Estrogen masculinizes neural pathways and sex-specific behaviors. *Cell* (2009) **139**:61–72. doi:10.1016/j.cell.2009.07.036
154. Hojo Y, Murakami G, Mukai H, Higo S, Hatanaka Y, Ogiue-Ikeda M, et al. Estrogen synthesis in the brain – role in synaptic plasticity and memory. *Mol Cell Endocrinol* (2008) **290**:31–43. doi:10.1016/j.mce.2008.04.017
155. Saldanha CJ, Remage-Healey L, Schlinger BA. Synaptocrine signaling: steroid synthesis and action at the synapse. *Endocr Rev* (2011) **32**:532–49. doi:10.1210/er.2011-0004
156. Remage-Healey L, Oyama RK, Schlinger BA. Elevated aromatase activity in forebrain synaptic terminals during song. *J Neuroendocrinol* (2009) **21**:191–9. doi:10.1111/j.1365-2826.2009.01820.x
157. Balthazart J, Baillien M, Cornil CA, Ball GF. Preoptic aromatase modulates male sexual behavior: slow and fast mechanisms of action. *Physiol Behav* (2004) **83**:247–70. doi:10.1016/j.physbeh.2004.08.025
158. Cornil CA, Seredynski AL, de Bournonville C, Dickens MJ, Charlier TD, Ball GF, et al. Rapid control of reproductive behaviour by locally synthesised oestrogens: focus on aromatase. *J Neuroendocrinol* (2013) **25**:1070–8. doi:10.1111/jne.12062
159. Roselli CE. Synergistic induction of aromatase activity in the rat brain by estradiol and 5 alpha-dihydrotestosterone. *Neuroendocrinology* (1991) **53**:79–84. doi:10.1159/000125701
160. Yilmaz MB, Wolfe A, Cheng YH, Glidewell-Kenney C, Jameson JL, Bulun SE. Aromatase promoter 1f is regulated by estrogen receptor alpha (ESR1) in mouse hypothalamic neuronal cell lines. *Biol Reprod* (2009) **81**:956–65. doi:10.1095/biolreprod.109.077206
161. Charlier TD, Newman AE, Heimovics SA, Po KW, Saldanha CJ, Soma KK. Rapid effects of aggressive interactions on aromatase activity and oestradiol in discrete brain regions of wild male white-crowned sparrows. *J Neuroendocrinol* (2011) **23**:742–53. doi:10.1111/j.1365-2826.2011.02170.x
162. Bronson FH, Eleftheriou BE. Adrenal response to fighting in mice: separation of physical and psychological causes. *Science* (1965) **147**:627–8. doi:10.1126/science.147.3658.627
163. Mainardi M, Valenti G, Valsecchi P, Parmigiani S, Brain PF, Vescovi P. Acute effects of low doses of two ACTH preparations on fighting behavior in male mice. *Med Sci Res* (1987) **15**:649–50.
164. Mikics E, Kruk MR, Haller J. Genomic and non-genomic effects of glucocorticoids on aggressive behavior in male rats. *Psychoneuroendocrinology* (2004) **29**:618–35. doi:10.1016/S0306-4530(03)00090-8
165. Mikics E, Barsy B, Barsvari B, Haller J. Behavioral specificity of non-genomic glucocorticoid effects in rats: effects on risk assessment in the elevated plus-maze and the open-field. *Horm Behav* (2005) **48**:152–62. doi:10.1016/j.yhbeh.2005.02.002
166. Schjolden J, Basic D, Winberg S. Aggression in rainbow trout is inhibited by both MR and GR antagonists. *Physiol Behav* (2009) **98**:625–30. doi:10.1016/j.physbeh.2009.09.018

167. Chang C, Li CY, Earley RL, Hsu Y. Aggression and related behavioral traits: the impact of winning and losing and the role of hormones. *Integr Comp Biol* (2012) **52**:801–13. doi:10.1093/icb/ics057
168. Kruk MR, Halasz J, Meelis W, Haller J. Fast positive feedback between the adrenocortical stress response and a brain mechanism involved in aggressive behavior. *Behav Neurosci* (2004) **118**:1062–70. doi:10.1037/0735-7044.118.5.1062
169. Mikics E, Barsy B, Haller J. The effect of glucocorticoids on aggressiveness in established colonies of rats. *Psychoneuroendocrinology* (2007) **32**:160–70. doi:10.1016/j.psyneuen.2006.12.002
170. Hayden-Hixson DM, Ferris CF. Cortisol exerts site-, context- and dose-dependent effects on agonistic responding in hamsters. *J Neuroendocrinol* (1991) **3**:613–22. doi:10.1111/j.1365-2826.1991.tb00326.x
171. Hayden-Hixson DM, Ferris CF. Steroid-specific regulation of agonistic responding in the anterior hypothalamus of male hamsters. *Physiol Behav* (1991) **50**:793–9. doi:10.1016/0031-9384(91)90020-O
172. Leshner AI, Korn SJ, Mixon JE, Rosenthal C, Besser AK. Effects of corticosterone on submissiveness in mice: some temporal and theoretical considerations. *Physiol Behav* (1980) **24**:283–8. doi:10.1016/0031-9384(80)90087-6
173. Blanchard DC, Spencer RL, Weiss SM, Blanchard RJ, McEwen B, Sakai RR. Visible burrow system as a model of chronic social stress: behavioral and neuroendocrine correlates. *Psychoneuroendocrinology* (1995) **20**:117–34. doi:10.1016/0306-4530(94)E0045-B
174. Blanchard RJ, Dulloog L, Markham C, Nishimura O, Nikulina Compton J, Jun A, et al. Sexual and aggressive interactions in a visible burrow system with provisioned burrows. *Physiol Behav* (2001) **72**:245–54. doi:10.1016/S0031-9384(00)00403-0
175. Haller J. The glucocorticoid/aggression relationship in animals and humans: an analysis sensitive to behavioral characteristics, glucocorticoid secretion patterns, and neural mechanisms. *Curr Top Behav Neurosci* (2014) **17**:73–109. doi:10.1007/7854_2014_284
176. Patchev VK, Almeida OF. Gonadal steroids exert facilitating and “buffering” effects on glucocorticoid-mediated transcriptional regulation of corticotropin-releasing hormone and corticosteroid receptor genes in rat brain. *J Neurosci* (1996) **16**:7077–84.
177. Liu J, Bisschop PH, Eggels L, Foppen E, Fliers E, Zhou JN, et al. Intrahypothalamic estradiol modulates hypothalamus-pituitary-adrenal-axis activity in female rats. *Endocrinology* (2012) **153**:3337–44. doi:10.1210/en.2011-2176
178. Lund TD, Munson DJ, Haldy ME, Handa RJ. Androgen inhibits, while oestrogen enhances, restraint-induced activation of neuropeptide neurones in the paraventricular nucleus of the hypothalamus. *J Neuroendocrinol* (2004) **16**:272–8. doi:10.1111/j.0953-8194.2004.01167.x
179. Anstrom KK, Miczek KA, Budygin EA. Increased phasic dopamine signaling in the mesolimbic pathway during social defeat in rats. *Neuroscience* (2009) **161**:3–12. doi:10.1016/j.neuroscience.2009.03.023
180. Barik J, Marti F, Morel C, Fernandez SP, Lanteri C, Godeheu G, et al. Chronic stress triggers social aversion via glucocorticoid receptor in dopaminergic neurons. *Science* (2013) **339**:332–5. doi:10.1126/science.1226767
181. Haller J, Halasz J, Makara GB, Kruk MR. Acute effects of glucocorticoids: behavioral and pharmacological perspectives. *Neurosci Biobehav Rev* (1998) **23**:337–44. doi:10.1016/S0149-7634(98)00035-9
182. Summers CH, Winberg S. Interactions between the neural regulation of stress and aggression. *J Exp Biol* (2006) **209**:4581–9. doi:10.1242/jeb.02565
183. Liu J, Hu P, Qi XR, Meng FT, Kalsbeek A, Zhou JN. Acute restraint stress increases intrahypothalamic oestradiol concentrations in conjunction with increased hypothalamic oestrogen receptor beta and aromatase mRNA expression in female rats. *J Neuroendocrinol* (2011) **23**:435–43. doi:10.1111/j.1365-2826.2011.02123.x
184. Dickens MJ, Cornil CA, Balthazart J. Neurochemical control of rapid stress-induced changes in brain aromatase activity. *J Neuroendocrinol* (2013) **25**:329–39. doi:10.1111/jne.12012
185. Brooks DC, Zhao H, Yilmaz MB, Coon VJ, Bulun SE. Glucocorticoid-induction of hypothalamic aromatase via its brain-specific promoter. *Mol Cell Endocrinol* (2012) **362**:85–90. doi:10.1016/j.mce.2012.05.012
186. Haller J, Mikics E, Makara GB. The effects of non-genomic glucocorticoid mechanisms on bodily functions and the central neural system. A critical evaluation of findings. *Front Neuroendocrinol* (2008) **29**:273–91. doi:10.1016/j.yfrne.2007.10.004
187. Hill MN, Patel S, Campolongo P, Tasker JG, Wotjak CT, Bains JS. Functional interactions between stress and the endocannabinoid system: from synaptic signaling to behavioral output. *J Neurosci* (2010) **30**:14980–6. doi:10.1523/JNEUROSCI.4283-10.2010
188. Choi DC, Evanson NK, Furay AR, Ulrich-Lai YM, Ostrander MM, Herman JP. The anteroventral bed nucleus of the stria terminalis differentially regulates hypothalamic-pituitary-adrenocortical axis responses to acute and chronic stress. *Endocrinology* (2008) **149**:818–26. doi:10.1210/en.2007-0883
189. Watters JJ, Dorsa DM. Transcriptional effects of estrogen on neuronal neurotensin gene expression involve cAMP/protein kinase A dependent mechanisms. *J Neurosci* (1998) **18**:6672–80.
190. Watters JJ, Campbell JS, Cunningham MJ, Krebs EG, Dorsa DM. Rapid membrane effects of steroids in neuroblastoma cells: effects of estrogen on mitogen activated protein kinase signalling cascade and c-fos immediate early gene transcription. *Endocrinology* (1997) **138**:4030–3. doi:10.1210/endo.138.9.5489
191. Titolo D, Mayer CM, Dhillon SS, Cai F, Belsham DD. Estrogen facilitates both phosphatidylinositol 3-kinase/Akt and ERK1/2 mitogen-activated protein kinase membrane signaling required for long-term neuropeptide Y transcriptional regulation in clonal, immortalized neurons. *J Neurosci* (2008) **28**:6473–82. doi:10.1523/JNEUROSCI.0514-08.2008
192. Constantin S, Caligioni CS, Stojilkovic S, Wray S. Kisspeptin-10 facilitates a plasma membrane-driven calcium oscillator in gonadotropin-releasing hormone-1 neurons. *Endocrinology* (2009) **150**:1400–12. doi:10.1210/en.2008-0979
193. Malyala A, Pattee P, Nagalla SR, Kelly MJ, Ronnekleiv OK. Suppression subtractive hybridization and microarray identification of estrogen-regulated hypothalamic genes. *Neurochem Res* (2004) **29**:1189–200. doi:10.1023/B:NERE.0000023606.13670.1d
194. Roepke TA, Malyala A, Bosch MA, Kelly MJ, Ronnekleiv OK. Estrogen regulation of genes important for K⁺ channel signaling in the arcuate nucleus. *Endocrinology* (2007) **148**:4937–51. doi:10.1210/en.2007-0605
195. Micevych PE, Chaban V, Ogi J, Dewing P, Lu JK, Sinchak K. Estradiol stimulates progesterone synthesis in hypothalamic astrocyte cultures. *Endocrinology* (2007) **148**:782–9. doi:10.1210/en.2006-0774
196. Remage-Healey L, Saldanha CJ, Schlinger BA. Estradiol synthesis and action at the synapse: evidence for “synaptocrine” signaling. *Front Endocrinol* (2011) **2**:28. doi:10.3389/fendo.2011.00028
197. Tian Z, Wang Y, Zhang N, Guo YY, Feng B, Liu SB, et al. Estrogen receptor GPR30 exerts anxiolytic effects by maintaining the balance between GABAergic and glutamatergic transmission in the basolateral amygdala of ovariectomized mice after stress. *Psychoneuroendocrinology* (2013) **38**:2218–33. doi:10.1016/j.psyneuen.2013.04.011
198. Lebesgue D, Reyna-Neyra A, Huang X, Etgen AM. GPR30 differentially regulates short latency responses of luteinizing hormone and prolactin secretion to oestradiol. *J Neuroendocrinol* (2009) **21**:743–52. doi:10.1111/j.1365-2826.2009.01893.x
199. Petrie WK, Dennis MK, Hu C, Dai D, Arterburn JB, Smith HO, et al. G protein-coupled estrogen receptor-selective ligands modulate endometrial tumor growth. *Obstet Gynecol Int* (2013) **2013**:472720. doi:10.1155/2013/472720
200. Sun J, Meyers MJ, Fink BE, Rajendran R, Katzenellenbogen JA, Katzenellenbogen BS. Novel ligands that function as selective estrogens or antiestrogens for estrogen receptor-alpha or estrogen receptor-beta. *Endocrinology* (1999) **140**:800–4. doi:10.1210/endo.140.2.6480
201. Meyers MJ, Sun J, Carlson KE, Marriner GA, Katzenellenbogen BS, Katzenellenbogen JA. Estrogen receptor-beta potency-selective ligands: structure-activity relationship studies of diarylpropionitriles and their acetylene and polar analogues. *J Med Chem* (2001) **44**:4230–51. doi:10.1021/jm010254a
202. Filardo EJ, Quinn JA, Frackelton AR Jr, Bland KI. Estrogen action via the G protein-coupled receptor, GPR30: stimulation of adenylyl cyclase and cAMP-mediated attenuation of the epidermal growth factor receptor-to-MAPK signaling axis. *Mol Endocrinol* (2002) **16**:70–84. doi:10.1210/mend.16.1.0758
203. Dennis MK, Field AS, Burai R, Ramesh C, Petrie WK, Bologna CG, et al. Identification of a GPER/GPR30 antagonist with improved estrogen receptor counter-selectivity. *J Steroid Biochem Mol Biol* (2011) **127**:358–66. doi:10.1016/j.jsbmb.2011.07.002
204. Lan NC, Graham B, Bartter FC, Baxter JD. Binding of steroids to mineralocorticoid receptors: implications for in vivo occupancy by glucocorticoids. *J Clin Endocrinol Metab* (1982) **54**:332–42. doi:10.1210/jcem-54-2-332

205. Pecci A, Alvarez LD, Veleiro AS, Ceballos NR, Lantos CP, Burton G. New lead compounds in the search for pure antigluocorticoids and the dissociation of antigluocorticoid effects. *J Steroid Biochem Mol Biol* (2009) **113**:155–62. doi:10.1016/j.jsbmb.2008.12.018

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